THE PLANT CELL

Burning questions for a warming and changing world: 15 unknowns in plant abiotic stress

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Abstract

We present unresolved questions in plant abiotic stress biology as posed by 15 research groups with expertise spanning eco-physiology to cell and molecular biology. Common themes of these questions include the need to better understand how plants detect water availability, temperature, salinity, and rising carbon dioxide (CO_2) levels; how environmental signals interface with endogenous signaling and development (e.g. circadian clock and flowering time); and how this

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integrated signaling controls downstream responses (e.g. stomatal regulation, proline metabolism, and growth versus defense balance). The plasma membrane comes up frequently as a site of key signaling and transport events (e.g. mechano-sensing and lipid-derived signaling, aquaporins). Adaptation to water extremes and rising CO_2 affects hydraulic architecture and transpiration, as well as root and shoot growth and morphology, in ways not fully understood. Environmental adaptation involves tradeoffs that limit ecological distribution and crop resilience in the face of changing and increasingly unpredictable environments. Exploration of plant diversity within and among species can help us know which of these tradeoffs represent fundamental limits and which ones can be circumvented by bringing new trait combinations together. Better defining what constitutes beneficial stress resistance in different contexts and making connections between genes and phenotypes, and between laboratory and field observations, are overarching challenges.

Introduction

(By Paul E. Verslues, editor)

"Before now, you just needed to know the answers to the questions you were given; now you need to know questions which you have not been given and for which there is no answer." This is the advice that many mentors have given, in one form or another, to those new to research. Here, we put ourselves to this test and present what several groups of scientists working in plant abiotic stress biology consider to be big questions for future research. This is a timely topic as climate change will bring about not just a general warming but also instability and extreme weather events of many types. Thus, climate change can increase the frequency and severity of single, combined, and even sequential abiotic stresses including drought, salinity, flooding, and even freezing. Rising carbon dioxide (CO_2) levels can also directly influence how plants respond to these stresses. A common theme of plant stress research is that we are trying to understand how plants respond to excesses: too hot, too cold, too little water, too much water, too much light, too little light, or too much salt. These excesses are gradually (or not so gradually in some cases) becoming more of the norm for plants in many parts of the world. In this article, we focus on questions of fundamental plant biology and how stress affects physiological and molecular processes and adaptation, rather than climate change mitigation strategies discussed by our colleagues. Ultimately, we want to use our physiological and molecular knowledge to both predict the effect of climate change on plants and intervene to improve those outcomes, particularly in terms of ecosystem resiliency or crop yield. Thus, perhaps one overriding challenge is the question of scale and how to move our knowledge from one scale to another. How does the opening and closing of a membrane channel that occurs at a time scale of seconds (or less) influence growth responses that occur over days? How do those growth responses affect yield or reproductive fitness which is the culmination of months, or more, of the plant life cycle? How does knowing where a gene is expressed at the cellular scale, or where a protein is localized at the subcellular scale, help us understand coordination of root and shoot responses at the whole plant scale?

Another common theme that emerges from our big questions is the challenge for measurements of stress phenotypes to keep pace with, and make best use of, the ever-increasing amount of genomic data. How can this growing body of genomics data help us to understand gene and protein function and, ultimately, deploy that knowledge for plant improvement or understanding natural systems? Despite advances such as automated plant phenotyping and image analysis systems, this "phenotype gap" (Miflin, 2000) continues to grow larger as -omics data accumulate. In the process of closing the phenotype gap, one needs to decide what phenotype(s) to measure and how to interpret the results. Readers of this article may also get a sense that there are several alternative meanings of "stress resistance" (this is perhaps most pronounced for drought where the term "drought tolerance" is often broadly used for both avoidance of water depletion and true tolerance of low water potentials). Does increased resistance (often referred to as increased tolerance, regardless of whether avoidance mechanisms are involved) mean the ability to better survive a near-lethal stress or the ability to remain more productive during a moderate severity stress? Those approaching plant abiotic stress from an agronomic versus ecophysiology perspective can have differing views of this question. Several types of data indicate that the mechanisms plants use to survive severe stress only partially overlap with mechanisms enabling greater productivity at more moderate stress severities. Thus, there is a need to clearly state and define which view of stress resistance/stress tolerance one is applying when interpreting data. In addition, the challenges of connecting phenotypes observed in the laboratory to real differences in a field environment (and vice versa the challenge of achieving a mechanistic understanding of quantitative traits related to yield and stress resistance) demand a certain level of circumspection from those working at all levels of plant stress biology.

For cellular studies of abiotic stresses such as drought and temperature stress, we have a particularly challenging question of how plants perceive the stress at the molecular scale. Such abiotic stress perception does not follow the familiar receptor–ligand paradigm many of us learned in biochemistry class (back when it was enough just to know the answers to the instructor's questions). Without knowing the beginning, how the plant perceives a change in its environment, it is much harder to understand the downstream

responses at any scale. One emerging area of interest is the plasma membrane, and its interfaces with the cell wall and cytoskeleton, as a logical place for plants to sense environmental signals and control water and solute transport while also initiating down-stream signaling and induction of signaling intermediates such as abscisic acid (ABA). Yet we know relatively little about how the cell wall–plasma membrane–cytoskeleton interface acts in stress sensing and signaling and what the key molecular players in stress sensing are.

If we scale up to the whole plant level, there are long-standing questions about how plants control the movement of water through the soil–plant–atmosphere continuum and how stress responses may be coordinated across different tissues. For drought stress, it is sometimes assumed that the initial sensing events occur in roots because they are directly exposed to drying soil. But this need not be the case as the whole plant is hydraulically connected and changes in water potential at the root will be quickly propagated through the plant. Conversely, one could make an equally logical hypothesis that water limitation is first sensed in leaves as these are the site of water loss to the atmosphere and the site where stomata must quickly respond to restrict water loss when water supply from the roots and vascular system is disrupted. There is also a question of whether there are nonhydraulic, chemical signals that move from root to shoot, presumably in the xylem, to communicate a change in water status. The type of signal that this could be remains uncertain, although peptide signaling has recently received increased attention (Takahashi et al., 2018; Reichardt et al., 2020). These hypotheses of root versus shoot sensing and nonhydraulic signaling versus hydraulic signaling are not mutually exclusive and it seems likely that something as important for the plant as sensing changes in water availability (or changes in salinity, temperature, or CO_2 levels) is likely to have multiple mechanisms which operate in both distinct and overlapping ways. Under severe water limitation, disruption of water transport via xylem embolism becomes more likely and there is ongoing debate on how and when (or whether) refilling of xylem and recovery of hydraulic conductance can occur. There is also debate on how much loss of vascular function and hydraulic conductivity is lethal, either to specific tissue which becomes cut off from water supply or to the plant as a whole.

The concept of tradeoffs, and how they are regulated, is also a recurring theme of plant stress research. The concept of a "growth-versus-defense" trade-off is now frequently mentioned in molecular studies (including sometimes in studies that have data for only one side of the proposed tradeoff). While the "defense" side of the tradeoff may refer to pathogen defenses, which often cause obvious disruption of plant growth, this has gradually broadened to include many types of stress responses that may, either directly or indirectly, affect growth. Another trade-off example is the concept that plants can take a "water spender" strategy of maximizing carbon acquisition even at the cost of high water use versus a "water saver" strategy of restricting water use and acquiring less carbon but maximizing water use efficiency (WUE; amount of carbon acquired per unit of water lost via transpiration). Which strategy is better for a species depends not only on the environment but also on which other plants share that environment. Saving water in the soil for later is less effective if your neighbor spends it first. An important question for research is whether these trade-offs, or other ecologically important tradeoffs, can be broken (high WUE and rapid growth, for example).

In the sections below, we consider these and related questions with the hope that other researchers will be informed and motivated to add, and answer, many other questions about plant abiotic stress that we do not yet know to exist.

Can plasticity in traits beneficial in both wet and dry soils be recognized and used to limit crop yield loss?

(By Julia Bailey-Serres)

Increased climate variability is responsible for excessive wet and dry soil conditions that affect irrigated and rain-fed agriculture. From this, there arises a question of whether there are genes and traits, or sets of genes and traits, associated with greater plant resilience in both of these extreme soil environments. One can find support for this notion in species that thrive in ephemeral wetlands. These possess constitutive adaptive traits or display plastic acclimation strategies that facilitate survival in areas that undergo a seasonal rise and ebb of the water table that inundates root systems and can partially or completely submerge aerial tissues. More often than not, wet and dry cycles occur in succession, necessitating traits that are plastic or beneficial under both extremes. Few crops withstand water-saturated soil (waterlogging), let alone submergence for more than several days. Rice is an exception, surviving by accelerating or dampening underwater growth. Flooding escape of seedlings is aided by ANAEROBIC GERMINATION 1, encoding a trehalose 6-phosphate phosphatase, that increases sink strength of the snorkel-like coleoptile, allowing the germinating seedling access to air (Kretzschmar et al., 2015). Deepwater rice can outgrow a seasonal rise in paddy depth of over 3 m. Within submerged stems, ethylene activates a gene suite (SNORKEL1/2, SEMIDWARF1, and ACCELERATOR1) that amplifies cell division at stem node meristems and subsequent internode elongation (Hattori et al., 2009; Kuroha et al., 2018; Nagai et al., 2020). In contrast to this adaptive strategy, the submergence tolerance regulator SUBMERGENCE1A (SUB1A), encoding an ethylene-responsive transcriptional regulator factor subfamily VII (ERF-VII), limits the exhaustion of leaf carbohydrate in leaf elongation (Fukao et al., 2006) and minimizes postsubmergence ROS and water deficit (Fukao et al., 2011). This transient tolerance protects semi-dwarf paddy rice from deep but short-term flash floods. While these studies focus on rice, there are species in all major crop families that are adapted to transient wet zones (*Oryza* and *Zea* in the Poaceae, *Lotus* in the Fabaceae, *Solanum dulcamara* in the Solanaceae, and

Rorippa in the Brassicaceae). These, along with rice and flooding tolerant Arabidopsis (*Arabidopsis thaliana*), provide insight into plastic survival strategies lost during crop domestication or selection for production agriculture.

Roots perceive subtle changes in soil moisture including flooding, which restricts diffusion of gases, elevating ethylene and depleting O_2. Can the discovery of regulatory mechanisms accelerate improvement of waterlogging resilience in crops without a yield penalty? Might this be accomplished even if flooding is followed by water deficit?

Let us consider root system traits that are associated with survival of waterlogged and anaerobic soils. When roots of diverse crops (e.g. rice, tomato, and Medicago) become O_2-deprived, a conserved low-O_2 gene regulatory network (GRN) is activated by SUB1A-like ERFs that are stabilized as O_2 levels fall (van Dongen and Licausi, 2015; Reynoso et al., 2019). The genes with conserved ERF-VII cis-regulation encode enzymes of anaerobic metabolism, turnover of ERF-VIIs upon reoxygenation, and ABA perception. Without aeration, as in root meristems of fully submerged rice, DNA synthesis and the cell cycle are attenuated until shoots are re-aerated (Reynoso et al., 2022).

Aeration in flooded soils is enhanced by conditional proliferation of roots with a shallow angle. In rice, this is augmented by loss-of-function of *SOIL SURFACE ROOTING1*, a homolog of *DEEPER ROOTING1* (*DRO1*) (Kitomi et al., 2020), limiting auxin-mediated gravitropism. Also important are shoot-borne (adventitious) roots that emerge near the air–water interface, capturing dissolved O_2 and nutrients of floodwaters (Lin et al., 2021). Their emergence is auxin-mediated, triggered by ethylene, and localized production of reactive oxygen species (ROS). Aeration is enhanced by internal or external passageways called aerenchyma that provide a low resistance path for the diffusion of O_2 and other gasses between aerated shoots and waterlogged roots (Pedersen et al., 2021). Aerenchyma form within the cortex of rice and maize roots through ethylene- and ROS-triggered programmed cell death (He et al., 1996; Yamauchi et al., 2016, 2017); auxin signaling is also implicated in rice (Yamauchi et al., 2019). Another characteristic of waterlogged roots is the accumulation of suberin lamellae, a layered polyester of poly(phenolic) and poly(aliphatic) fatty acids, in the apoplasm of the exterior side of the outermost cortical layer (exodermis) or periderm (epidermis of older roots and stems) (Pedersen et al., 2021). Accumulation of the suberin lamellae is mediated by ABA in rice (Shiono et al., 2022). The extension of this gas and water impermeable barrier toward the root tip limits the outward diffusion of O_2 en route to root meristems.

Aeration traits can be constitutive, as observed in paddy weeds (i.e. *Echinochloa* species; Ejiri and Shiono, 2019) and Amazonian *Oryzae* (Ejiri et al., 2020), or induced by waterlogging or prolonged (stagnant) flooding. Both cortical aerenchyma and exodermal suberin are constitutive in the wetland teosinte *Zea nicaraguensis*, but induced by ethylene

in domesticated maize cultivars (Abiko et al., 2012). The mapping and marker-assisted breeding of genetic determinants of these plastic traits are important for crop improvement. Promising examples include a locus associated with adventitious rooting upon waterlogging in soybean that enhances yield stability (Ye et al., 2018) and loci determining aeration traits in teosinte that have been pyramided into maize cultivars (Mano and Omori, 2013; Mano and Nakazono, 2021).

Can root aeration traits be beneficial in dry soils? Water deficit also activates the formation of aerenchyma, a strategy that purportedly reduces metabolic costs of deep water-seeking roots (Lynch, 2015). It also reinforces the exodermal suberin barrier to limit water loss by diffusion between the root tip zone and distal regions. In contrast to the shallow-angle roots of wetland rice, constitutive deep rooting is characteristic of drought-resilient crops. In upland rice, a functional *DRO1* allele promotes auxin-mediated gravitropism resulting in a deep root angle (Uga et al., 2013). Yet plants with shallow root systems that allow greater access to phosphate, nitrogen and other nutrients can display hydrotropism (growth toward moisture through gradient sensing) and hydropatterning (Bao et al., 2014; Robbins and Dinneny, 2018; Lind et al., 2021), discerned as auxin-mediated but ABA-limited lateral branching on the moist side of a root in Arabidopsis (Orosa-Puente et al., 2018). These traits, along with the prioritization of elongation of established crown (nodal) or other roots with access to moisture, may provide sufficient plasticity for survival of a wet to dry transition (Figure 1). But a challenge may be whether spatial or temporal perception of ABA promotes or inhibits root growth in the specific environmental context and species. Another consideration is the consequence of waterlogging on beneficial plant–microbe interactions. Rhizobial nitrogen fixation in legumes and mycorrhizal fungal interactions are limited by waterlogging, yet both recover as soils dry (Justino and Sodek, 2013; Groen et al., 2022).

Comparative genomics, systems biology at the tissue and cell level, and genome editing have expanded opportunities to address these questions to overcome the urgent challenge to increase water extreme resilience in crops.

How do plants sense and communicate water deficit?

(By Alexander Christmann and Erwin Grill)

Homeostasis of water status is a major challenge for plants. The gradient of water potential (ψ_w) from the soil to the plant and subsequently to the atmosphere generates a hydraulic continuum that efficiently mobilizes soil-borne water via stomata transpiration into the air. Water availability and the water-conducting capacity of plants impose constraints on this water flux. Regulatory processes governing long-term adjustments and stomata responses to changes in water status are well understood on a molecular level and involve the phytohormone ABA (Yoshida et al., 2019a, 2019b; Zhang
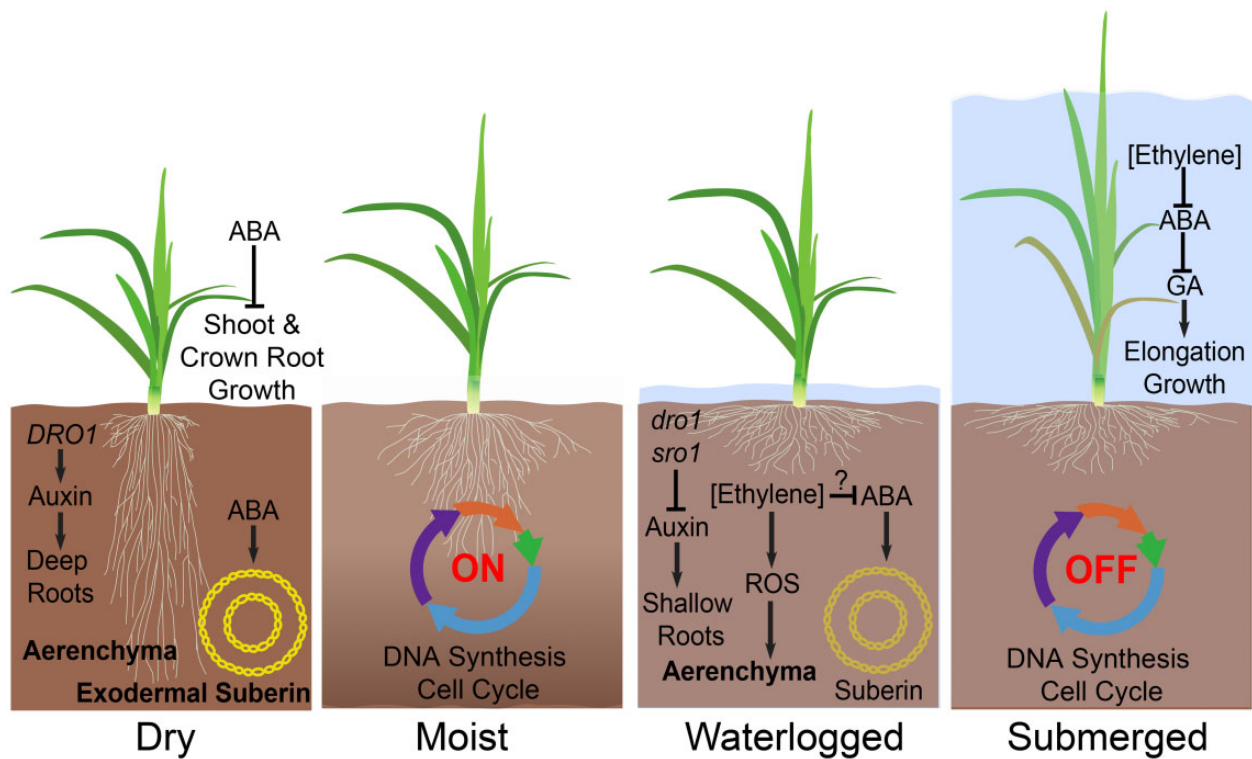
Figure 1 Characteristics of shoot and root growth plasticity associated with variation in soil moisture using semi-dwarf rice as the model. Roots in dry soils grow toward water reserves and limit water loss by deposition of a pronounced suberin barrier, particularly surrounding the exterior side of the exodermis. In moist soils, root architecture is more compact, crown root initiation and elongation are active, and exodermal suberization is limited. Waterlogged roots containing aerenchyma proliferate near the soil surface and exodermal suberin prevents oxygen loss by radial diffusion. In fully submerged plants, cell division activity in root systems is rapidly switched off and leaf elongation is enhanced, promoting escape to the air. Key phytohormones, their relationships and genes mentioned in the main text are shown.

et al., 2022). However, the molecular components of sensing water deficit and communicating these cues within the plant remain largely speculative.

Effects of water deficit

Uptake of soil water by plants requires a root ψ_w more negative than the surrounding soil, which is achieved predominantly by osmotic adjustment and negative hydraulic pressure due to transpiration. A negative water balance (i.e. water uptake lower than its release by transpiration) immediately changes a number of parameters within the plant. In the substomatal cavity, the major evaporation site, ψ_w of the apoplastic fluid becomes more negative, causing neighboring cells to lose water, and consequently causing their turgor and volume to decrease (Figure 2). Simultaneously, the hydraulic tension increases within the xylem. These water deficit-induced changes are relayed within the tissue and, depending on the extent of water loss and water capacitance of the plant tissue, possibly to other organs. Changes in hydraulic parameters serve as a fast long-distance signal ($>40\,\mathrm{cm}\ \mathrm{min}^{-1}$) (Christmann et al., 2013). Suppression of the hydraulic signal from desiccating roots blocked the ABA-induced leaf response, that is, stomatal closure and ABA-dependent gene expression (Christmann et al., 2007).

Reduction of stomatal aperture by ABA signaling readjusts the water balance by lowering transpiration. Stomatal responses to high water vapour deficit are regulated in a largely guard cell-autonomous manner but with pavement cell–guard cell interaction. Signaling elements of this response include several protein kinases, Raf-like proteins, and the receptor-like kinase GHR1, which act upstream of the ABA response mediator Open Stomata 1 (OST1) (Hsu et al., 2021). Improved water uptake via increases in root hydraulic conductance (Maurel et al., 2016) and osmotic adjustment recover leaf gas exchange at the expense of increasingly negative plant ψ_w. These adjustments are incremental and occur even under nonstress water conditions.

How is the change in ψ_w in the plant perceived?

Changes of tissue ψ_w relative to its surroundings affect cellular water flux, osmolarity, and mechanical forces including turgor acting on the cell wall, biomembranes (e.g. plasma membrane, tonoplast, and endomembranes), and the cytoskeleton. There is mounting evidence for a mechano-sensitive mechanism that detects these changes. Classical paradigms for mechano-sensitive and ψ_w change-mediated plant responses are touch-induced leaf closure of the Venus flytrap and tendril coiling of Bryonia that are very sensitive
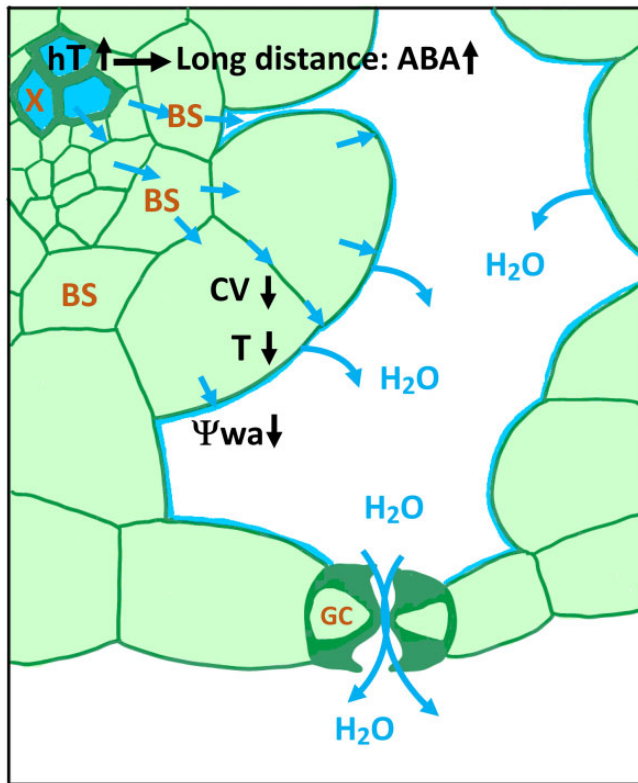
Figure 2 Water flux from the xylem vessel to the evaporation site in a schematic leaf cross section. Water transported through the xylem (X) of a small leaf vein passes through bundle sheath (BS) cells to mesophyll and epidermal cells, which surround guard cells (GCs), the sites of transpiration. GCs are symplastically isolated (have no plasmodesmata) and water supply occurs from the apoplast. Transpiration exceeding water replenishing at the evaporation sites results in a reduced water potential of the apoplastic fluid (ψ_{wa}). This key event triggers a number of changes: a reduction of turgor (T) and cell volume within the mesophyll, and an increase in the hydraulic tension (hT) within the vasculature. The change in hT serves as fast long-distance signal and there might be parallels with mechanical sensing and relay of touch and wounding. The hydraulic signal is converted into the chemical signal ABA to adjust water conductance and stomatal aperture. Mechano-sensitive components are likely involved in all cell types but have not been unequivocally identified.

to external force, at least as sensitive as human touch. (Klusener et al., 1995; Escalante-Perez et al., 2011) Thus, touch-sensitive signaling offers a good model to conceptualize the types of mechanisms that could be involved in low ψ_w perception and initial signaling.

The Venus flytrap uses multicellular trigger hairs to sense prey by converting mechanical forces exerted by the insect into a turgor-driven snap. The response to insect touch is relayed to neighboring cells by electrical signaling involving a calcium (Ca^{2+}) wave within the leaf lobe (Suda et al., 2020). The sensory cells of the trigger hair are enriched in ion channels associated with mechano-perception and Ca^{2+} entry (Iosip et al., 2020; Procko et al., 2021). These include homologs of the Arabidopsis mechano-sensitive-like channel 10 (MSL10) (Basu and Haswell, 2020), the glutamate

receptor Ca^{2+} channel GLR3.6 and hyperosmolality-induced [Ca^{2+}]-increase (OSCA)-family channels. Members of the OSCA family are mechanically activated and ion nonselective (Murthy et al., 2018). Touch-induced depolarization of the sensory cells is mediated by MSL10 and initiates action potentials that propagate via GLR3.6 and OSCAs. MSL10 and GLR3.6 are known components of electrical signal propagation upon wounding (Toyota et al., 2018; Farmer et al., 2020; Moe-Lange et al., 2021). In the case of wounding, hydraulic pressure waves are initiated that propagate through the xylem and trigger an electrical and Ca^{2+} wave in the vasculature. The waves are relayed (10 cm min^{-1}; Farmer et al., 2020) by MSL10 and may require downstream-acting GLRs localized to endomembranes, namely the phloem-expressed GLR3.3 and GLR3.6, which are also highly expressed in xylem-contacting cells (Nguyen et al., 2018; Moe-Lange et al., 2021). Hence, mechano-sensing in the Venus flytrap and wounding response utilize many of the same molecular components and also share a signal propagation mechanism that includes a wave of increased intracellular Ca^{2+}. Endomembrane compartments also play a critical role in Ca^{2+} release during mechano-stimulated responses (Klusener et al., 1995). Mechanical forces acting from outside the cell can be relayed to these intracellular cell compartments by the cytoskeleton and cytoskeleton-associated proteins (Bhaskara et al., 2017; Hamant et al., 2019) or by the loss of water causing osmotic disequilibrium between intracellular compartments which may activate stretch related sensing on intracellular membranes in addition to the plasma membrane. The involvement of endomembrane signaling in stress response is also indicated by observations that loss of the chloroplast-localized mechano-sensitive channels MSL2 and MSL3 leads to osmotic imbalance between chloroplast and cytosol and constitutive activation of low ψ_w response in unstressed plants (Wilson et al., 2014).

The electrical wave-induced cell depolarization involves additional components shared with ABA responses such as voltage-dependent anion and cation channels, NADPH-oxidase, and the proton ATPase driving re-polarization (Farmer et al., 2020; Iosip et al., 2020). While wounding results in a sudden relaxation of the hydraulic tension at the severed xylem, water deficit increases this tension. The increased xylem tension translates into a stronger pulling force acting on the xylem-contacting cells. A sudden change of this force might distort and stretch domains at the plasma membrane. Pulling forces of −0.1 bar resulted in half-maximum MSL10 activation in the Venus flytrap (Procko et al., 2021) and even relatively mild water deficit could be expected to generate similar or stronger forces. Touch- and wound-activated responses also induce chemical signals including oxo-phytodienoic acid and jasmonate for subsequent phytohormone signaling (Escalante-Perez et al., 2011; Farmer et al., 2020). We currently do not know the extent to which touch- and wound-induced signaling mechanisms overlap with the mechanisms used to detect and respond

to water-deficit. If such mechanisms are involved in water deficit signaling they would be expected to induce ABA accumulation as a key signaling factor to turn on further downstream stress responses.

Genetic screens for Arabidopsis mutants impaired in regulating ABA-responsive reporter expression under hyperosmotic stress have failed so far in identifying water-deficit sensing receptors (Wang et al., 2011; personal experience). Genetic and functional redundancy provides an explanation, in which receptors act in parallel pathways that converge on ABA. This might be the case considering the multiple cellular effects of leaf transpiration exceeding water uptake. Use of Arabidopsis natural variation in low ψ_w-induced ABA accumulation identified candidate loci affecting ABA accumulation (Kalladan et al., 2017); however, the possible role of these candidate loci in stress-signaling needs to be validated and further studied. The increased tension of xylem water may activate MSL-type channels of contact cells or mechanical force-sensing Ca^{2+} channels as part of long-distance communication. These contact cells and the surrounding parenchyma of the vasculature play a prominent role as specific sites of ABA biosynthesis (Endo et al., 2008). Cells lining the substomatal evaporation sites are particularly challenged with water efflux and turgor decrease. A decrease in cell volume and the concomitant reduction in plasma membrane surface (Sack et al., 2018) could be sensed similar to yeast target of rapamycin (TOR) complex2 that balances plasma membrane constraints with membrane lipid level (Riggi et al., 2019). Turgor-sensing and—controlling mechanisms need to be activated to re-establish water balance. However, bona fide turgor sensors of plants are not known yet. In yeast, the histidine-kinase SLN1 senses turgor and controls the hyperosmolarity response (Reiser et al., 2003). Interestingly, several structurally related histidine kinases of Arabidopsis, including AHK1 and the cytokinin receptors AHK2, AHK3, and AHK4, can complement the turgor-sensing function of SLN1 either alone or in the presence of cytokinin for AHK4 (Reiser et al., 2003; Tran et al., 2007). However, it is unclear whether AHKs act as water stress sensors in plants (Kumar et al., 2013). RAF-like protein kinases involved in osmotic adjustments are promising candidates for downstream-acting signaling components (Lin et al., 2020; Soma et al., 2020).

Components that maintain or monitor cell wall integrity are critical in stressful conditions exerted by high turgor pressures. Such components include pectate lyase (Chen et al., 2021) and several plasma membrane-localized protein kinases like FERONIA, required to avoid root cell burst in response to salt stress (Feng et al., 2018), CrRLK1L/BUPS1 (Zhou et al., 2021), THESEUS (Bacete et al., 2022), and STRUBBELIG (Chaudhary et al., 2021).

In summary, water deficit responses are activated during increasing transpirational demand to restore plant water homeostasis and sustain photosynthesis. Understanding how plants sense and communicate water deficit on a molecular level provides a promising tool to increase WUE without yield penalty needed for crops of the future (Yang et al., 2019).

How, where, and when are water deficit signals integrated during floral transition?

(By Lucio Conti)

Plant physiology textbooks emphasize how plants are continuously challenged by their surrounding environment and how this triggers developmental adjustments. But what happens if the environment deteriorates to the point of threatening survival? Stressors like water deficit can pose such a challenge for plants and one survival strategy relies on the high flexibility of the flowering program. The switch to flowering is a crucial decision for plants, determining the cessation of the vegetative phase and initiation of reproductive development. It occurs at the shoot apex upon receipt of environmental and endogenous signals and precedes shoot elongation, specification, and outgrowth of floral organs. Depending on the ecological context appropriate timing of the floral transition positively influences inflorescence growth, architecture, and the number of flowers produced (as these processes rely on carbon assimilation from vegetative leaves). Several studies describe the rapid natural and artificial selection for early flowering phenotypes to evade harsh summer drought scenarios (Franks, 2011; Kenney et al., 2014; Monroe et al., 2018; Groen et al., 2020), a strategy referred to as drought escape (DE). DE permits the completion of the life cycle before water deficit conditions become extreme, even if at the expense of fitness. Flowering time regulation is also extremely plastic in the face of unpredictable environmental constraints (Blackman, 2017). Similarly, DE can also be adaptive (Figure 3A) which means that upon experiencing water deficit, some species can activate an earlier floral transition, although significant variations in genotype by water deficit interaction exist (Franks, 2011; Kenney et al., 2014).

Where does DE originate? Flowering is regulated through a network of genes that perceive environmental/endogenous signals and genes that integrate these signals to regulate floral induction. Florigen genes are important integrators expressed in the vascular tissue of vegetative leaves. They encode a class of small globular proteins that move long-distance through phloem vessels at the shoot apex to cause transcriptional reprogramming of meristem cells and floral specification of new primordia. While florigen's transcriptional activation usually occurs upon perception of critical daylengths, studies in Arabidopsis, rice, and tomato further point to the contribution of water deficit signals (Riboni et al., 2013; Du et al., 2018; Chong et al., 2022). Transcriptional upregulation of florigen genes in response to water deficit appears to be necessary and sufficient to cause DE in these species, despite their evolutionary separation. The phytohormone ABA emerges as a conserved molecule regulating florigen expression, although different mechanisms are involved (Figure 3B). In tomato, ABA stimulates phosphorylation and subsequent nuclear shuttling of the
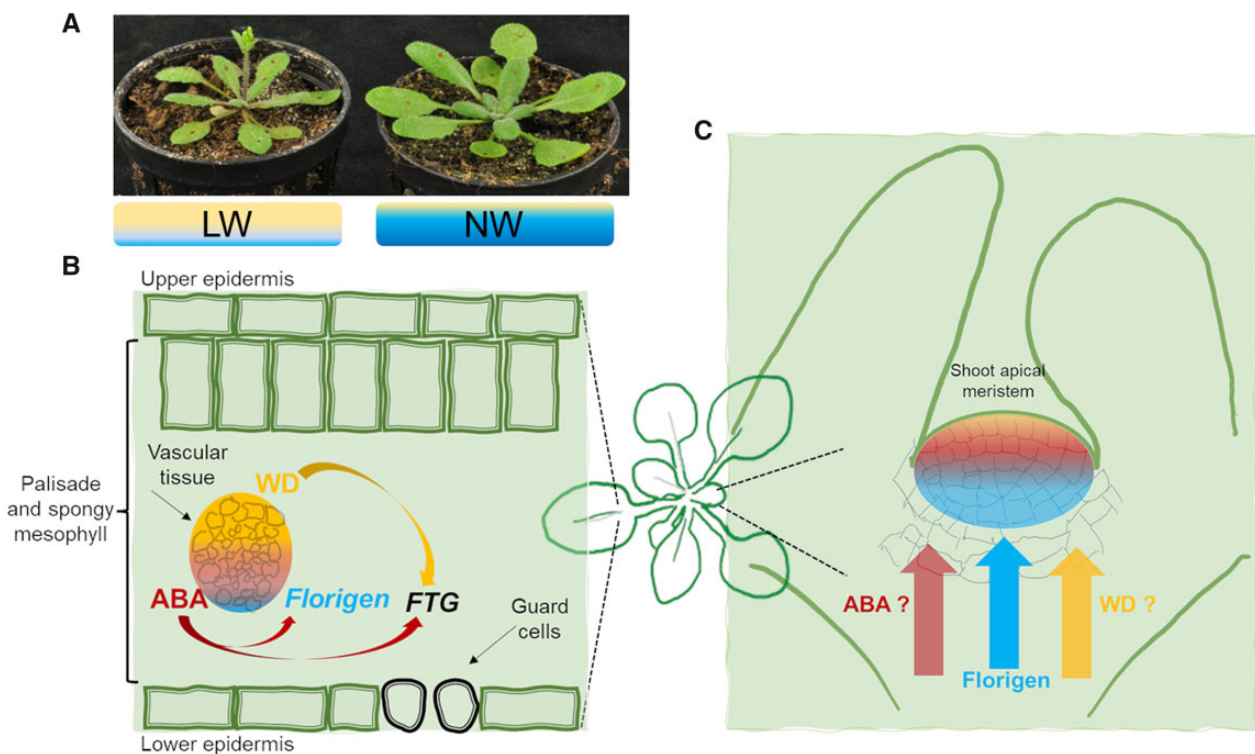
Figure 3 Known and uncharacterized signals causing DE and their sites of action. A, wild-type Arabidopsis plant photographed at ~3 weeks from germination. Plants were grown under normal or low-watering (NW and LW, respectively) regimes. LW plants exhibit DE. B, Cartoon outlining a leaf cross-section. Water deficit signals (WD) are transmitted through the vasculature to stimulate ABA production. ABA positively contributes to florigen transcriptional activation in leaves in response to water deficit. ABA and WD promote expression of many other flowering time genes regulating guard cell activity and drought tolerance traits. C, Besides florigen proteins, different signals converge at the shoot apical meristem to relay water deficit information and regulate floral transition accordingly. Despite the ability of the shoot meristem cells to integrate these cues, it is still unclear how water deficit signals (WD) can reach the apex and if significant ABA translocation occurs.

transcription factor VASCULAR PLANT ONE-ZINC FINGER 1 to activate the florigen SINGLE FLOWER TRUSS (Chong et al., 2022). In Arabidopsis and rice, ABA orchestrates different transcriptional and posttranscriptional mechanisms, with key contribution of a class of ABA-regulated basic leucine-zipper (bZIP) transcription factors that, in Arabidopsis, are incorporated into multimeric protein complexes to activate (indirectly) florigen expression (Yoshida et al., 2014; Hwang et al., 2019).

Are florigens unique in relaying water deficit information at the shoot apex? Considering that ABA is an important driver of DE, its redistribution at the shoot could also influence the flowering process (Figure 3C). Phosphoproteomics studies in Arabidopsis reveal several flowering time regulators as substrates of ABA-activated signaling, suggesting multiple and spatially separate points of control of the floral network (Wang et al., 2013a). Is there ABA translocation at the shoot apex or are shoot meristem cells capable of de novo ABA production in response to water deficit signals? Can ABA directly affect cell fates at this site? These questions remain largely unsolved. In Arabidopsis, ABA biosynthesis occurs in roots and vascular bundles of leaves, largely overlapping with the main sites of florigen expression (Kuromori et al., 2014). ABA is loaded into the phloem to be distributed across different tissues (Kuromori et al., 2018; Daszkowska-Golec, 2022). While measuring ABA cellular concentrations in specific shoot cells remains challenging, there is evidence that florigen proteins can directly control the expression of different ABA signaling genes at the shoot apex (Zhu et al., 2020). ABA and its signaling cascade regulate leaf emergence rates, suggesting a direct influence on meristem cell activity, possibly mediated by regulation of primary metabolism (Yoshida et al., 2019b). These findings could set the stage for a better understanding of ABA-regulated cell fate reprogramming in response to, or in parallel with, florigen mobilization at the shoot in response to water deficit. Additionally, this would stimulate further questions about the role of ABA accumulation and signaling in flowering time regulation under optimal irrigation conditions and its conservation across species. For example, reducing ABA sensitivity of rice plants by mutations in a clade of ABA receptors causes late flowering, a phenotype that is not observed in Arabidopsis (Miao et al., 2018).

Many questions remain concerning the number of signals elicited in response to water deficit, their integration in reproductive development, and the role of flowering time genes in conferring drought protection (Figure 3, B and C). Water deficit-stimulated ABA production inhibits shoot

growth, thereby delaying the appearance of floral structures (LeNoble et al., 2004). Thus, water deficit may lead to uncoupling chronological time to flower (the appearance of floral organs) from floral transition per se. Furthermore, unknown signals can influence shoot meristem function, flowering time, and florigen expression, depending on the level of water deficit imposed (Galbiati et al., 2016; Du et al., 2018). How cells can distinguish between varying levels of water deficit is unclear. ABA also activates negative regulators of flowering, including FLOWERING LOCUS C (FLC) in Arabidopsis (Wang et al., 2013b). FLC contributes to delaying flowering, which is most apparent when water deficit occurs under noninductive daylength conditions (i.e. when florigen expression is repressed) (Riboni et al., 2013). Because FLC antagonizes the expression of multiple floral genes in leaves and at the shoot apical meristem, its activation may be required to curb excess florigen signaling and finetune inflorescence development according to stress intensity. Concurrent regulation of antagonistic flowering mechanisms may also generally impact drought tolerance traits. For example, plants carrying functional alleles of FLC and its upstream regulator FRIGIDA (FRI), display strong reductions in water use (Mckay et al., 2003). SHORT VEGETATIVE PHASE, a floral repressor related to FLC, promotes ABA accumulation (Wang et al., 2018b). In contrast, the florigen gene FLOWERING LOCUS T (FT) regulates stomatal opening, favoring transpiration (Kinoshita et al., 2011). Considering the importance of the duration of crop cycles on yield and the role of ABA in reducing water loss, identification of DE molecular mechanisms and their natural genetic variations could offer targeted strategies to balance flowering time and drought tolerance traits.

How do we incorporate plant diversity into our molecular understanding of environmental stress adaptation?

(By José R. Dinneny)

It is an obvious fact, but worth reiterating, that plants have evolved to occupy nearly every environment on the earth's surface (Corlett, 2016). Furthermore, through agriculture, humans have bred plants that are now cultivated across 12.6% of the total terrestrial landscape (Global Cropland Extent Product at 30 m (GCEP30) (Thenkabail et al., 2021). The ability of plants to occupy this breadth of environments involved the evolution or breeding of plant physiological mechanisms to meet the diverse environmental challenges that are faced in each ecosystem and agricultural management system. Despite this clear abundance of physiological diversity, the majority of mechanistic research in plants is still focused on a small collection of stress-sensitive model systems. This is not to say that such discoveries are unimportant or limited in impact, however, we have a patchy understanding as to whether such studies will identify broadly relevant principles, or rather species-specific details. Addressing the question of how plant–environmental responses are diversified across the kingdom will provide

insight into the major innovations plants have evolved to survive in different environments and will also inform strategies for introducing such mechanisms into a broader range of crop plants.

Much of the past emphasis on investing in molecular genetic model systems such as Arabidopsis was based on the historic limitation in the availability of genomic resources. Furthermore, these concerted efforts established a critical mass of researchers focused on determining a baseline understanding of plant molecular biology (Provart et al., 2016); however, this is no longer applicable. The Kew Royal Botanic Gardens recently estimated that ~374,000 plant species have been discovered (Christenhusz and Byng, 2016) and recent efforts have led to genome sequences being available for over 350 species. Botanic gardens have made efforts to sequence the genomes of their collections and the Ruili Botanic Gardens in China has done so for 689 species (Liu et al., 2019). The 10,000 Plant Genomes Project run by the Beijing Genome Institute promises to expand this list of available sequences far beyond what is currently available (https://db.cngb.org/datamart/plant/DATApla1/). Furthermore, transcriptomic profiling of species across the green plant lineage (Leebens-Mack et al., 2019), or occurring in specific ecological niches, such as the Atacama Desert (Eshel et al., 2021), has clarified phylogenetic relationships and illuminated physiological adaptations of plants to arid climates. In short, the book of life for plants is being revealed at an extraordinary pace.

Despite this rapid progress, the pace of studies that functionally explore this glut of genomic data has not kept pace. The bottlenecks that limit our ability to functionalize genome sequences and discover the molecular mechanisms governing adaptations to the environment are three-fold. The first limitation is our understanding of the diverse physiological adaptations that plants use to survive environmental challenges. The second is the paucity of methods available in nonmodel plants to enable the functional characterization of a plant's genomic sequence. Finally, limitations in the ability to manipulate the genomes of a diverse array of species through genetic engineering prevent hypothesis testing about genotype–phenotype relationships and the application of this knowledge.

The lack of understanding of the diversity of physiological adaptations to the environment is particularly apparent for the root system, which by its nature has remained hidden behind a veil of soil. Roots can be considered the sustainability organ system (Lynch, 2007). They function to provide the plant anchorage and prevent displacement from a fixed position in the ground (Hostetler et al., 2021b), but other processes are also relevant. Roots are the major conduit for the absorption of water and nutrients and roots engage in a metabolic bartering system with soil microbes, which facilitates nutrient uptake in exchange for the products of photosynthesis (Fitzpatrick et al., 2018). Plants also communicate with each other through their roots and this can affect the density and diversity of local communities

(Mommer et al., 2016). Despite these varied functions, very little molecular insight into how roots perform these functions has been described beyond in a few model species. Furthermore, of the tissues and cell types that are thought to compose most roots, little is understood as to whether plant species have evolved innovations in cellular function that allow them to survive in the vast array of soil types and terrains on earth. Innovations in the preparation of plant tissue for light-based microscopy, such as ClearSee (Ursache et al., 2018), and in the use of other imaging modalities, such as microscopic computed tomography (microCT) (Mairhofer et al., 2011; Morris et al., 2017), have opened up opportunities for the quantitative exploration of diverse plant anatomical structures and of root systems grown in soil.

Specific plant families such as the Brassicaceae (Koenig and Weigel, 2015), Solanaceae (Fernandez-Pozo et al., 2015), and Poaceae (Buell, 2009) have emerged as models for comparative genomic studies. Encompassing ∼3,630 species, the Brassicaceae family is home to plant species used in agriculture including oilseed crops *Brassica napus* (canola) and *Camelina sativa*, salt-tolerant halophytes species *Eutrema salsugineum* and *Schrenkiella parvula*, and the well-characterized model molecular-genetic plant Arabidopsis (Figure 4A). Arabidopsis provides a nested model system within the Brassicaceae family for exploring the diversification of stress responses. The sequencing of over 1,000 accessions facilitated the identification of genetic loci under selection, and identified the ABA signaling pathway as being important (1001 Genomes Consortium, 2016). ABA, which is induced under drought and salinity stress (Cutler et al., 2010), suppresses root growth in Arabidopsis, particularly at concentrations above 1 μM, while in other species like *S. parvula*, an extremophyte plant living at the edge of a hyper-saline lake, ABA accelerates growth (Sun et al., 2022). These data suggest that even for well-characterized signaling pathways, diametric changes in response to ABA are possible. Further exploration of the diversification of stress response pathways will help to reveal the principles behind the tuning of such pathways during evolution.

Methodological innovations are beginning to provide functional insight into nonmodel plants (Figure 4, B and C). Single-cell RNA sequencing (scRNA-seq) now allows cell type-specific transcriptional profiles to be obtained from a diverse range of species (Tarashansky et al., 2021). This provides insight into the diversification of cell type functions and the potential discovery of new cell types. DNA affinity purification and sequencing (DAP-seq), which allows for the in vitro reconstitution of transcription factor–genome interactions (O'Malley et al., 2016), enables the determination of GRNs in nonmodel species (Sun et al., 2022). Comparisons of GRN architecture between related species provide a means of understanding how the evolution of genomic sequence leads to the rewiring of GRNs and the regulation of downstream physiological processes important for stress acclimation. These advances, together with improvements in gene editing and plant transformation (Nadakuduti and Enciso-Rodríguez, 2020; Anjanappa and Gruissem, 2021) dramatically brighten the horizon for studies in nonmodel species.

How do plants integrate climate signals?

(By Scott Hayes and Christa Testerink)

Integrating stress signals: from field observations to understanding cellular events

Over the past two decades, several canonical environmental signaling pathways have been established. These pathways trace environmental signals from perception to transduction and finally an altered transcriptional and phenotypic response. These advances are a notable achievement. Studying "clean" responses to a single stress has proved a sensible
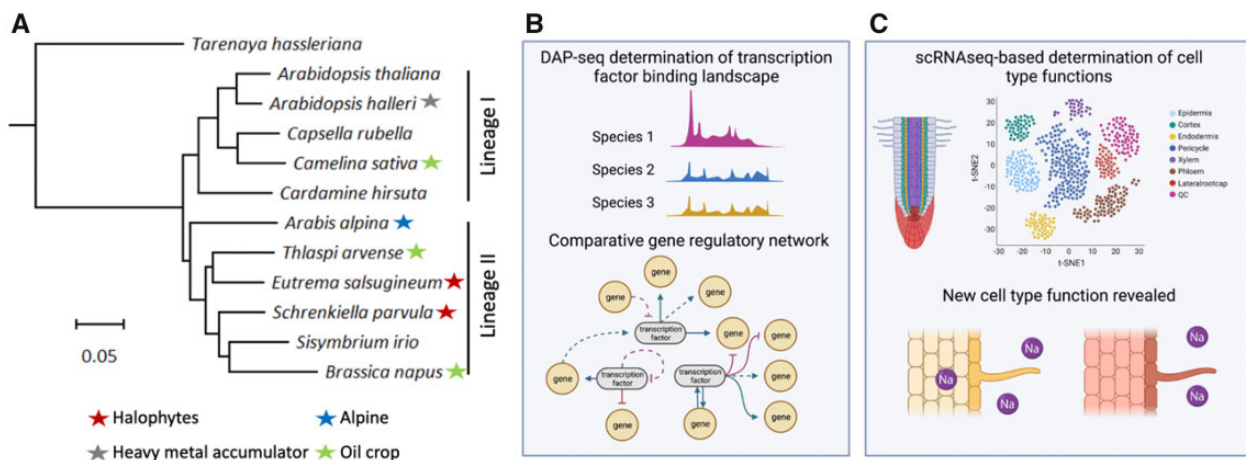

Figure 4 The Brassicaceae family contains species with diverse physiological traits and agronomic value. A, Phylogeny of species in the Brassicaceae family with sequenced genomes. Stars indicate different physiological or agronomic properties of interest. Figure courtesy of Dong-Ha Oh. B, DAP-seq facilitates the rapid determination of transcription factor–genome interaction landscapes. GRN architecture can be compared between species to identify rewiring events. C, scRNA-seq enables cell-type functions to be explored between species and for new functions to be uncovered.

and effective approach to identifying sensors and signaling pathways (Lamers et al., 2020). What is becoming increasingly clear, however, is that many of these signaling pathways are context-dependent. Environmental cues are often transduced through overlapping molecular components, leading to highly contextual molecular responses. While many agronomical and crop science studies have already extensively addressed naturally occurring combinations of environmental stress factors (Rivero et al., 2022), the molecular mechanisms underpinning these interactions are often obscure.

To understand plant growth in complex environments, we must improve our understanding of how different cues are integrated into plant development. In this section, we focus on the cellular pathways governing the integration of abiotic signals, with a focus on water availability and temperature as relevant climate change-related cues, that often coincide (Livneh and Hoerling, 2016). We highlight important factors that need to be taken into consideration when studying signal integration, and we put forward conceptual frameworks through which to study these processes.

Genome-wide studies identify interesting patterns

Phenotypic and transcriptomic studies have offered the first clues to the molecular mechanisms involved in signal integration. Pioneering studies found that the transcriptomic and metabolic response of plants treated with both drought and heat stress differed dramatically from when these stresses were applied in isolation (Rizhsky et al., 2004). Proline accumulated in response to drought, but not in response to a combination of drought and heat stress. Instead of proline, sucrose was produced as an osmoprotectant in these conditions (Rizhsky et al., 2004). More in-depth studies, using up to six combinations of five stresses (cold, high light, salinity, heat, or flagellin) found that around 60% of transcriptional responses could not be predicted from the response to single stresses alone (Rasmussen et al., 2013). Sewelam et al. (2014) tested the transcriptional response to salt, mannitol, heat stress, and combinations of the three and also found that transcripts in the combined treatment could not be accounted for by the data for single stresses alone (Sewelam et al., 2014). Most studies into transcriptional signal integration have opted for severe stress levels, and one could argue that the unexplained transcriptional response was caused by tissue damage. However, Prasch and Sonnewald (2013) found that relatively moderate soil drought (30% of field capacity) substantially altered the transcriptional response to warm temperature ($32°C/28°C$). Several other -omics approaches have expanded our knowledge by documenting the responses to combined abiotic stresses, but few have shed light on the molecular and cellular mechanisms involved in signal integration (Zandalinas et al., 2021). Genome-wide association studies have allowed for the identification of genomic loci important for the interaction between nutrient deficiency signaling and salt stress (Kawa et al., 2016), but these loci remain to be further

characterized. And while in-depth phenotypic and transcriptomic analysis has led to the identification of several genes involved in the cross-talk between nutrient deficiency stress (Kellermeier et al., 2014), we still lack a coherent framework for understanding this interaction.

Progress toward understanding the mechanisms of signal integration

Several studies have started to probe environmental signal integration on a mechanistic and cellular level. For example, low levels of soil salinity suppress shade avoidance (Hayes et al., 2019). Soil salinity acts through the ABA pathway to suppress brassinosteroid-activated transcription factors, thus limiting shade-induced growth. There have also been significant advances in our understanding of how light and temperature signals are integrated (Hayes et al., 2021). In several Arabidopsis accessions, simulated neighbor shade triggers an increase in petiole elongation at $22°C$, but not at $16°C$. This striking, temperature-dependent response to shade involves the receptor-like kinase ERECTA (Patel et al., 2013). More recently, it has become clear that shoot temperature perception is heavily integrated with light signaling pathways. Phytochrome and phototropin photoreceptors revert more quickly to their inactive forms at warm temperatures (Legris et al., 2016; Jung et al., 2016a; Fujii et al., 2017), with the result that light signaling is suppressed in these conditions. Additionally, the light-suppressed transcription factor PHYTOCHROME INTERACTING FACTOR 7 (PIF7) exhibits enhanced translation at warm temperatures due to changes in its mRNA structure (Chung et al., 2020). In accordance with the antagonistic relationship between light and warm temperature, shade-avoidance is more aggressive at high ambient temperatures (Romero-Montepaone et al., 2021).

The future outlook

Recently, data obtained by Prasch and Sonnewald (2013) were re-analyzed and used by (Azodi et al., 2020) to train machine learning models to predict cis-regulatory elements required for the synergistic response to multiple stresses. Novel approaches using computational models including cellular and functional structural modeling are a promising avenue to capture and predict interactions. In addition, Morales et al. (2021) reported detailed phenotypes of Arabidopsis plants in response to drought and temperature stress in combination with recovery from flooding. Together these studies provide a starting point to investigate the mechanisms underlying consequences of combined stress on plant development and resilience.

A major limitation of most studies on signal integration is the use of single stress intensities. Many stresses have a nonlinear effect on plant traits, dependent on intensity (Figure 5A). It is likely that different signaling networks act at different intensities of the same stress. When investigating the integration of two stresses, it may be useful to plot traits in a matrix of severity (Figure 5B). Comparing heat map matrices of mutants and wild types may eventually allow us to
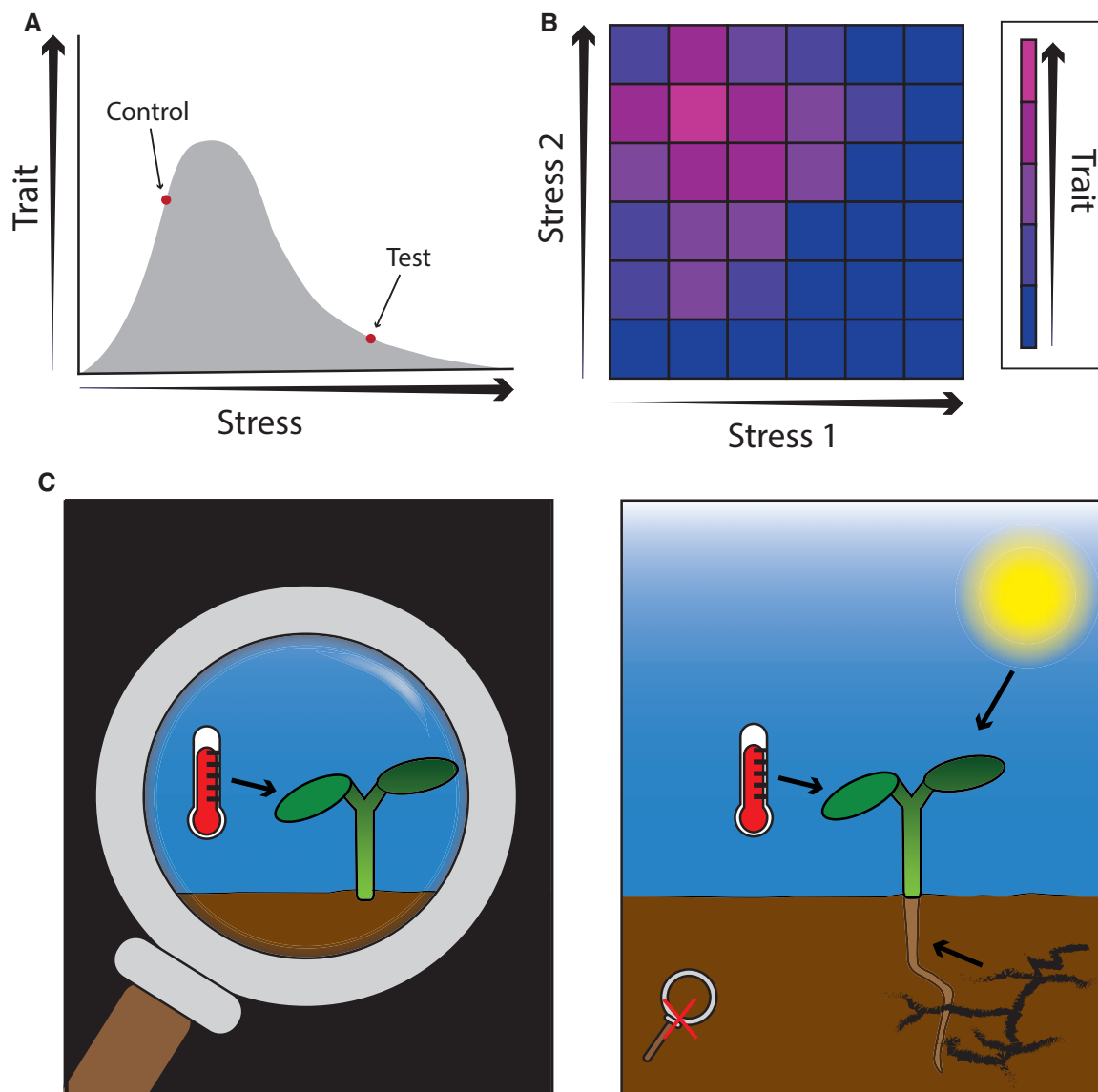
Figure 5 How do plants integrate climate signals? A, Most environmental cues do not have linear effects on plant development. When investigating the molecular basis for a particular cue, it is important to consider that different intensities of the same stress likely act through different molecular pathways. B, Representing gradients of two stresses as a heat map could be a potential way to conceptualize complex interactions. Note that the intensity of Stress 1 affects the way plants respond to the intensity of Stress 2. C, Through taking a step back and studying the interactions between signaling pathways in their relevant environmental context, we will come closer to understanding the molecular basis for plant development in natural environments.

place signaling networks within specific environmental contexts. It will also highlight environmental contexts in which phenotypes cannot be explained by known developmental regulators and offer potential avenues for future research. Transitioning to gradients of environmental conditions (rather than one, often severe stress condition) will require conceptual and computational advances. It should also be noted that environmental integration is likely to be different for different organs, tissues, or even cell types. Temperature signaling in the roots, for example, can act independently of the light signaling components in the shoot (Bellstaedt et al., 2019; Ludwig et al., 2021). Providing spatial and dose-dependent context of other relevant environmental factors (Figure 5C) will ultimately allow us to understand plant responses to abiotic stress in a realistic, complex environment.

How do trade-offs impact abiotic stress responses and climate adaptation?

(By Robert W. Heckman and Thomas E. Juenger)

Trade-offs occur when a phenotype that confers an advantage in one context also confers a disadvantage, whether in the same or a different context (Agrawal, 2020). A classic trade-off is between carbon gain and water loss during photosynthesis: when stomata open to absorb CO_2, they lose

H_2O (Schulze and Hall, 1982). Numerous ways of mitigating this trade-off have evolved in plants, from minor adaptations like changes in stomatal behavior or development to major innovations like C4 and CAM photosynthesis. From an evolutionary perspective, trade-offs impose costs that constrain the ability of plant populations to evolve in response to selection (Roff and Fairbairn, 2007; Donovan et al., 2011). These constraints may prevent populations from reaching adaptive peaks, particularly in new environments, which can limit species' ranges and the distribution of habitats suitable for particular crops (Blows and Hoffmann, 2005; Shaw and Etterson, 2012; Dwivedi et al., 2021). Trade-offs can originate from different biological processes, like the specialization of ecotypes in different habitats (Agrawal, 2020). These differences in the origins of trade-offs can impact how plants acclimate, and how plant populations evolve, in response to climate change.

Here, we focus on trade-offs that occur within species as they are most likely to be the subject of ongoing evolution that can drive adaptation. Trade-offs within a species or population can result from pleiotropy and genetic linkage and the degree of standing genetic variation (i.e. genetic architecture) (Saltz et al., 2017). Pleiotropy occurs when the same gene encodes multiple traits; linkage occurs when genes encoding different traits are located in close physical proximity on a chromosome, reducing recombination and resulting in coinheritance (Lynch and Walsh, 1998; Mackay, 2001). In the short term, trade-offs can constrain evolution by linking particular trait combinations and making other trait combinations less likely (Walsh and Blows, 2009). Trade-offs can also be reinforced by correlational selection, which occurs when certain combinations of traits, rather than traits in isolation, are advantageous (Sinervo and Svensson, 2002). Trade-offs can also occur among populations within species. These trade-offs often arise due to local adaptation, which occurs when plants perform better in their home environment than plants from a different environment (Kawecki and Ebert, 2004). Local adaptation results from multiple factors, including antagonistic pleiotropy, where an allele at a genetic locus leads to high relative performance in one environment and low relative performance in a contrasting environment, and conditional neutrality, where an allele confers high (or low) relative performance in one environment and has no impact on performance in a contrasting environment (Anderson et al., 2011).

Understanding how and when trade-offs operate will help biologists to gain the benefits of a trade-off while avoiding the costs. To do this, context is often critical. Trade-offs in allocation to different functions may be hidden when plants grow under benign conditions, such as those in many agronomic and laboratory settings (Roff and Fairbairn, 2007). When plants grow under more natural, stressful conditions, the trade-offs may be exposed, leading to unexpectedly suboptimal performance (MacTavish and Anderson, 2020). This can occur because benign conditions often increase the total resource acquisition by plants. For example, a trade-off between root and shoot biomass is commonly invoked, because when resources are fixed, any allocation to root biomass must come at the expense of allocation to shoot biomass (Shipley and Meziane, 2002). But, in nature, many plants with large root systems also have large shoots, suggesting that no trade-off exists. This failure to detect an allocation trade-off occurs when failing to account for differences in resource acquisition.

Context is also key when considering attributes of wild species in agronomic environments. Often, the characteristics that make wild plants successful become liabilities in crops. In wild plants, many stress-response strategies ensure plant survival via reduced growth rates or stress tolerance strategies, including slow growth, leaf abscission, or early flowering (Fang and Xiong, 2015). In agronomic conditions, where rapid growth and high yield are prized, these stress responses are often maladaptive (Maggio et al., 2018). This may make extremely well-adapted, stress-tolerant wild relatives (or their adaptive strategies) poor targets for domestication or crop improvement. Instead, biologists should re-examine some ideas about what makes plants successful in resource-rich, agronomic environments. For instance, domestication simultaneously increased growth rates and reduced drought tolerance in *Helianthus annuus* (Koziol et al., 2012). Traits like stay-green sorghum and maize circumvent the plant's natural stress response (Zheng et al., 2009; Jordan et al., 2012) and are usually beneficial in an agronomic setting. Reducing other natural stress responses like high water-use efficiency, which often leads plants to leave water in the soil at the expense of potential growth, could be targets of similar innovation (Leakey et al., 2019).

When trade-offs occur within species and there is standing genetic variation in performance or yield, biologists can leverage the possibility of continued evolutionary change to break trade-offs and remove conditionally deleterious variants. In most cases, trade-offs can be broken by consistent selection orthogonal to the direction of the trade-off or by crossing locally adapted genotypes to break up linkage (Conner, 2003; Agrawal et al., 2010) and through multi-trait genomic prediction-based breeding for large mega-environments (Wallace et al., 2018). The trade-offs among locally adapted genotypes that result from conditional neutrality are an ideal target. Since there is no cost of conditionally neutral alleles in a disfavored environment (Anderson et al., 2011), breeding that focuses on combining many conditionally neutral alleles may quickly generate generalist ideotypes, avoiding trade-offs altogether. A similar approach can be used in a molecular context: Kudo et al. (2019) found that drought tolerant rice plants over-express stress-inducible genes, like *DREB1A*, which reduces growth. By breeding plants that co-expressed growth-promoting genes, like *GA5* and *PHYTOCHROME INTERACTING FACTOR4* (*PIF4*), this trade-off could be eliminated (Kudo et al., 2019).

The consequences of failing to break trade-offs can be severe, especially when trade-offs constrain the ability of plant populations to adapt to keep pace with ongoing global

change (Chapman et al., 2012). Left alone, species that cannot adapt to rapid global change will need to acclimate or will likely go locally extinct (Hoffmann and Sgrò, 2011; Christmas et al., 2016). In these cases, biologists can also use our understanding of trade-offs to move species with desirable attributes to matching environments (e.g. targeting crop species to the most suitable environments or combining certain species to enhance ecosystem services) (Willi and Van Buskirk, 2022).

Trade-offs are, to some extent, inevitable in biology. But, trade-offs do not need to hamstring our ability to respond to climate change. As the climate becomes more variable and moves further away from historical averages, the target environments for crop breeding will become less obvious (Chapman et al., 2012). Thus, the specialized strategies exhibited by locally adapted crops may become less valuable. A major challenge will be to identify the trade-offs that are most limiting to desired outcomes (e.g. increased production or resilience to climatic extremes) and use our biological insight and engineering principles to break, alleviate, or circumvent those trade-offs. Gene stacking to produce generalist ideotypes and artificial selection in the direction orthogonal to a trade-off are promising approaches that may be feasible in the short term. In the longer term, more extreme interventions, like genetically engineering crops to use C4 photosynthesis or to become perennials, may be required to break or alleviate some of the most recalcitrant trade-offs. Given the importance of trade-offs for so many aspects of biology, their study can be a promising approach for enhancing the resilience of our agricultural and natural systems in an increasingly variable world.

How does the circadian clock "gate" plant responses to abiotic stress?

(By Paloma Mas)

Severe drought, extreme temperatures, and changes in salinity all disturb plant cellular homeostasis and cause deleterious effects on crop growth and productivity. Throughout evolution, plants have developed a battery of responses to reach a cellular status that is tolerant or compatible with harsh conditions (Markham and Greenham, 2021). Understanding the array of responses triggered by abiotic stress can provide useful information to obtain crop varieties adapted to stress (Iannacone et al., 2012; González-Guzmán et al., 2022). Among the different signaling pathways involved in plant responses to stress, the circadian clock stands out as a main cellular mechanism able to measure time and to coordinate key biological processes in synchrony with the environment (Sanchez and Kay, 2016). The proper function of the circadian clock enables plants to anticipate the daily changes in the environment, controlling the timing of growth, development, and responses to biotic and abiotic stresses (Sanchez and Kay, 2016).

Mutation of clock components disturbs the ability of plants to adapt to environmental stress conditions (Seo and Mas, 2015; Bonnot et al., 2021). The number of stresses connected with the circadian clock is ample and includes drought, heat, cold, or redox imbalance (Grundy et al., 2015). Interestingly, the circadian clock not only regulates daily or seasonal oscillatory stresses such as heat stress during the day or severe cold during the night, but also continuous stresses with no obvious oscillations, like high salinity (Park et al., 2016). The anticipation provided by the circadian clock may enable plants to prepare in advance against stresses that diurnally or seasonally oscillate, and thus conferring an adaptive advantage. However, the regulation of constant stresses may rely on specific clock components that act independently of their function within the clock. It is also possible that the constant stress is related to other stresses and pathways in which circadian timing is relevant.

Based on the pervasive role of the circadian clock in the regulation of abiotic stress responses, the focus of attention is directed toward the use of the circadian system for improved tolerance to a broad combination of stresses without imposing detrimental pleiotropic effects such as growth arrest, or reduced yield. To that end, multiple strategies can be adopted, but particularly interesting are those related to a central function of the circadian system, known as "gating," whereby the clock differentially regulates the magnitude of the plant response to environmental signals depending on the time-of-day (Seo and Mas, 2015). The circadian peak of expression of many genes involved in stress responses coincides with the recurrent peak time of the stress (Bonnot et al., 2021), and so, a large proportion of the heat- and cold-responsive transcriptome is gated by the clock (Covington et al., 2008; Blair et al., 2019). Interestingly, the clock is also able to gate stress responses depending on the time of the year (Lee and Thomashow, 2012). For example, key factors involved in cold responses oscillate with higher amplitude under short-day conditions, providing improved tolerance to cold conditions during the winter time. Thus, using circadian gating might be advantageous for plants to restrict their response only to the most appropriate and needed time, as opposed to a constitutive response normally associated with high energy demand and with the penalty on growth and yield.

But, what are the mechanisms behind the gating function and regulation? Although most studies have focused on transcriptional control, posttranscriptional regulation can be also gated by the clock. For example, alternative splicing has been associated with heat stress responses (Ling et al., 2021), and Splicing Factor 30 could be one of the many plausible candidates linking the clock with alternative splicing and heat stress (Bonnot and Nagel, 2021). Identifying all the components of the splicing machinery gated by the clock, and the alternative splicing isoforms functionally relevant under single or combined stress conditions is an interesting area of ongoing and future research. Similarly, studies showing the connection of alternative polyadenylation with the clock (Yang et al., 2020), and with stress (Yang et al., 2021) open the way for functional studies on the circadian gating of alternative polyadenylation. New areas of study can

specifically focus on the circadian coupling of transcription with polyadenylation, splicing, or RNA modification (N6-methyladenosine, 5-methylcytosine, or pseudouridine) (Yang et al., 2021). The clock also selectively redefines the pool of mRNAs to be translated under heat stress, controlling about one-third of the circadian- and heat-dependent translated proteins (Bonnot and Nagel, 2021). Expanding these studies to other abiotic stresses will provide a global view of how circadian clock gating impacts the abiotic stress-related translatome. The key role of posttranslational modifications on abiotic stress responses (Hashiguchi and Komatsu, 2016) also paves the way for future studies on how circadian gating controls other posttranslational modifications such as phosphorylation, ubiquitination, glycosylation, etc. to regulate protein function and localization in response to abiotic stress (Figure 6).

Changes in DNA methylation and histone modifications result in epigenetic variation that provides phenotypic plasticity and plant adaptation to changing environments (Miryeganeh, 2021). As the circadian clock is also closely connected with chromatin remodeling (Chen and Mas, 2019), it would be interesting to fully explore the circadian gating of epigenetic responses to stress. Research could focus on identifying the full array of chromatin "readers" and "erasers" controlled by the clock and their connection with the stress-responsive loci. The fact that epigenetic marks can trigger stress memory in primed plants also places the spotlight on identifying the role of the clock in providing a long-term memory for the stress response. To obtain meaningful results, the stress-related experiments should be performed under growing conditions that mimic as close as possible the natural growth environment (Panter et al., 2019). Likewise, analyzing combinatorial stresses that usually appear simultaneously or sequentially in nature (e.g. heat and drought) might provide much more reliable and relevant information to use for enhancing crop tolerance to abiotic stress (Rivero et al., 2022). Studies could also switch the focus from whole plants to single cells in order to get meaningful conclusions about stress perception, signaling, and responses in specific cell types. Also, looking back to plant ancestors and comparisons of domesticated crops and their wild relatives (Markham and Greenham, 2021; Figure 6) will surely provide novel avenues to understand how the circadian gating by the clock can be exploited for adaptation to environmental stresses, thus providing novel opportunities for targeted approaches for improved tolerance to abiotic stress (Bhatnagar-Mathur et al., 2008).

How do lipid-derived second messengers translate abiotic stress information into cellular responses for stress acclimation?

(By Teun Munnik)

Phospholipids are crucial building blocks for membrane function. They create the bilayered, liquid structure that surrounds every cell, organelle, and endosomal compartment, and hosts thousands of integral and peripheral membrane proteins essential for membrane energization, signal detection and transduction, as well as primary and secondary metabolism (Munnik et al., 2021). Phospholipids are indispensable for cell function (Noack and Jaillais, 2020).

Besides a structural function, phospholipids have also emerged as crucial signaling molecules, either as precursors of signaling molecules or as lipid second messengers themselves. The best examples come from polyphosphoinositides (PPIs), which are inositol-containing phospholipids that can be phosphorylated at the D-3, D-4, and/or D-5 position of the inositol ring, thus creating five distinct molecular species in plants (i.e. PI3P, PI4P, PI5P, PI(3,5)P_2, and PI(4,5)P_2) as well as two more that are present in metazoans but have not been detected in plants [PI(3,4)P_2 and PI(3,4,5)P_3] (Munnik and Testerink, 2009; Gerth et al., 2017). PPIs are typically low abundance lipids that escape detection by common mass spectrometry methods, and turn over rapidly. They act as biochemical and biophysical landmarks that contribute to membrane identity, signaling, and compartment morphodynamics, with each PPI species accumulating in different set of endomembranes thus helping to define membrane identity (Gerth et al., 2017; Dubois and Jaillais, 2021).

In the world of PPI signaling in animals there has been something of a revolution over the last decade that has manifested itself in an increasing appreciation of PI(4,5)P_2 as a regulator of cellular events in its own right. PI(4,5)P_2 has been known for a long time to fulfill a crucial role as the substrate for two major signaling pathways: the phosphoinositide (PI)–phospholipase C (PLC) pathway that generates inositol-1,4,5-trisphosphate (IP$_3$) and Ca^{2+} release, as well as diacylglycerol (DAG) to activate protein kinase C (PKC); and the PI 3-kinase pathway that generates PI(3,4,5)P_3, a crucial lipid second messenger in regulating cell proliferation and many other physiologically important processes (Balla, 2013; Dickson and Hille, 2019).

Yet, higher plants clearly have a different agenda. They do have the PI–PLC pathway to generate the above-mentioned signaling molecules, but they have no recognizable PKC or IP$_3$ receptors, and no PI 3-kinases of Type I, the group that uses PI(4,5)P_2 as a substrate (Munnik and Testerink, 2009). Yet Arabidopsis has 11 Type I PIP 5-kinases (PIP5Ks), nearly four times as many as us animals! And why despite this do plants maintain PI(4,5)P_2 levels 1–2 orders of magnitude lower than metazoans? The only logical answer to these questions is that PI(4,5)P_2 has an even more central role as a signaling entity in its own right than it does in animal biology. In plants, there seems to be a proliferation of PI(4,5)P_2 functions in conjunction with an increasing understanding of PI(4,5)P_2 compartmentalization (Munnik, 2014; Doumane et al., 2022). Together, these new insights are revolutionizing how we think about this ancient and original PPI.

Knockout of some Arabidopsis PIP5Ks (PIP5K1–PIP5K6) clearly indicate a role for PI(4,5)P_2 in cell polarity, in particular during cell division and polar growth of root hairs and pollen tubes (van Leeuwen et al., 2007; Kusano et al., 2008; Ischebeck et al., 2010; Tejos et al., 2014). In contrast, no or
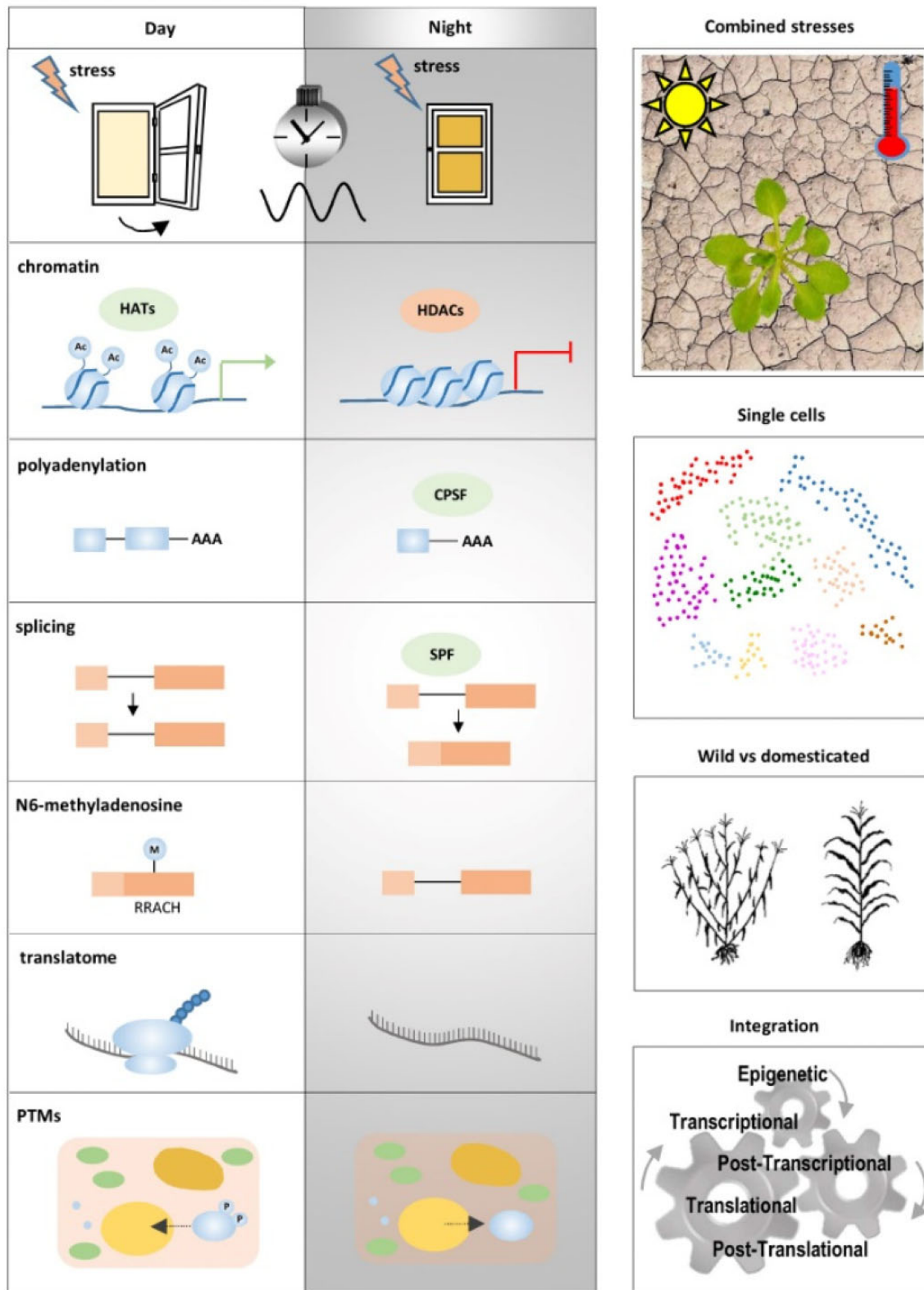
Figure 6 Circadian gating by the clock differentially regulates plant responses to stress depending on the time of day. Figurative examples of open questions related to the circadian gating via changes in chromatin, posttranscriptional, translational, and posttranslational mechanisms of regulation that ultimately define gene expression and protein function under stress conditions. Single-cell approaches with plants grown under conditions that closely follow the natural environment, including combined stresses, and comparisons of domesticated crops and their wild relatives are circadian research areas of increasing interest. Understanding the coupling among the different gating mechanisms will pave new ways for using the circadian clock to tackle the effects of abiotic stress on plants.

only subtle developmental phenotypes are observed for PIP5K7–PIP5K9 mutants as these PIP5Ks are instead involved in salinity stress and response to polyamines (Zarza et al., 2020; Kuroda et al., 2021). Genetically encoded-$PI(4,5)P_2$ biosensors (van Leeuwen et al., 2007; Simon et al., 2014) revealed that $PI(4,5)P_2$ typically accumulates at the plasma membrane, except during heat stress where additional punctates appeared in cytosol and near the nuclear envelope (Mishkind et al., 2009). What these punctate compartments are, and which PIP5K generates them, is still unknown. Genetic manipulation of $PI(4,5)P_2$ by inducible production or depletion revealed crucial roles for $PI(4,5)P_2$ in endocytosis and regulating the actin- and microtubule cytoskeleton, with dramatic consequences for development (Gujas et al., 2017; Doumane et al., 2021).

But how is $PI(4,5)P_2$ managing all this? In metazoan systems, several protein targets have been characterized, including $PI(4,5)P_2$ specific-binding domains, for example, PH, Tubby, and SEC14 (De Jong and Munnik, 2021). While several plant proteins contain such domains, their $PI(4,5)P_2$-binding specificity and functionality has remained largely unexplored (De Jong and Munnik, 2021). Since plant cells contain much lower $PI(4,5)P_2$ levels than animals, and since it is no problem to stably express $PI(4,5)P_2$-biosensors (based on exogenous PH and Tubby domains) in Arabidopsis without causing any phenotype (van Leeuwen et al., 2007; Simon et al., 2014), plants are likely to contain distinct $PI(4,5)P_2$-binding domains with a much higher affinity. Such domains, however, remain to be identified. In animals, the gating of most K^+ channels is regulated by $PI(4,5)P_2$, and there are indications this occurs in plants too (Zarza et al., 2020).

Phosphatidic acid (PA) is another important plant lipid second messenger that is typically triggered upon abiotic stress, including heat, cold, drought, and salinity stress, but also in response to pathogens and wounding (Munnik, 2001; Kim and Wang, 2020). In general, PA responses are fast (min) and generated through hydrolysis of structural phospholipids by phospholipase D (PLD) and/or through phosphorylation of DAG by DAG kinase (DGK) (Munnik and Testerink, 2009). Arabidopsis contains 12 PLDs and 7 DGKs, and 9 PI–PLCs that produce DAG by hydrolyzing PIP4 or, if generated, $PI(4,5)P_2$. How PA is involved in the different stress responses, via which PLD, DGK, and PLC, and at which cell or compartment this takes place, has been a central theme of the last decade, and will still be in the next. Complementary approaches include KO and OE mutants on PA production, isolation, and characterization of PA targets, and the construction of genetically encoded-PA biosensors to monitor PA in living cells (Munnik and Testerink, 2009; Platre et al., 2018; Kim and Wang, 2020; Scholz et al., 2022). While these tools helped to establish PA as a lipid second messenger that rivals the importance of Ca^{2+}, we still know very little of either PA versus Ca^{2+} specificity in responding to different stresses or their potential collaboration. Monitoring both molecules simultaneously with ratiometric biosensors would certainly help clarifying this.

Another challenge remaining is to understand PI–PLC signaling in plants (D'Ambrosio et al., 2017; van Wijk et al., 2018; Zhang et al., 2018c, 2018d). In vitro, the enzyme equally likes PI4P and $PI(4,5)P_2$ as substrate, but the general absence of $PI(4,5)P_2$ in plant plasma membranes, while PI4P is relatively abundant, makes it more likely that IP_2 is generated rather than IP_3 in vivo. This would also explain the lack of IP_3 receptors (Munnik and Vermeer, 2010; Munnik, 2014). For DAG, it makes no difference to be converted into PA. IP_2 can be step-wise phosphorylated into various inositolpolyphosphates (IPPs) that are emerging as signaling molecules, including IP_5 and IP_6, but also IPP-pyrophosphates IP_7 and IP_8 (Lorenzo-Orts et al., 2020). IP-related signaling functions in plants include intracellular Ca^{2+} release (ABA), acting as co-factor for auxin and jasmonate signaling (via TIR1 and COI1), as well as functions in RNA transport and P_i sensing (Figure 7; Munnik, 2014; Lorenzo-Orts et al., 2020). Linking all these cellular phenotypes to PI–PLC signaling remains challenging (Zhang et al., 2018c, 2018d).

While the above mostly deals with questions downstream of abiotic stress-triggered lipid signaling, we are equally in the dark on how lipid kinases, phosphatases, and phospholipases are actually activated upstream. The next decade will be an exciting venture into a much deeper understanding of how plants use signaling lipids and inositol phosphates to control their physiology and respond to stress. It will not just widen our basic knowledge of plant biology, but very likely also our understanding of stress responses that will most benefit efforts to improve plant resilience.

How can laboratory stress research be applied to continuously stressed plants in the field?

(By Hilde Nelissen)

One of the outstanding questions in plant abiotic stress biology is why the vast amount of knowledge on physiological and molecular responses to abiotic stresses has resulted in so few climate resilient crop varieties that are currently on the market (Sanchez, 2013; Blum, 2014; Nuccio et al., 2018). Among the possible reasons that hamper the translation of molecular knowledge toward application, the gap between environmentally controlled conditions versus more variable field conditions was identified as one of the bottlenecks (Nuccio et al., 2018). Controlled conditions, here used as a collective name for greenhouse and growth chamber experiments, allow higher throughput, increased speed of innovation and reduced cost compared with field trials (Simmons et al., 2021). In addition, more detailed mechanistic insights and mode of action studies often require controlled conditions, where responses to changes in the environment can be monitored with respect to time, space, amplitude, and other factors related to both the stress applied and the response output (Verslues et al., 2006; Bao et al., 2014; Alejo-Jacuinde et al., 2022; Hu et al., 2022). On the other hand, field trials provide valuable information about the
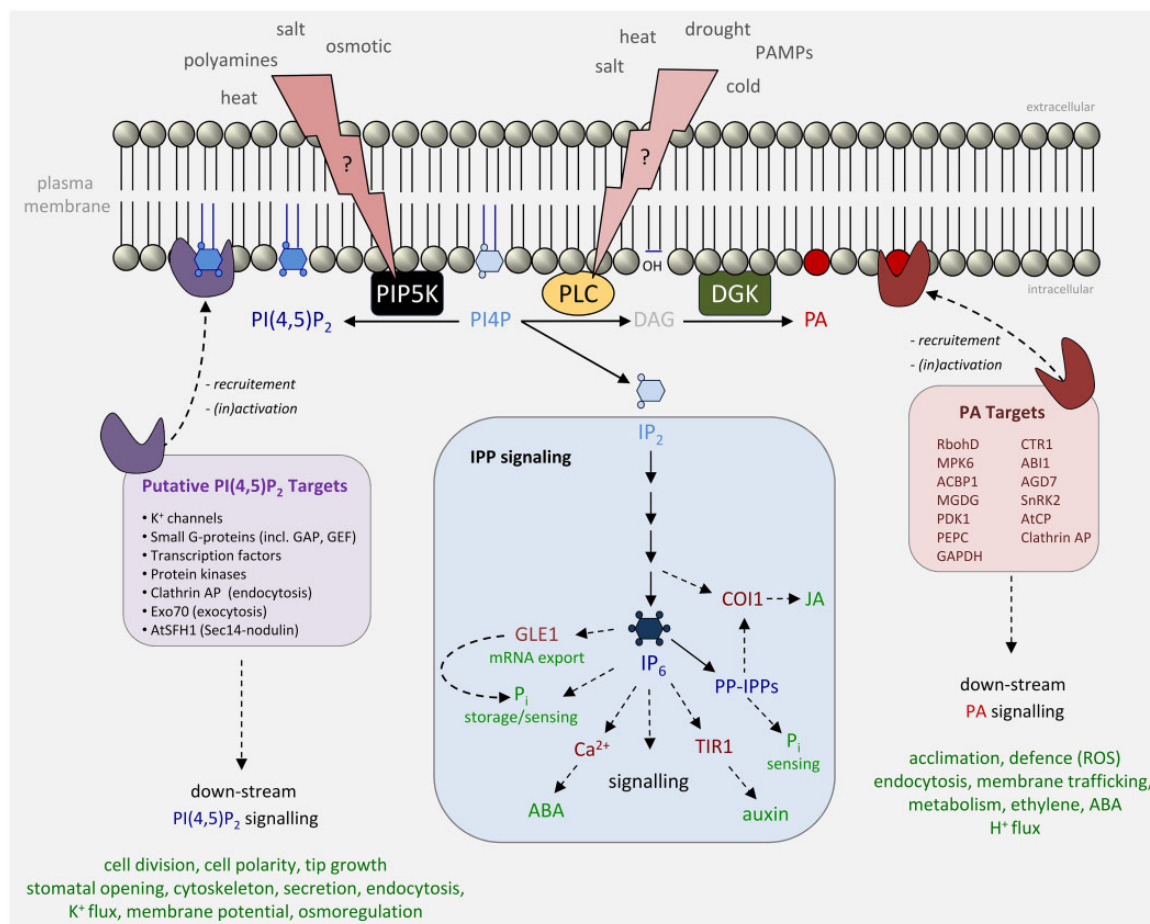
Figure 7 Possible signaling mechanisms of PI(4,5)P$_2$, PA, and PLC in plant abiotic-stress signaling. PI(4,5)P$_2$ concentrations in plants are extremely low but synthesis is rapidly triggered by activation of one or more of the 11 PIP5Ks of Arabidopsis via unknown receptors or activation mechanisms (indicated by "?"). As such, PI(4,5)P$_2$ is triggered by heat, salinity, osmotic stress, and polyamines, but also during polar growth, predominantly accumulating at the plasma membrane. This likely causes recruitment of target proteins to the membrane or (in)activation of targets that are already present at the membrane. Putative plant targets are based on the mammalian literature and correlations with plant literature. Plant PLCs likely hydrolyse PI4P and generate membrane-bound DAG and the water-soluble headgroup, IP$_2$. Again, receptors and activation mechanisms remain unknown (indicated by "?"). The newly formed DAG is rapidly phosphorylated by DGK to form the signaling lipid PA for which several protein targets have been identified. IP$_2$ can be stepwise phosphorylated by IPP multi-kinases to produce IP$_5$ and IP$_6$, and IPP-pyrophosphates IP$_7$ and IP$_8$, for which several signaling functions and targets are emerging. Protein targets are indicated in purple [PI(4,5)P$_2$] or dark red [PA]. Lipid (-derived) signals are indicated in blue. Solid lines indicate metabolic conversions; dashed lines represent "activation" or "cause." Effects are indicated in green.

potential of a stress-tolerant product for marketable yield stability without yield drag. However, field evaluations are costly and labor-intensive because of the experimental design and replication needed to detect a desired effect, management logistics, and weather hazards. Thus, given the throughput and the costs involved, the logic pipeline for plant improvement is a funnel-like screening in which many lines are screened in controlled conditions, of which the most promising leads subsequently undergo field evaluations (Simmons et al., 2021). However, the realization is growing that the inability of typical laboratory- or greenhouse-controlled conditions to properly model agronomic environments hampers the translation of knowledge on abiotic stresses to climate resilient crops (Sanchez, 2013; Blum, 2014; Nuccio et al., 2018; Simmons et al., 2021).

As the name implies, controlled conditions aim to maximally standardize the growing conditions to reassure reproducibility and to facilitate detailed studies on the effects of a limited perturbation. In controlled condition stress experiments, typically temperature, humidity, irrigation, or light is altered or a substance is added to mimic an abiotic stress response. Ideally, the other environmental parameters are kept as standardized and stable as possible, so that only one or a combination of a few abiotic stresses occur simultaneously. However, to fully understand the impact of the applied stress(es), it might be necessary to monitor additional parameters, such as soil water content in drought experiments. The stress(es) can be maintained for a substantial time of the plant's development, sometimes followed by stress alleviation, but more frequently stress treatments in

greenhouse and growth chamber conditions are short and severe to evoke molecular changes. The setup of such an "ideal" laboratory stress assay is often more dependent on the greenhouse and growth chamber facilities and the assay robustness than on reflecting the actual field conditions.

Anyone who has performed field trials or has a private garden realizes that there is no typical growth season that could be represented by standardized, control conditions. Even when temperature and precipitation are close to the multi-year average, there will be periods of extreme weather conditions. Besides the conspicuous extreme weather conditions, there are also less obvious consequences of climate change that can have detrimental effects on crop yield. For example, the global rising temperature is not only experienced during the daytime. Increased night temperature is also a problem for plants in that it enhances night respiration that increases utilization of photo-assimilates and thereby reduces the amount of carbon available for grain filling (Desai et al., 2021; Xu et al., 2021). In the field, some stresses are continuously present, like heavy metals in the soil, but other stresses build up more gradually, like drought, and can occur during any developmental stage with distinct impacts on yield (Verbraeken et al., 2021). In field conditions, stresses rarely occur alone or in a coordinated way, and temperature, humidity, irrigation, light, and several additional factors fluctuate continuously rendering the plants constantly stressed.

The fact that field-grown plants are continuously in a stressed state is reflected by the transcriptomic changes between plants of the same genotypes, grown in controlled conditions and multiyear field trials. Several stress-related genes that confer abiotic stress tolerance in over-expression lines exposed to controlled stress conditions were massively upregulated in field-grown versus laboratory-grown plants, even under relatively normal field conditions (Nelissen et al., 2020). Even neighboring plants of the same genotype grown in one field displayed transcriptional differences in stress-related genes (Cruz et al., 2020). Studies that also take into account the weather conditions when analyzing field transcriptomics (Nagano et al., 2012) or that incorporate measurements of plant water status show that individual field-grown plants sense a micro-environment and react molecularly to local changes in environmental changes by altering stress-related genes, some of which were already identified in controlled conditions.

We showed that not only known stress-related genes were differentially expressed between the controlled conditions and the field (Nelissen et al., 2020). Genes involved in processes such as shade avoidance were also differentially expressed between the laboratory and the field, which, in turn, may interact with stress responses (Hayes et al., 2019), complicating the situation even more. In addition, shade avoidance is a response that breeding attenuated to achieve higher planting density, an agronomic practice that is often overlooked in laboratory or greenhouse experiments. In such experiments, plants are grown in pots filled with potting soil, so that individual plant irrigation schemes or nutrient applications can be monitored. The pots are arranged to optimize greenhouse space, irrespective of whether the plants under study are typically grown as high-density row crops or subjected to other management regimes. In this way, differences in inter-plant population dynamics, interactions with other organisms, or effects of crop rotation or inter-cropping are not accounted for when screening in controlled abiotic stress conditions. By individualizing every plant, field-born local differences that cause molecular responses (Cruz et al., 2020) or trigger differences in developmental programming (Robbins and Dinneny, 2015) can be overlooked, along with the effects of soil differences related to recent cropping history or biotic interactions (Beirinckx et al., 2020).

The fact that abiotic stress-related genes, identified in controlled abiotic stress experiments, are transcriptionally regulated in the field, confirms their potential usefulness in improving plant resilience. However, this also urges plant scientists to gain a better insight into the molecular "state of mind" of field-grown plants in order to come up with more clever ways to modulate these genes and the corresponding networks to improve plant performance. To achieve this, the fields of breeding and molecular biology need to work together more closely to grow and molecularly profile new varieties with improved traits in both field and controlled conditions, irrespective of whether the lines were generated by traditional breeding, genome editing, or transgenic approaches. This would enable a virtuous cycle of each new variety to be field phenotyped and molecularly profiled, providing information for further improvements (Figure 8). However, for such efforts to have a maximal impact on agriculture, there is also a need to ease and harmonize regulations that govern the use of genome-edited or transgenic crops for sustainable agriculture and food production.

Where is the plant most hydraulically vulnerable?

(By Lawren Sack, Craig Brodersen, and Thomas N. Buckley)

When storms threaten the power grids distributing electricity across a continent, we need to know their weakest links to better prepare for and quickly remediate calamitous failures (Weiss and Weiss, 2019). Equally, we need to ask, as increasingly frequent droughts face plants throughout our globally important and vulnerable ecosystems (Hammond et al., 2022), where are their weakest points? That is, where within the plant does the impact of water stress trigger the most severe reductions in leaf gas exchange and whole plant productivity, the most irrecoverable damage, and the greatest risk of mortality—especially with aggravating stresses such as insect outbreaks and fire. Addressing this question is critical to predicting the impacts of climate change on the local and global distribution of ecosystems, the future of agricultural and forest gas fluxes, and even the behavior of the atmosphere and climate system. Equally, answering this
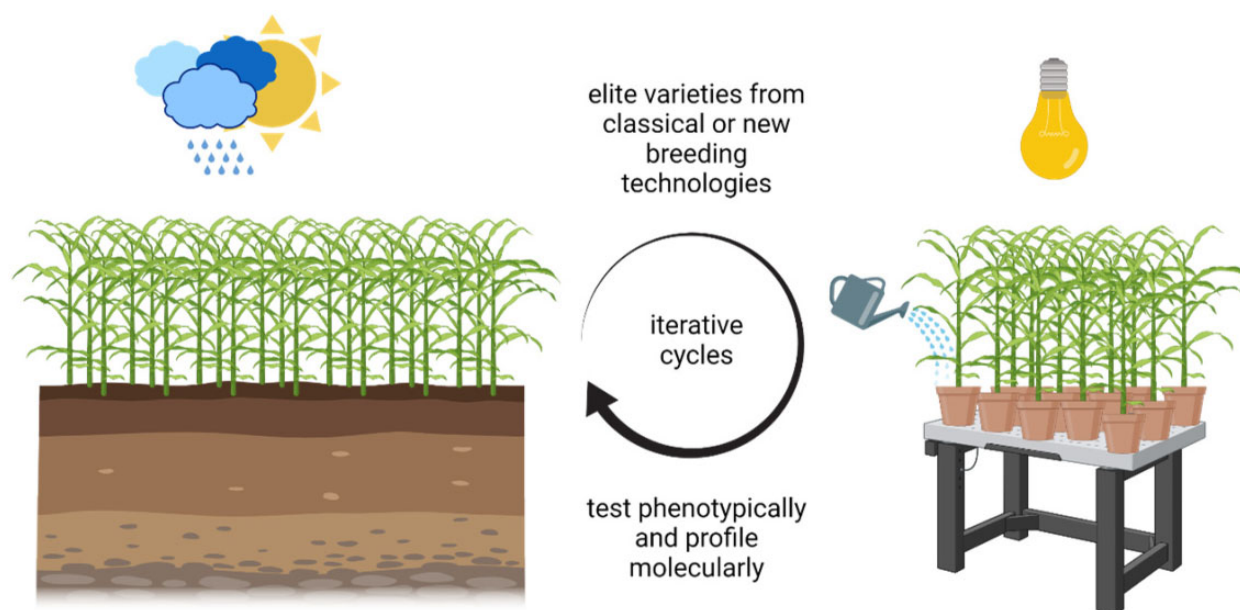
Figure 8 Schematic representation of the differences in management, light, and irrigation schemes between plants grown in the field (left) and in controlled conditions. Iterative cycles of field testing and molecular profiling of all lines with putatively improved traits will be necessary to increase our understanding how the stresses that plants experience in the field relate to the controlled abiotic stress conditions. Created with BioRender.com.

question will inform the breeding of drought-resistant crops for food security.

Sensational or not, the analogy of plants as power grids is in fact a well-established quantitative approach. Much understanding of plant hydraulics arose from the classical comparison of the soil–plant–atmosphere continuum to an electrical circuit, where flow paths are resistors and water potential gradients are voltages, subject to the application of the analogy to Ohm's Law (Van den Honert, 1948; Tyree and Zimmermann, 2002):

$$\Delta V = IR,$$

where V is the voltage, I the current, and R the electrical resistance,

By analogy,

$$\Delta \psi_w = ER,$$

where ψ_w is the water potential, E the transpiration rate, and R the hydraulic resistance. Thus, the stronger the flow rate, and the resistance to water movement, the stronger the drop in ψ_w (or pressure) across the organ, or whole plant, between the soil and the atmosphere; conversely, the stronger the ψ_w gradient driving force, the stronger the flow rate through the system or any component. This electrical analogy stands in for more detailed theory of fluid mechanics and irreversible thermodynamics (Edlefsen, 1941; Slatyer and Taylor, 1960; Gibbs, 1961; Granger, 1995), and can have drawbacks (namely, when factors other than ψ_w, such as thermal gradients drive water movement; Rockwell et al., 2014; Buckley et al., 2015). On the other hand, the analysis of the plant and its environment as a network of electronic

components yields a wealth of predictions and mechanistic representations by considering analogies of tissues, plants, and ecosystems as including fixed resistors, variable resistors (potentiometers), capacitors, and diodes. Studies using this approach to analyze the distribution of hydraulic resistance within the plant network suggest that the extremities of the plant represent key bottlenecks to water flow, with the leaves and roots accounting for $>75\%$ of total resistance and stems $<25\%$, in a wide variety of growth forms (Tsuda and Tyree, 2000; Sack et al., 2003; Domec et al., 2009). Moreover, these resistances are dynamic, subject to internal and environmental control. Most famously, in every component of the system, hydraulic conductance (K), the inverse of resistance, declines precipitously at lower ψ_w, a "hydraulic vulnerability" arising from a multiplicity of processes (Figure 9, A–F). In soils, air replaces water and thus removes flow paths for water to be sucked into the plant. In xylem throughout the plant, under strong dehydration, cavitation occurs, which is the formation of air- or vapor-filled conduits that block water flow (Figure 9, E and F; Tyree and Zimmermann, 2002), However, in leaves and roots, water flows not only through xylem but also through living tissues (on the way from the soil to the root xylem, and from the leaf xylem to the stomata). Before embolism forms in the xylem, in dehydrating roots, cortical lacunae may form that break the hydraulic connection between the root xylem and the soil (Figure 9, A and B; Cuneo et al., 2021), and in dehydrating stems and leaves, tissues shrink, which may influence flow pathways around and within cells, as the water channel proteins that traverse cell membranes, known as aquaporins (AQPs), can be gated even under mild dehydration (Scoffoni
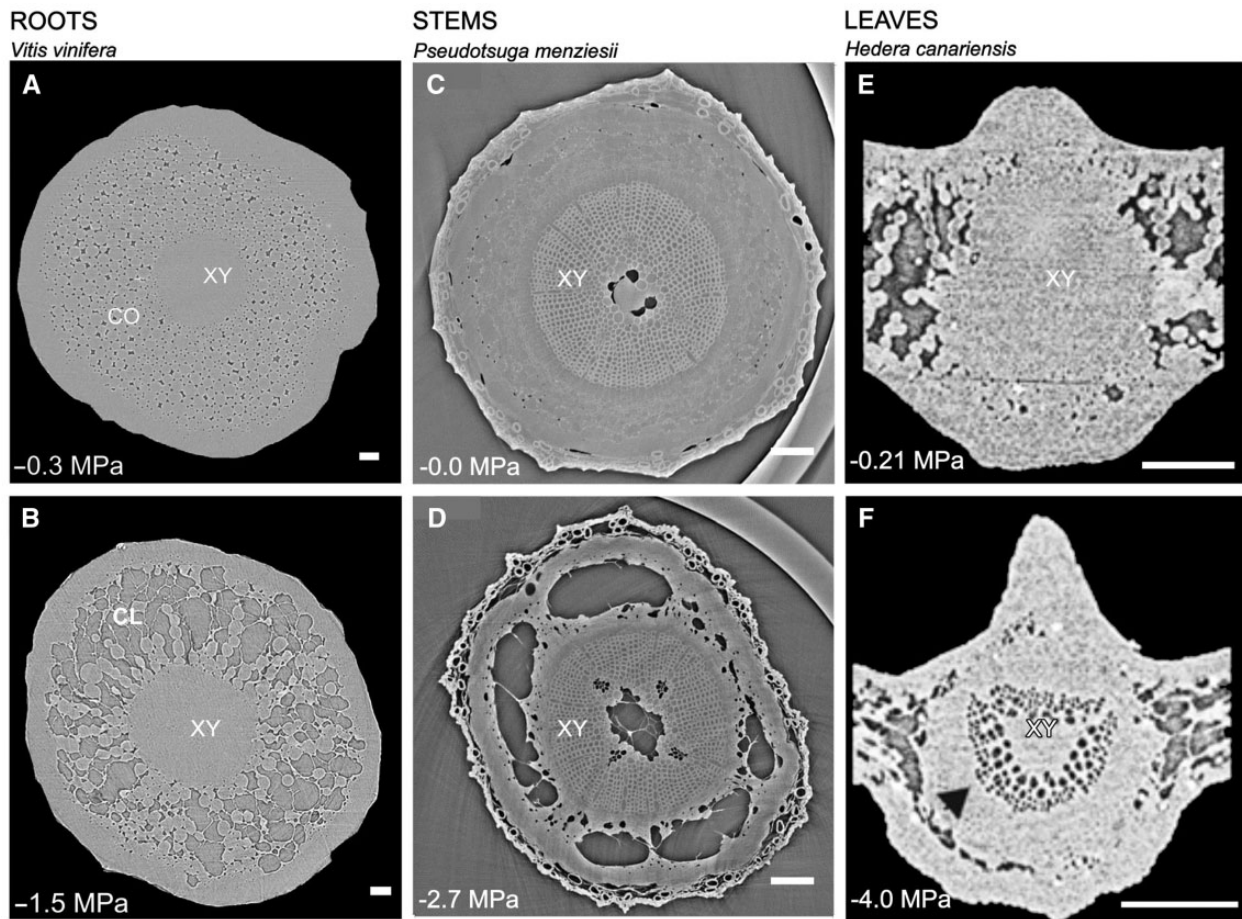
ROOTS
Vitis vinifera

STEMS
Pseudotsuga menziesii

LEAVES
Hedera canariensis

Figure 9 The effects of drought on tissues across plant organs. Representative transverse X-ray micro-computed tomography images show the in vivo functional status of the vascular system and the surrounding tissues under well-watered (A, C, E) and drought (B, D, F) conditions in roots, stems, and leaves. In (A) and (B), the roots of grapevine (*Vitis vinifera*) the xylem network (XY) remains functional under a range of water potentials (ψ_w) but mild drought (−1.5 MPa) leads to massive cellular damage and formation of cortical lacunae (CL in B) that physically decouple the vascular system from the surrounding soil (Cuneo et al., 2021). In (C) and (D), the vascular systems of the immature stems of conifer seedlings (*Pseudotsuga menziesii*) are highly drought tolerant (XY), but deformation of the cortex under strong drought in these resembles that in roots (Miller et al., 2020). In (E) and (F), leaves of ivy (*Hedera canariensis*), subjected to very strong dehydration (−4.0 MPa) show xylem cavitation in the midrib (XY) and the strong mesophyll tissue deformation that occurs during drought exceeding the turgor loss point (Scoffoni et al., 2017). Bars = 200 μm in (A) and (B); 100 μm in (C) and (D); 250 μm in (E) and (F). Values in the lower left corner of each panel indicate the ψ_w of the plant during the experiment.

et al., 2014; Figure 9, C–F). Given their extra-xylem pathways, and despite similarity of their xylem in resistance to embolism (Zhu et al., 2016a, 2016b; Klepsch et al., 2018; Li et al., 2020; Smith-Martin et al., 2020), leaves and roots tend to be more hydraulically vulnerable than stems (Scoffoni et al., 2017; Albuquerque et al., 2020; Cuneo et al., 2021).

Does the combination of the greater bottlenecks and vulnerabilities within leaves and roots than stems make these extremities the plant's most sensitive fuses under strong drought? Indeed, many have concluded that these organs should be more vulnerable, to protect stem xylem from tensions that would drive irreparable cavitation, as the stem is longer-lived and more costly to replace, ideas known as the "hydraulic-segmentation" and "vulnerability-segmentation" hypotheses (Tsuda and Tyree, 1997; Tyree and Zimmermann, 2002; Pivovaroff et al., 2014). Yet, the

challenge faced by plant organs will shift as drought proceeds, along with the distribution of different ψ_w within the plant. In drought mild enough that stomata remain open, transpiration causes ψ_w to be lowest in the leaves, and given that the leaf is a bottleneck and highly vulnerable, K loss in leaves can be drastic (Hernandez-Santana et al., 2016; Scoffoni and Sack, 2017; Albuquerque et al., 2020). Leaf K loss may, however, act as a brake on water loss by amplifying stomatal closure (Scoffoni and Sack, 2017). In severe droughts, where turgor is lost and stomata are fully closed, the plant will be close to equilibrium with the soil, with all organs experiencing similar ψ_w. As the soil and plant dehydrate further, exacerbated by the low "minimum conductance" from incompletely closed stomata and/or across the cuticle (Martin-StPaul et al., 2017), the leaf xylem may suffer embolism, starting with major veins and proceeding

to minor veins, and this embolism may trigger leaf death (Brodribb et al., 2021). When substantial cavitation occurs in the stem xylem, it usually kills the stem, because cavitation tends to beget more cavitation, in a vicious cycle of "catastrophic embolism"—with gas bubbles spreading, uncontrolled, among conduits (Tyree and Zimmermann, 2002). As for the roots, the vulnerability of their xylem and outside-xylem pathways may also be strong (Brunner et al., 2015; Rodriguez-Dominguez and Brodribb, 2020). Across species, this sequence of hydraulic decline—leaves before stems and roots—tends to be typical, and species' thresholds for declines of K in organs and the effects of drought-induced damage are correlated (Bartlett et al., 2016; Dayer et al., 2020).

This overall simple scenario, however, has been tested in a few hundred plant species at best, and not in sufficient detail to fully parameterize the hydraulic network for the bulk of plants. Particularly little is known of the hydraulic vulnerability of roots, due to technical difficulties; recent studies on potted plants of a few species have proposed that the decline of hydraulic conductance in the root and/or root–soil interface can be strong enough even in relatively moist soil to contribute to stomatal closure (Abdalla et al., 2021; Bourbia et al., 2021; Duddek et al., 2022). Further, the potential role of the root sheath and mycorrhizae in modulating or perhaps protecting root and root interface hydraulic conductance has yet to be fully clarified (e.g. Boomsma and Vyn, 2008; Brunner et al., 2015). Indeed, the fine roots are more vulnerable than older roots, and their vulnerability needs separate quantification, especially if fine root death and turnover occur even under mild soil moisture deficit (Cuneo et al., 2021).

Given these numerous unknowns, the location of hydraulic triggers for declines in gas exchange and death is a critical avenue for research. When enough hydraulic conductance is lost, water cannot be transported into and throughout the plant, and a spiral of mortality begins, potentially exacerbated by carbon starvation and other biotic and abiotic stresses (Choat et al., 2018; Hammond et al., 2022; McDowell et al., 2022). Yet, water storage can buffer given organs from loss of hydraulic conductance, and protect the plant from dehydration, especially when the plant retracts its roots to prevent water loss to the soil. The general magnitude and role of water storage "capacitors" is another, related critical unanswered question (McCulloh et al., 2014; Knipfer et al., 2019). Least of all is known about the triggers for death, and their timing and general order during dehydration, for cells within tissues, tissues within organs, and organs throughout the plant. Indeed, new concepts are needed—there is no generally agreed definition for the time of death of an organ, tissue, or whole plant—and plants may be incredibly diverse in this death pattern (Hammond and Adams, 2019). In many species, leaf cells are apparently damaged or killed by dehydration below the turgor loss point, but in resurrection plants, cells can recover completely (Stuart, 1968; Alpert, 2000; Prats and Brodersen, 2021). In deciduous species, the leaves die first, and then the buds on the stem, but in some species, roots apparently die early on and spell the death of the plant (Sack, 2004). Plants with multiple stems and/or sectoriality among stems and roots may better survive drought due to redundancy and/or resprouting after mortality of stems and roots (Schenk et al., 2008; Zeppel et al., 2015; McElrone et al., 2021). Answering the critical question of the location of hydraulic bottlenecks, vulnerabilities, and triggers of mortality within plants, and of the traits with predominant influence across diverse plants, will open the door to the prediction of plant mortality and ecosystem shifts, and the design of drought-hardy crop varieties. These imperatives are as urgent as readying our power grids for the storms of climate change.

How does the continuing rise in CO_2 affect the regulation of stomatal apertures and water-use efficiency of plants?

(By Julian I. Schroeder and Po-Kai Hsu)

The atmospheric CO_2 concentration is continuing to rise and is now ∼50% larger than before the industrial revolution. This is resulting in increased absorption of infrared radiation by CO_2, which in turn is causing temperatures to rise on Earth. Plants remove CO_2 from the atmosphere via photosynthesis. CO_2 can be viewed as an abundant atmospheric fertilizer that contributes to plant growth, if nutrients and water are sufficiently available and depending on the species and conditions (De Kauwe et al., 2021). Indeed, satellite monitoring of photosynthesis and plant growth has shown global "greening" (Zhu et al., 2016a, 2016b; Chen et al., 2022). However, warming temperatures globally can increase vapor pressure deficit, which arguably may tend to counteract this greening trend (Zhu et al., 2016a, 2016b; Wang et al., 2020a, 2020b), with this hypothesis being a matter of debate and further studies being warranted.

CO_2 enters leaves for photosynthesis via stomatal pores. Typically, plants lose ∼150 to >500 water molecules via stomatal transpiration for every CO_2 molecule that is taken in and assimilated by photosynthesis. The CO_2 concentration itself is a regulator of the rapid stomatal closing and opening responses. Elevated CO_2 in the intercellular spaces of leaves, $[CO_2]i$, occurs at night in C3 and C4 plants due to respiration, triggering stomatal closing. During light periods, photosynthesis reduces $[CO_2]i$, which mediates stomatal opening, together with a light-triggered signal transduction network. The atmospheric $[CO_2]$ rise is adding to these diurnal changes in $[CO_2]i$, thereby causing a narrowing of stomatal pores globally (Medlyn et al., 2001; Franks et al., 2013). This CO_2 response can be beneficial to plants. Reduction in stomatal apertures resulting from elevated atmospheric $[CO_2]$ can enable plants to maintain photosynthetic CO_2 assimilation rates, while losing substantially less water, thereby improving water use efficiency (WUE). However, there are limitations, including that many plants have a "weak" stomatal CO_2 response (Raschke, 1975), thereby showing less

or no improvement in WUE. C_4 plants show saturation of assimilation at relatively low $[CO_2]i$ levels and reduction of stomatal conductance may improve WUE depending on the species and conditions (De Kauwe et al., 2021). Moreover, this is also relevant for C_3 plants (Doheny-Adams et al., 2012), including in forestry, where reduction in transpiration from trees could slow depletion of soil water content.

On the other hand, in agricultural regions with sufficient rainfall, soil nutrients, and favorable growth conditions, the high CO_2-induced reduction in stomatal apertures could limit photosynthesis. This applies particularly to C3 plants for which photosynthetic CO_2 assimilation is not yet saturated at typical $[CO_2]i$ levels. C3 plants represent ∼85% of plant species globally. Research is also needed to examine the impact of CO_2-induced stomatal closing during heat stress, given that the CO_2 response appears to be weaker at high temperatures and heat itself is a signal that promotes stomatal opening (Raschke, 1975).

A better understanding of the molecular mechanisms that enable CO_2 control of stomatal movements could aid in future molecular enhanced breeding-, engineering-, and/or gene editing-driven improvement of stomatal WUE traits that are better adapted to diverse environments in a high CO_2 and climate change-challenged world. In recent years, advances have been made toward understanding the molecular mechanisms that cause CO_2-regulated stomatal movements, with some key questions remaining to be resolved as highlighted here.

Studies have shown CO_2 sensing by guard cells, but also a role for the mesophyll (Mott et al., 2008), in sensing or amplifying the stomatal CO_2 response. Forward genetic screens have thus far identified guard cell localized molecular mechanisms that function in CO_2 control of stomatal movements. Thus, the rapid mesophyll-derived signal remains one of the open questions. We discuss guard cell CO_2 signaling mechanisms and open questions in the following (Figure 10). In brief, CO_2 entry into guard cells could be facilitated by the CO_2-permeable PIP2 aquaporins (Mori et al., 2014; for review Zhang et al., 2018a). Carbonic anhydrases in guard cells accelerate the stomatal response to CO_2 shifts in Arabidopsis, rice, and maize (Hu et al., 2010; Chen et al., 2017; Kolbe et al., 2018). Carbonic anhydrases mediate reversible catalysis of CO_2 in guard cells to bicarbonate and protons. The βCA4 isoform of carbonic anhydrases is located at the plasma membrane of guard cells and interacts directly with the PIP2;1 aquaporin (for review, Zhang et al., 2018a). Studies have suggested that intracellular bicarbonate (HCO_3^-) plays an important role as a second messenger in transducing the CO_2 signal in guard cells (e.g. Hu et al., 2010). However, the primary bicarbonate/CO_2 sensor in guard cells that controls stomatal movements has remained elusive (Note that a *secondary* sensor has been identified, as discussed later). Identification of the primary HCO_3^-/CO_2 sensor that controls stomatal movements will be a key to modifying dynamic CO_2-dependent WUE.
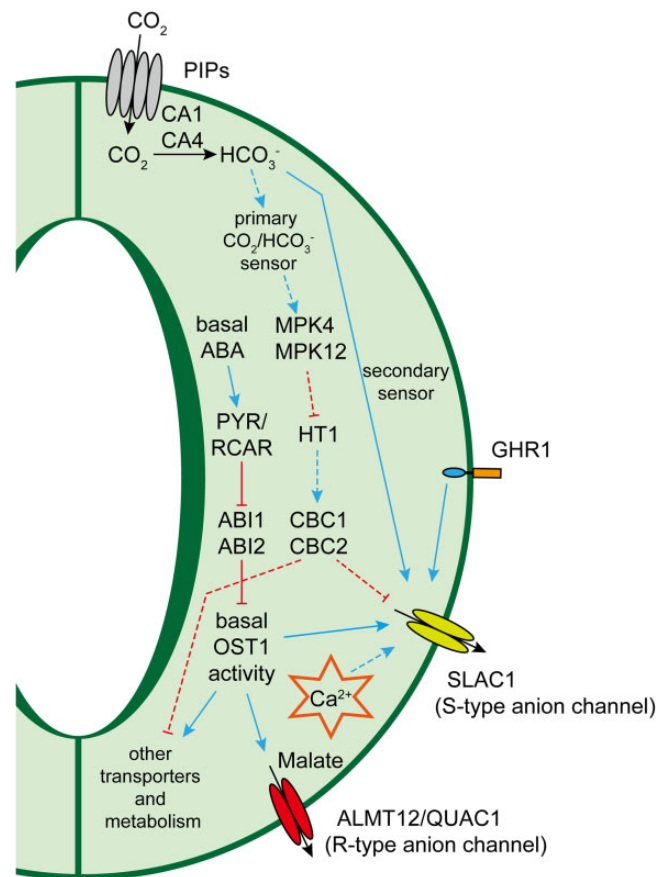

Figure 10 Simplified model for CO_2 sensing and signaling in stomatal closure. Schematic diagram of signaling components and pathways in CO_2-induced stomatal closure. Black arrows indicate the directions of transport or enzymatic reaction in response to elevated CO_2. Blue arrows and red blocks represent positive and negative regulation of high CO_2-triggered stomatal closure, respectively. PIP2 AQPs (PIPs) and β-carbonic anhydrases (CA1/CA4) could facilitate CO_2 influx and HCO_3^- and proton production in guard cells. Protein kinases including MPK4/MPK12, HT1, and CBC1/CBC2 are known early signaling components involved in regulating ion transporters. However, many detailed mechanisms in CO_2 sensing and signal transduction remain to be investigated as indicated by dash arrows and red dashed blocks. Basal ABA signaling and basal OST1/SnRK2 kinase activities facilitate high CO_2-induced stomatal closure. A receptor-like pseudokinase GHR1 is required for SLAC1 activation under CO_2 elevation and several other environmental stimuli.

A forward genetic CO_2 response screen identified recessive *high temperature 1* (*ht1*) mutant alleles in a RAF-like protein kinase that cause a strong insensitivity to low CO_2-induced stomatal opening, while showing responsiveness to blue light and ABA (Hashimoto et al., 2006). Mapping of natural variants in WUE and in stomatal regulation have independently identified the mitogen-activated protein kinase MPK12 as a rate-limiting genetic locus (Des Marais et al., 2014; Jakobson et al., 2016). Mechanistic research has shown that in Arabidopsis double mutants of *mpk12* together with the close homolog *mpk4* and in tobacco silencing of the close homolog *NtMPK4* disrupt high CO_2-induced stomatal

closing, while ABA-induced stomatal closing remains intact (Marten et al., 2008; Tõldsepp et al., 2018). Neither MPK12 nor MPK4 protein kinase activities were found to be regulated by CO_2/HCO_3^- directly in vitro under several examined conditions (Tõldsepp et al., 2018).

Two other RAF-like protein kinases were discovered, CONVERGENCE OF BLUE LIGHT AND CO_2 (CBC1 and CBC2), for which double mutants show an impairment in low CO_2-induced stomatal opening (Hiyama et al., 2017). In contrast to recessive *ht1* mutant alleles (Hashimoto et al., 2006), *cbc1/cbc2* double mutants disrupt blue light-induced stomatal opening as well (Hiyama et al., 2017). Therefore, CBC1 and CBC2 are proposed to represent a convergence point of low CO_2 and blue light-mediated stomatal opening (Hiyama et al., 2017). HT1 can phosphorylate CBC1 in vitro (Hiyama et al., 2017), but the physiological relevance of this reaction for CO_2-mediated stomatal regulation is unknown.

The carbonic anhydrases (βCAs), HT1, and MPK12/MPK4 proteins function in the early guard cell specific CO_2 response pathway as positive regulators (βCAs, MPK12/MPK4) and negative regulators (HT1) (Figure 10). Downstream of early CO_2 signaling, a network of guard cell ion channels, pumps, transporters, and regulators in the plasma membrane and vacuolar membrane as well as metabolic responses (e.g. Flütsch et al., 2020) mediate CO_2-regulated turgor driven stomatal movements. Elevated CO_2 activates both slow "S-type" and rapid "R-type" anion channels in guard cells (Raschke et al., 2003). Elevated CO_2 can be predicted to inhibit plasma membrane proton pumps that drive stomatal opening, although direct evidence is needed. Furthermore, elevated bicarbonate enhances the activity of the S-type anion channel SLAC1 in heterologous cells and in guard cells. Residues in SLAC1 that are required for this response have been identified and the respective SLAC1 mutants show impaired CO_2 regulation, but intact ABA regulation, of stomatal closing in intact plants, leading to the model that SLAC1 can function as a secondary CO_2/HCO_3^- sensor in guard cells (Zhang et al., 2018b). Since SLAC1 activation is known to require phosphorylation, the upstream primary CO_2/HCO_3^- sensor remains, however, unknown (Zhang et al., 2018b).

An important question remains on how early CO_2 signaling mechanisms control these mediators of stomatal movements. Research suggested that the elevated CO_2 response is mediated by the ABA receptor signaling pathway (Dittrich et al., 2019). However, CO_2-regulated stomatal conductance findings in ABA receptor mutants (Dittrich et al., 2019) could not be confirmed using several approaches and showed CO_2 responsiveness (e.g. Hsu et al., 2018; Zhang et al., 2020). Moreover, recent research has led to the unexpected findings that CO_2 triggers stomatal closing without further activating SNF1-related protein kinase2 (SnRK2s), including OST1 (Hsu et al., 2018; Zhang et al., 2020), that are activated by ABA. Moreover, basal ABA levels and a basal activity of SnRK2/OST1 protein kinases were found in guard cells, and these are required for amplifying the CO_2 response

even though CO_2 increases did not rapidly change their activities (Hsu et al., 2018; Zhang et al., 2020). Thus, the link from early CO_2 signaling mechanisms to downstream targets that mediate stomatal closure remains to be discovered.

There is still much to learn about how CO_2 regulates stomatal apertures, from CO_2/HCO_3^- sensors to a biochemical and physiological understanding of the signaling network, which could drive future improvements in WUE of plants including trees, depending on the species, with a need for future quantitative field research (De Kauwe et al., 2021). Furthermore, leaf-level stomatal conductance models are a crucial part of Earth system climate models, and molecular insights could improve the accuracy of these models.

How can one aquaporin have so many roles in a plant?

(By Stephen D. Tyerman)

Much research on plant AQPs has assessed their impact on water transport across membranes in response to drought and salinity stress, but low temperature, anoxia, and nutrient stress and combinations also feature (Kapilan et al., 2018). Many studies show that overexpression of an AQP can confer tolerance to these stresses, sometimes multiple stresses, but it is by no means clear how such stress tolerance occurs in the strict context of water transport. Some AQPs, originally designated as true water channels, have been shown to transport multiple substrates besides water, including signaling molecules (e.g. H_2O_2), neutral substrates for synthesis (CO_2, O_2, and NH_3), and even ions (Na^+, K^+, and NO_3^-) (Tyerman et al., 2021) that can all feature in responses to abiotic stress or photosynthesis (Ermakova et al., 2021). AQPs are also under the control of many hormones (Maurel et al., 2021) and are important for regulating growth (Wang et al., 2020a, 2020b). However, genetic evidence indicating AQP involvement in stress resistance, for example, the location of AQP genes under stress-related QTLs, is uncommon (Hostetler et al., 2021a), perhaps due to their complex regulation (Grondin et al., 2016) and multifunctionality.

The plant AQPs can be divided into several subfamilies (Figure 11A) and not all of them are good water channels. Within the NIPs (mainly), as well as some PIPs and XIPs, transported substrates include metaloids, protonated organic acids, or metal complexes (Tyerman et al., 2021). Water can also be transported but not always. In each case, the transport is passive (down-hill) in response to the free energy gradient for water, concentration gradients for the solute, or electrochemical gradients for those shown to also pass ions (Tomkins et al., 2021; Tyerman et al., 2021). Multiple substrate transport through the same protein at the same time (e.g. ions and water, CO_2 and water, and H_2O_2 and water) could lead to interactions (Tyerman et al., 2021). It remains to be seen how some substrates permeate and the fifth pore through the center of the tetramer is a candidate for ions and CO_2 (Figure 11, B–E).

What may still occur, even for passive transport, is rectification (i.e. a greater flow or diffusion in one direction
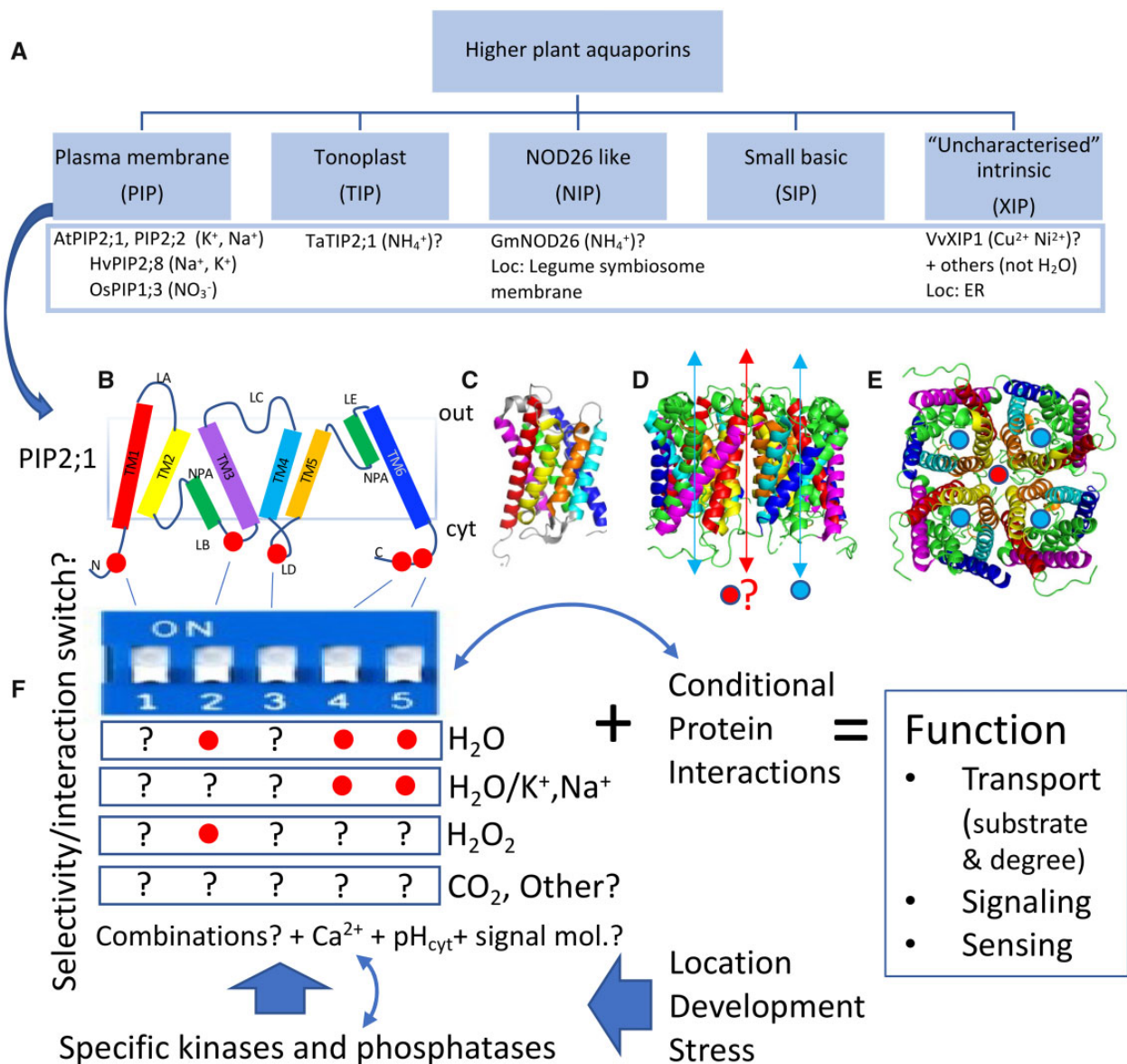
Figure 11 How an aquaporin (AQP) can be multifunctional. A, AQP family with those indicated to transport ions electrogenically (under each subfamily, ? = maybe, Loc = membrane location for specific isoform, note many NIPs are located on the plasma membrane) (Tyerman et al., 2021). B, Features of PIP2;1 showing transmembrane helices (TM1-6), loops (A–E) and half helices forming the central selectivity NPA filter. Red dots = phosphorylation sites. C, Folded monomer structure (SoPIP2;1 PDB 1z98) (Törnroth-Horsefield et al., 2006). D, Homotetramer as the functional unit with four monomeric pores that transport water (blue arrows, dots) and probably H_2O_2, also NH_3 in TIP2;1 (Kirscht et al., 2016) and the central pore (red arrow, dot) implicated in ion transport for mammalian AQP1 (Henderson et al., 2022). AQPs also likely form heterotetramers (e.g. ZmPIP1;2 + ZmPIP2;1) (Chaumont and Tyerman, 2014). E, View normal to the membrane plane on the cytoplasmic face. F, Hypothesis for multi-functionality of PIP2;1. Phosphorylation by specific kinases occurs at one or more of the sites on cytoplasmic loops like a digital dipswitch that determine protein interactions and/or substrate selectivity, for example, ion water reciprocity (Qiu et al., 2020) or H_2O_2 permeation (Rodrigues et al., 2017), or protein interactions (Prado et al., 2019). Selectivity is fixed to a certain degree in the monomeric pores by the structure of the ar/R selectivity filter and other residues. The hydrophobic central pore may still allow ion permeation and is proposed to be gated at a low probability dependent on monomer gating (Tyerman et al., 2021). There are likely feedbacks (small blue arrows) due to some substrates and protein interactions also affecting the signaling that determines the kinase/phosphatase activity.

compared with the other depending on the direction and magnitude of the gradient) as is well known for some ion channels. The aromatic/Arginine (ar/R) selectivity filter present on the lumen side of the monomeric pore may give rise to voltage and ion dependence (Mom et al., 2021) that could lead to rectification. Rectification has not been well studied for plant AQPs at the molecular level though it was well studied in the past for water transport across plant cell membranes. This could occur for water to be trapped within the root xylem (Pascut et al., 2021) but reverse flow through a root with reversed gradients would argue against this. Ion flow through AtPIP2;1 and AtPIP2;2 can show rectification

with certain divalent cations (Ca^{2+} and Mg^{2+}) present (Kourghi et al., 2017).

Some PIP AQPs can account for significant portions of shoot and root hydraulic conductivity (Lpr) (Prado et al., 2019; Ding et al., 2020; Domec et al., 2021). To determine the contribution of AQPs to Lpr requires a combination of sophisticated models and measurements of water transport (Ding et al., 2021; Knipfer et al., 2021). The regulation of root AQPs in response to abiotic stress such as salinity and anoxia via gating or removal from the membrane results in rapid changes in Lpr (Tournaire-Roux et al., 2003; Boursiac et al., 2008). Shoot signals are also implicated in regulation of root AQPs (Chaumont and Tyerman, 2014). Changes in the Lpr can have large effects on stomatal conductance, shoot water relations, and growth (Ding et al., 2020; Knipfer et al., 2021) and perhaps ion content of the shoot related to the reflection coefficient of the root (Knipfer et al., 2021) or the capacity of some AQPs to transport ions. Ultimately, the amount of water relative to the amount of ions transported to the xylem determines the xylem ion concentration for delivery to the shoot. A common signal element that links plant Na^+/K^+ ratios under salinity with control of AQPs in the root is the production and transport of H_2O_2 (Ma et al., 2011; Martiniere et al., 2019).

Taking the PIP subfamily (with two clades; PIP1 and PIP2) as the best studied example of multifunctionality, one PIP2 isoform can be permeable to water, cations, CO_2, and the signaling molecule H_2O_2. For the cases of cations, water, and H_2O_2, phosphorylation on certain residues appears to be key (Figure 11) though there are some variations in the literature regarding the impact on water flow. Arabidopsis AtPIP2;1 has been shown to be required for circadian variation in rosette Lp with 14-3-3 proteins depending on C-term phosphorylation (Prado et al., 2019), for auxin regulation of lateral root outgrowth (Peret et al., 2012), CO_2 uptake into guard cells with a carbonic anhydrase (Wang et al., 2016), H_2O_2 signaling in guard cells dependent on a Loop B phosphorylation (Rodrigues et al., 2017), and univalent cation transport dependent on C-term phosphorylation (Qiu et al., 2020). The very similar AtPIP2;2, also permeable to cations but with increased sensitivity to Ca^{2+} compared with AtPIP2;1 (Kourghi et al., 2017), contributes to Lpr derived from root exudation (Lp^{rex}) but not Lpr derived from pressure gradients (Lpr^{pres}) (Javot et al., 2003). Root exudation is important for xylem water continuity and may be more complicated than ion pumping into the xylem with the subsequent osmotic gradient driving flow (Schenk et al., 2021). Interestingly, AtPIP1;2 when knocked out reduces Lpr^{pres} but not Lp^{rex} pointing to an interesting division of labor that may depend on different locations of the two AQPs (Postaire et al., 2010). Another PIP2 from rose is implicated in a drought signaling hub releasing a membrane bound transcription factor depending on phosphorylation status and environmental signals to control growth under water stress (Zhang et al., 2019). The well-studied maize PIP2;5 is also multifunctional being implicated in guard cell

signaling for ABA responses due to H_2O_2 permeation (Ding et al., 2021) but also for its water permeation in the root (Ding et al., 2020).

Many introductions to papers on AQPs describe them as exclusively important for water transport, implying that there is only water transport. But the situation now emerging is far more complex and the PIPs seem to be implicated in various aspects of plant water relations that are more than just water transport across a membrane. This makes interpretation of phenotypes in both reverse and forward genetics rather challenging. The question as to how a single AQP can function in such a broad range of cell types with different transported substrates or signaling roles is addressed by the hypothesis in Figure 11F. The answer may lie in a digital-like switch through posttranslational modifications and signaling pathways combined with many and varied protein interactions.

How do plants balance growth and abiotic stress responses?

(By Taishi Umezawa)

Plant growth is affected by many environmental factors that alter the balance of energy use (Crepin and Rolland, 2019). Under favorable environmental conditions, plants are able to assign the energy obtained from photosynthesis to growth, especially during the vegetative growth phase. Conversely, under adverse environmental conditions, plants must redirect their energy to stress responses to overcome the challenge and ensure individual survival. However, when stress conditions are prolonged, it is not always a suitable strategy to inhibit growth indefinitely. Under such conditions, it may be advantageous for plants to maintain growth to some extent, or to switch to their reproductive stage to preserve the next generation. It is likely that plants have evolved to adjust their energy balance precisely in response to abiotic stresses, because they are often forced to make such decisions throughout their life cycles. The question is how plants tilt the balance toward stress responses or toward growth regulation (Figure 12).

In the case of drought, plant responses to this stress have been divided into three alternative strategies: drought tolerance, drought avoidance, and drought escape (Kooyers, 2015). For short-term drought, drought avoidance or tolerance can be effective. For example, it is well known that plants can quickly close their stomata to prevent water loss from the leaf surface. Similarly, plant cells can rapidly adjust their osmotic potential to maintain water status (Zhu, 2002; Yamaguchi-Shinozaki and Shinozaki, 2006). Molecular mechanisms that turn on such stress responses have been well studied, especially responses induced by the phytohormone ABA. The major ABA signaling pathway consists of three core components: ABA receptors (PYL/RCAR), Clade A of the type 2C protein phosphatases (PP2C), and SnRK2 kinases (Cutler et al., 2010; Umezawa et al., 2010). Under normal conditions, PP2Cs directly dephosphorylate and inactivate SnRK2s (Umezawa et al., 2009; Vlad et al., 2009).
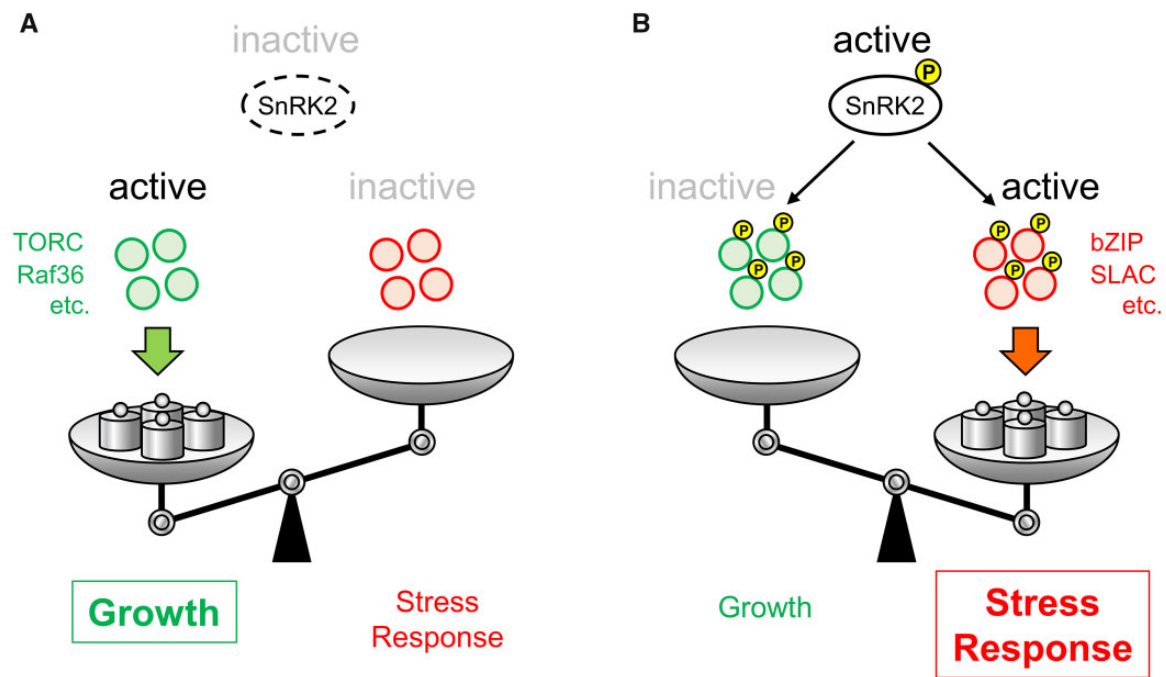
Figure 12 SnRK2 kinases regulate growth and stress response under drought. A, Under normal conditions, plants use energy to grow, and TORC or Raf36 are involved in this process. B, Under drought conditions, SnRK2s are activated and phosphorylate substrates, for example, bZIP transcription factors or slow anion channels (SLAC), to induce stress responses. SnRK2s also phosphorylates TORC or Raf36 to inhibit plant growth.

In response to ABA, this inhibition is abated and active SnRK2s can phosphorylate various protein substrates to induce ABA-associated responses including stomatal closure and large-scale changes in gene expression. Since SnRK2s are central players in drought responses, many studies have used SnRK2s as a starting point to identify signaling proteins involved in ABA or osmotic stress signaling (Wang et al., 2018a; Kamiyama et al., 2021).

In addition to their central role in ABA signaling, SnRK2s also function to regulate plant growth under drought stress conditions. Recently, SnRK2s were shown to directly phosphorylate Raptor, a component of the TOR complex (TORC) that regulates plant growth (Figure 12). Under stress conditions, phosphorylation of Raptor by SnRK2 resulted in dissociation of TORC and inhibition of plant growth (Wang et al., 2018a, 2018b). Separately, a recent study demonstrated that Raf36, a group C Raf-like protein kinase, promotes growth under normal conditions, and is degraded in response to ABA by SnRK2-dependent phosphorylation (Kamiyama et al., 2021). These two examples highlight an important role for SnRK2s not only for ABA-dependent stress responses, but also for mediating growth inhibition under short-term and severe drought stress. In addition to the SnRK2 pathway, Clade E Growth-Regulating (EGR) phosphatases and Microtubule-Associated Stress Protein 1 are involved in growth regulation during drought stress (Longkumer et al., 2022). It is expected that identifying and functionally characterizing additional SnRK2 or EGR substrates will aid our understanding of the mechanisms of growth inhibition under drought stress.

In nature, sudden and severe drought stress on plants is not likely to occur. In most cases, drought stress gradually increases in stages. When drought stress is mild and prolonged, it is likely not beneficial for plants to spend energy only on stress responses, and under such conditions, plants may continue to grow as part of their drought avoidance or escape strategies (Kooyers, 2015). For example, root growth often increases during mild drought as a means to increase access to available water. In rice, a root angle QTL, DRO1, was shown to be effective for selection of drought tolerance in rice, demonstrating that drought avoidance is one of the promising breeding targets for drought resistance (Uga et al., 2013).

In the drought escape response, plants accelerate floral development and the transition to the next generation (Kooyers, 2015). ABA is involved in early flowering as a drought escape response, and multiple pathways are proposed to link ABA and flowering (Martignago et al., 2020). For instance, several bZIP transcription factors, AREB/ABFs, are phosphorylated by SnRK2s and involved in drought escape (Hwang et al., 2019). In addition, previous studies proposed that the photoperiodic flowering pathway, consisting of GIGANTEA (GI), CONSTANS, and FT, is essential for early flowering (Riboni et al., 2013, 2016). However, the connection between ABA and GI-dependent FT pathway is still under investigation.

Drought stress is not constant and stress intensity fluctuates over time in nature. Once drought stress reaches a certain level, plants cross a threshold and change the balance between growth and stress response. It will be important to identify the molecular switch involved in such a stage-gate

of drought stress. Furthermore, if the intensity of drought stress changes frequently, plants can acquire stress memory. It is known that some epigenetic modifications could be involved in stress memory (Sharma et al., 2022) and may regulate intra- or inter-generational responses to stress conditions. This is another topic to be clarified.

As discussed in this section, plant growth regulation is complex, and current knowledge of plant growth regulation under stress conditions is just beginning to scratch the surface. Further studies will be required to understand how plants balance stress response and growth regulation, and in depth understanding of such mechanisms could facilitate molecular breeding for yield and quality of agricultural production under drought conditions.

Proline metabolism: protector, scavenger, or executioner?

(By Paul E. Verslues)

Proline is highly soluble and zwitterionic, hallmarks of compatible solutes that accumulate to reduce cellular osmotic potential while also protecting protein and membrane structure from dehydration-induced damage. However, making proline is not the only impact of proline metabolism and the protective role of proline itself is not the only purpose of stress-induced proline accumulation (Bhaskara et al., 2015; Alvarez et al., 2022). The proline cycle (Figure 13) consists of synthesis from glutamate by Δ^1-pyrroline-5-carboxylate (P5C) synthetase (P5CS) and P5C reductase (P5CR) while proline catabolism back to glutamate is catalyzed by proline dehydrogenase (ProDH) and P5C dehydrogenase (P5CDH). In Arabidopsis, *P5CS1* and *ProDH1* are the most stress-responsive of the proline cycle genes and the proteins they encode catalyze the rate-limiting steps of proline synthesis and catabolism, respectively. One protective function of the proline cycle is to regenerate $NADP^+$ to help ensure the supply of a safe electron acceptors during stress and when leaf CO_2 becomes limited (Hebbelmann et al., 2011; Sharma et al., 2011; Signorelli, 2016). Both P5CS1 and P5CR prefer NADPH over NADH as an electron donor (Giberti et al., 2014; Forlani et al., 2015; Sabbioni et al., 2021) and *p5cs1* mutants have a reduced $NADP^+$/NADPH ratio during low water potential (ψ_w) stress (Sharma et al., 2011). *p5cs1-4* also has substantial changes in photosynthesis-related gene expression (Shinde et al., 2016). How such a proline synthesis-photosynthesis redox link could work depends on the subcellular localization of P5CS1 and P5CR and how it is affected by stress. P5CS1 is likely to be localized in the cytoplasm; however, this is ambiguous as both cytoplasmic (Funck et al., 2020) and chloroplast, or chloroplast-associated, localization (Székely et al., 2008) have been reported. Similarly, fluorescently tagged P5CR was localized mainly in the cytoplasm (Funck et al., 2012) while biochemical or immunoblot assays indicated that it could also be present in plastids (Szoke et al., 1992; Murahama et al., 2001). The preference of P5CS1 and P5CR for NADPH is also consistent with the proposal that proline synthesis is linked

to the pentose-phosphate pathway in a redox cycle (Hare and Cress, 1997). However, this proposal has been little tested.

ProDH1 and P5CDH are also scavengers in that plants can use proline as an alternative respiratory substrate during senescence and dark-induced starvation (Zhang and Becker, 2015). This is facilitated by the fact that ProDH transfers reductant directly to ubiquinone via its FADH cofactor (Zheng et al., 2021). During stress recovery (after restoration of water supply), when proline levels rapidly decline, or in response to exogenous proline, *ProDH1* and *P5CDH* expression is induced (Figure 13) and proline catabolism can feed so much reductant into mitochondrial electron transport that some of it needs to be vented off by alternative oxidase to prevent oxidative stress (Oh et al., 2022). The rapid catabolism of proline after re-watering may be a way to channel the nitrogen from proline to other amino acids needed during the resumption of growth.

Interestingly, *p5cs1* mutants, which have greatly reduced proline accumulation, and *prodh1* mutants, which have increased proline accumulation, have similar low ψ_w-sensitive phenotypes (Sharma et al., 2011; Bhaskara et al., 2015). Low ψ_w stress leads to *ProDH1* downregulation in most of the plant tissues. However, meristematic and growing cells have steady or increased *ProDH1* and *P5CDH* expression during low ψ_w stress (Sharma et al., 2011). This indicates that the proline cycle, with the synthesis versus catabolism sides of the cycle spatially separated, can also facilitate the movement of reducing potential, stored in the form of proline, away from photosynthetic tissue where it is in excess, to root and meristem tissue where proline is used for energy metabolism or osmotic adjustment (or in other words: to be a good protector, it is important to know when to also be a scavenger).

Proline metabolism shows its executioner side during the hypersensitive response (HR) to incompatible pathogen infection (Figure 13). Proline accumulation mediated by P5CS2 and proline catabolism by ProDH1 and ProDH2 is required for HR-induced cell death and associated ROS burst (Fabro et al., 2004, 2016; Cecchini et al., 2011; Senthil-Kumar and Mysore, 2012). During infection, cells marked for death have up-regulated expression of *P5CS2*, *P5CR*, and *ProDH1* but *P5CDH*, leading to partial proline catabolism that is associated with cell death (Alvarez et al., 2022; Figure 13). Whether the cell death is caused by proline-dependent ROS production or a yet unknown signaling function of P5C (or a combination of the two) is unclear. P5C is the common intermediate of both proline synthesis and catabolism and it has also been proposed that P5C may be exported from the mitochondria and used for proline synthesis in the cytoplasm, thus forming a proline–P5C cycle which could amplify ProDH-dependent ROS production in the mitochondria (Miller et al., 2009). However, evidence supporting such a P5C cycle in plants is circumstantial and a mitochondrial P5C translocator has not been identified.
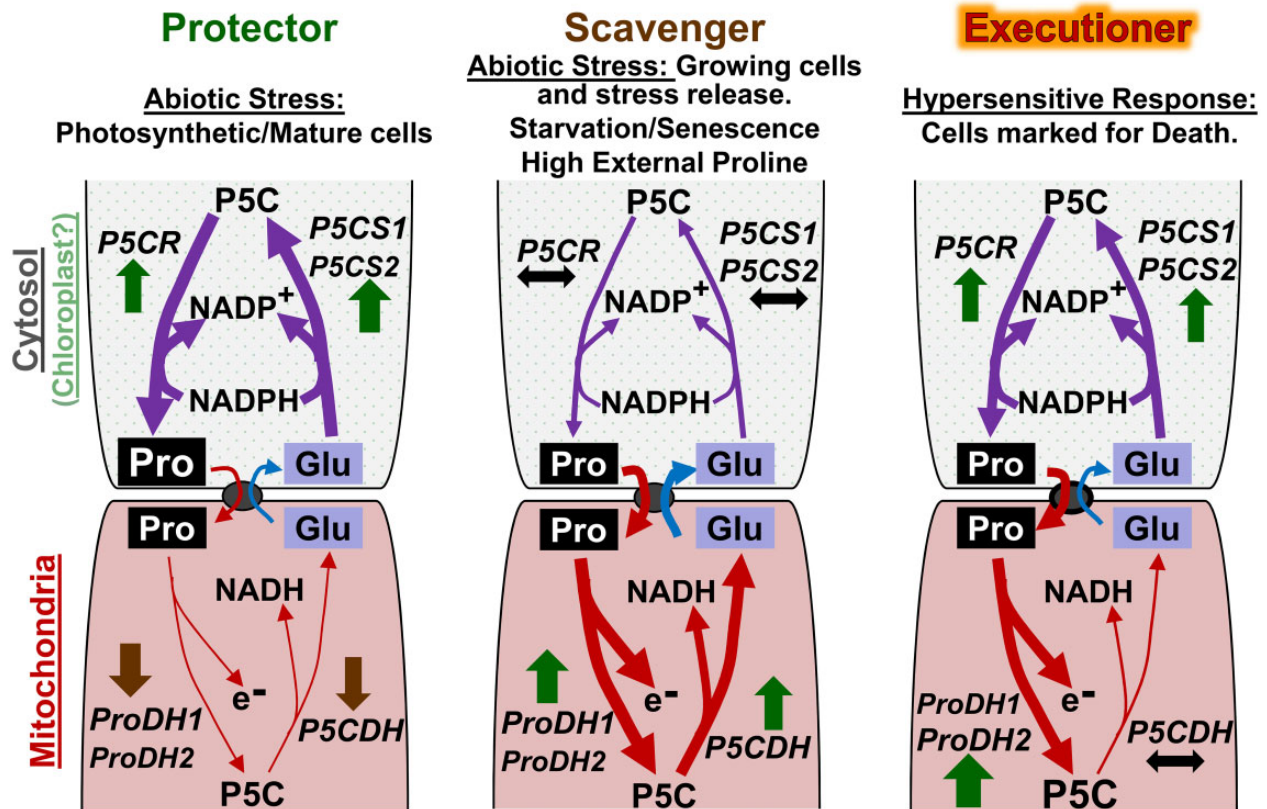
Figure 13 The protector, scavenger and executioner modes of proline metabolism and the conditions or tissue in which they are observed. Up and down arrows (red, brown, or gray) indicate changes in gene expression compared with unstressed control. In most cases where data are available, protein levels of the corresponding enzyme change in the same direction. Thickness of the red or purple arrows represents predicted relative metabolic flux through different steps in proline metabolism. Abiotic stress refers to drought, freezing or high salt treatments where sustained accumulation of free proline is observed in many plant species.

Whether proline metabolism operates in protector, scavenger, or executioner mode depends on unknown environmental and metabolic signals. Thus, proline metabolism is not only interesting in its own right in terms of how it protects or kills plant cells, it is also a useful model to discover new aspects of stress signaling. For example, what sensing and signaling events occur during drought stress to allow high levels of proline to accumulate without inducing *ProDH1* and without having proline metabolism switch into executioner mode to promote cell death (Miller et al., 2005)? This is especially interesting as the sensing and upstream signaling mechanism(s) plants use to detect and respond to reduced water availability during drought stress remain unknown. For proline metabolism, the relative fluxes through different parts of the proline cycle (indicated by different arrow thicknesses in Figure 13) are inferred from gene expression or protein levels of proline metabolism enzymes but there is little information on actual metabolic flux rates through the proline cycle under different conditions. This is important information for determining the conditions where proline catabolism is rapid enough to either significantly contribute to respiration (scavenger mode) or significantly increase ROS levels (executioner mode) and how this is coordinated with mitochondrial mechanisms to dissipate

excess reducing potential, including alternative oxidases and uncoupling proteins. Posttranslational modifications of P5CS1 and ProDH1 (Alvarez et al., 2022) or interactions with regulatory proteins (Ren et al., 2018) are likely to affect their enzymatic properties but the roles of such factors in controlling the protector–scavenger–executioner modes of proline metabolism are unknown. For P5CS1 and P5CR, knowledge of their subcellular localization is also strikingly limited. Surprisingly, Savoure and co-workers have reported that a *prodh1prodh2* double mutant, in which the only two ProDH genes in the Arabidopsis genome are knocked out, is viable despite having no known way to catabolize proline (Alvarez et al., 2022). Is there a metabolic work around that allows these plants to metabolize proline? And, Figure 13 depicts a mitochondrial proline–glutamate exchanger and such an activity, along with that of mitochondrial proline importer(s), has been biochemically observed (Di Martino et al., 2006); however, the genes encoding these activities remain unknown.

Perhaps one of the most striking paradoxes of proline and stress, given all the evidence of the importance of proline to stress resistance, is that some plants apparently do without it. For example, most Arabidopsis accessions accumulate high levels of proline during low ψ_w stress; but a few have

very low levels of P5CS1 and greatly reduced levels of proline accumulation similar to *p5cs1* knockout mutants (Kesari et al., 2012). Are these accessions more sensitive to drought (or salt) stress, or have they found a substitute for the stress-protective (and executioner) effects of proline metabolism? Also, some plants adapted to chronically dry conditions have relatively low levels of free proline accumulation but instead convert proline to proline betaine or hydroxy-proline betaine as these may be more effective osmoprotectants (Hanson et al., 1994). However, the implications of this conversion for the proline cycle are unknown and these compounds are likely to be more difficult to catabolize, thus impeding redeployment of nitrogen and reducing potential when the stress subsides.

Is proline metabolism a protector, scavenger, or executioner? It depends. Depends on what is the real question; a question whose answer will reveal much about the sensing, signaling, and metabolic mechanisms that plants use to cope with abiotic stresses that are of increasing concern for a warming and changing world.

Temperature sensing: how do plants adapt to different climates?

(By Philip A. Wigge)

A remarkable feature of plants is their ability to adapt to a wide range of climates, occupying almost every niche from the tropics and hot springs to polar regions. To do this, plants have evolved an array of responses to temperature, over multiple scales, from minutes to months, which enable a suite of developmental and cell biological responses to maximize survival. Understanding how plants are able to adapt to different climates is a major open question, and of particular relevance during a period of unprecedented rapid global heating (Battisti and Naylor, 2009).

Broadly, we can consider active and passive responses to temperature. Passive responses refer to adaptations such as membrane fluidity and protein stability. Proteins at high temperature tend to denature and unfold. Thermophiles therefore have proteins with increased ionic interactions and a larger stable hydrophobic core. At low temperature, there is reduced molecular motion due to low entropy and enthalpy, and psychrophilic organisms adapt by having proteins with fewer salt bridges and hydrogen bonds to facilitate flexibility (Brininger et al., 2018).

A major strategy of plants has been to evolve active temperature sensing and response pathways. These enable the anticipation of both seasonal temperature shifts as well as shorter term temperature stresses. Temperature measurements over the year, in concert with photoperiod, enable plants to avoid unfavorable seasons in a dormant stage, and grow and flower during suitable months. While there is enormous diversity in the habitats and climates that plants have adapted to, conservation of major signaling components appears to be a common theme.

Broadly, two major approaches have been followed to identify the genes and mechanisms underlying adaptation to different temperatures. From a population perspective, it is possible to harness the power of genetics to identify genomic regions and loci that show signatures of natural selection in populations from different locations (Hancock et al., 2011). Another strategy is to identify the underlying temperature sensors in genetic screens or using a candidate gene approach. This method is complicated by the often pleiotropic and redundant nature of temperature signaling pathways, reflecting their central role in many essential responses. The use of carefully designed and controlled temperature screens can, however, overcome some of these issues. Sensors may then be investigated for natural variation in the context of their mode of action. This approach has the advantage that it is targeted and enables a test of functionality, as well as identification of variation which directly impacts temperature signaling. We will discuss some well-studied temperature responsive networks that suggest pathways to adaptation.

Vernalization

Vernalization, the response of plants to prolonged cold, is a classical temperature response in Arabidopsis. Extensive natural variation exists at the level of the major regulators *FLC* and *FRI* (Shindo et al., 2005; Figure 14). The complex nature of *FLC* silencing in response to cold via a proposed antisense RNA also lends itself to natural variation and modulation in terms of the extent and duration of cold requirement (Duncan et al., 2015).

Thermomorphogenesis

Accelerated growth in response to warm temperature and flowering in Arabidopsis is termed thermomorphogenesis. This process is dependent on enhanced activity of PIF4 (Quint et al., 2016). PIF4 is regulated posttranslationally by the thermosensor phytochrome B (phyB). phyB measures temperature through its dark reversion rate (Jung et al., 2016a; Legris et al., 2016). Different rates of dark reversion can evolve orthogonally to light sensing and cause a corresponding change in temperature sensitivity. This could enable thermorphogenesis to be tuned to the local environment. In addition to the identification of phyB in a natural variation screen for thermal responsiveness, the gene *EARLY FLOWERING3* (*ELF3*) was also identified (Box et al., 2014; Figure 14). ELF3 contains a thermoresponsive predicted prion domain (Jung et al., 2020). This has been shown to have extensive natural variation in the length of a polyQ repeat (Undurraga et al., 2012), and variation both within Arabidopsis as well as with other plants such as *Brachypodium distachyon* directly changes temperature responsive behavior. The polyQ region is encoded by a short tandem repeat (STR), which can expand or contract during replication through DNA polymerase slippage, enabling the generation of functional variation in ELF3 (Kashi and King, 2006). Since the thermal responsiveness of the protein is proportional to the size of the repeat, this may allow for adaptation to different temperature ranges. It will be interesting to see if this STR-based mechanism occurs in other
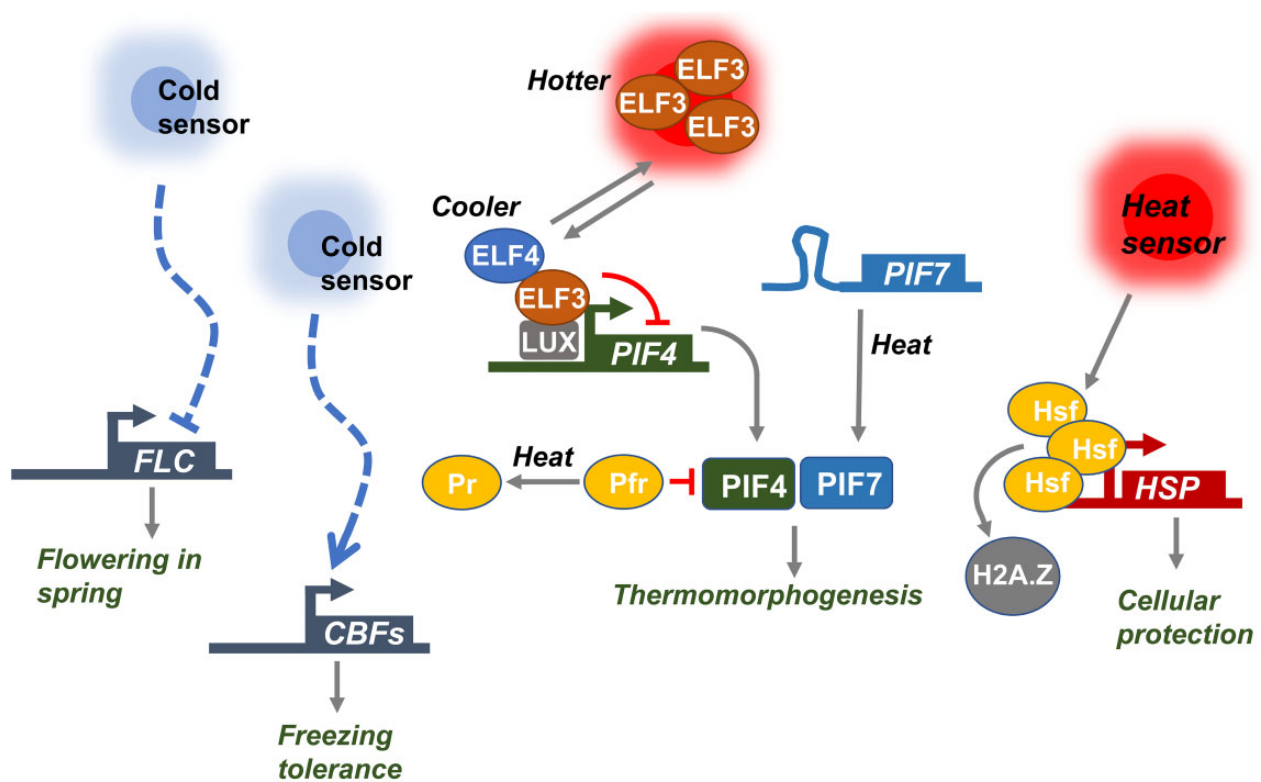
Figure 14 Temperature response mechanisms. Temperature controls many key adaptive traits in plants such as freezing tolerance, growth, flowering, and heat stress protection. Vernalization results in the stable repression of expression of the floral repressor *FLC*. Thermomorphogenesis integrates temperature information from at least three distinct biophysical responses: protein phase separation of ELF3, thermal reversion of phyB (Pfr to Pr) and enhanced translation of *PIF7* in warm conditions. Heat stress also induces the induction of protective heat shock proteins via the activation of HSFs via an unknown mechanism.

temperature-responsive proteins, as has been suggested for Drosophila (Sawyer et al., 1997). An additional thermosensory mechanism is displayed by the RNA secondary structure in the 5′ UTR of *PIF7*, which facilitates enhanced translation at higher temperatures (Chung et al., 2020). Since 5′ UTR sequences can evolve independently of protein function, this represents a mechanism to alter the levels of protein rapidly in response to temperature. It is not known if this is a widespread mechanism in plants.

Temperature stress

Survival of freezing stress is mediated by the CBF transcription factors in Arabidopsis, which are activated by cold perception (Jaglo-Ottosen et al., 1998). Heat stress responses are activated by the conserved heat shock factors (HSFs), which activate protective heat shock proteins. In plants, HSFs have undergone a remarkable expansion from 1 to 3 family members in yeast and mammals to 21 members in Arabidopsis (Baniwal et al., 2004). The basis for this is not clear, but suggests an important role for this family of TFs in mediating plant survival (Figure 14).

Critical to understanding adaptation to climate will be determining how many temperature sensors are present in plants. The very distinct genetic and physiological responses to cold during vernalization and cold stress suggest independent sensors, while the heat stress is similarly independent from thermomorphogenesis. Nevertheless, it is plausible to propose as few as 10–20 distinct temperature sensors may account for most of the transcriptional responses to warm and cold temperatures observed in Arabidopsis. Thermomorphogenesis is perhaps the most well-studied system, and in this case, it is interesting that multiple discrete sensors act at different scales and levels (transcriptional, translation, and posttranslation). Such redundancy may represent a mechanism to filter the inherent noise from temperature signals. How temperature information is remembered and integrated over many months is also not understood. Heat stress in the field occurs in a complex environmental context, often with drought stress, and therefore how these various stresses are integrated is important. Analysis of natural variation and field studies of a broader range of plants will also be critical for understanding mechanisms by which sequence variation can achieve rapid changes in the temperature response range of thermosensors. This knowledge will enable the rational editing of crops to enhance thermal resilience.

Acknowledgments

References

1001 Genomes Consortium (2016) 1,135 Genomes reveal the global pattern of polymorphism in *Arabidopsis thaliana*. Cell **166**: 481–491

Abdalla M, Carminati A, Cai GC, Javaux M, Ahmed MA (2021) Stomatal closure of tomato under drought is driven by an increase in soil–root hydraulic resistance. Plant Cell Environ **44**: 425–431

Agrawal AA (2020) A scale-dependent framework for trade-offs, syndromes, and specialization in organismal biology. Ecology **101**: e02924

Agrawal AA, Conner JK, Rasmann S (2010) Tradeoffs and negative correlations in evolutionary ecology. *In* Evolution Since Darwin: The First 150 Years. Sinauer Association, Sunderland, MA, pp 243–268.

Abiko T, Kotula L, Shiono K, Malik AI, Colmer TD, Nakazono M (2012) Enhanced formation of aerenchyma and induction of a barrier to radial oxygen loss in adventitious roots of *Zea nicaraguensis* contribute to its waterlogging tolerance as compared with maize (*Zea mays* ssp. mays). Plant Cell Environ **35**: 1618–1630

Albuquerque C, Scoffoni C, Brodersen CR, Buckley TN, Sack L, McElrone AJ (2020) Coordinated decline of leaf hydraulic and stomatal conductances under drought is not linked to leaf xylem embolism for different grapevine cultivars. J Exp Bot **71**: 7286–7300

Alejo-Jacuinde G, Kean-Galeno T, Martínez-Gallardo N, Tejero-Díez JD, Mehltreter K, Délano-Frier JP, Oliver MJ, Simpson J, Herrera-Estrella L (2022) Viability markers for determination of desiccation tolerance and critical stages during dehydration in *Selaginella* species. J Exp Bot **73**: 3898–3912

Alpert P (2000) The discovery, scope, and puzzle of desiccation tolerance in plants. Plant Ecol **151**: 5–17

Alvarez ME, Savoure A, Szabados L (2022) Proline metabolism as regulatory hub. Trends Plant Sci **27**: 39–55

Anderson JT, Willis JH, Mitchell-Olds T (2011) Evolutionary genetics of plant adaptation. Trends Genet **27**: 258–266

Anjanappa RB, Gruissem W (2021) Current progress and challenges in crop genetic transformation. J Plant Physiol **261**: 153411

Azodi CB, Lloyd JP, Shiu SH (2020) The cis-regulatory codes of response to combined heat and drought stress in *Arabidopsis thaliana*. NAR Genom Bioinform **2**: lqaa049

Bacete L, Schulz J, Engelsdorf T, Bartosova Z, Vaahtera L, Yan G, Gerhold JM, Ticha T, Ovstebo C, Gigli-Bisceglia N, et al. (2022) THESEUS1 modulates cell wall stiffness and abscisic acid production in *Arabidopsis thaliana*. Proc Natl Acad Sci USA **119**: e2119258119

Balla T (2013) Phosphoinositides: tiny lipids with giant impact on cell regulation. Physiol Rev **93**: 1019–1037

Baniwal SK, Bharti K, Chan KY, Fauth M, Ganguli A, Kotak S, Mishra SK, Nover L, Port M, Scharf KD, et al. (2004) Heat stress response in plants: a complex game with chaperones and more than twenty heat stress transcription factors. J Biosci **29**: 471–487

Bao Y, Aggarwal P, Robbins NE, 2nd, Sturrock CJ, Thompson MC, Tan HQ, Tham C, Duan L, Rodriguez PL, Vernoux T, et al. (2014) Plant roots use a patterning mechanism to position lateral root branches toward available water. Proc Natl Acad Sci USA **111**: 9319–9324

Bartlett MK, Klein T, Jansen S, Choat B, Sack L (2016) The correlations and sequence of plant stomatal, hydraulic, and wilting responses to drought. Proc Natl Acad Sci USA **113**: 13098–13103

Basu D, Haswell ES (2020) The mechanosensitive ion channel MSL10 potentiates responses to cell swelling in Arabidopsis seedlings. Curr Biol **30**: 2716–2728 e2716

Battisti DS, Naylor RL (2009) Historical warnings of future food insecurity with unprecedented seasonal heat. Science **323**: 240–244

Bellstaedt J, Trenner J, Lippmann R, Poeschl Y, Zhang X, Friml J, Quint M, Delker C (2019) A mobile auxin signal connects temperature sensing in cotyledons with growth responses in hypocotyls. Plant Physiol **180**: 757–766

Beirinckx S, Viaene T, Haegeman A, Debode J, Amery F, Vandenabeele S, Nelissen H, Inzé D, Tito R, Raes J, et al. (2020) Tapping into the maize root microbiome to identify bacteria that promote growth under chilling conditions. Microbiome **8**: 54

Bhaskara GB, Wen TN, Nguyen TT, Verslues PE (2017) Protein phosphatase 2Cs and microtubule-associated stress protein 1 control microtubule stability, plant growth, and drought response. Plant Cell **29**: 169–191

Bhaskara GB, Yang TH, Verslues PE (2015) Dynamic proline metabolism: importance and regulation in water limited environments. Front Plant Sci **6**: 484

Bhatnagar-Mathur P, Vadez V, Sharma KK (2008) Transgenic approaches for abiotic stress tolerance in plants: retrospect and prospects. Plant Cell Rep **27**: 411–424

Blackman BK (2017) Changing responses to changing seasons: natural variation in the plasticity of flowering time. Plant Physiol **173**: 16–26

Blair EJ, Bonnot T, Hummel M, Hay E, Marzolino JM, Quijada IA, Nagel DH (2019) Contribution of time of day and the circadian clock to the heat stress responsive transcriptome in Arabidopsis. Sci Rep **919**: 1–12

Blows MW, Hoffmann AA (2005) A reassessment of genetic limits to evolutionary change. Ecology **86**: 1371–1384

Blum A (2014) Genomics for drought resistance—getting down to earth. Funct Plant Biol **41**: 1191–1198

Boomsma CR, Vyn TJ (2008) Maize drought tolerance: potential improvements through arbuscular mycorrhizal symbiosis? Field Crops Res **108**: 14–31

Bonnot T, Blair EJ, Cordingley SJ, Nagel DH (2021) Circadian coordination of cellular processes and abiotic stress responses. Curr Opin Plant Biol **64**: 102133

Bonnot T, Nagel DH (2021) Time of the day prioritizes the pool of translating mRNAs in response to heat stress. Plant Cell **33**: 2164–2182

Bourbia I, Pritzkow C, Brodribb TJ (2021) Herb and conifer roots show similar high sensitivity to water deficit. Plant Physiol **186**: 1908–1918

Boursiac Y, Boudet J, Postaire O, Luu DT, Tournaire-Roux C, Maurel C (2008) Stimulus-induced downregulation of root water transport involves reactive oxygen species-activated cell signalling and plasma membrane intrinsic protein internalization. Plant J **56**: 207–218

Box MS, Huang BE, Domijan M, Jaeger KE, Khattak AK, Yoo SJ, Sedivy EL, Jones DM, Hearn TJ, Webb AAR, et al. (2014) ELF3 controls thermoresponsive growth in Arabidopsis. Curr Biol **25**: 194–199

Buell CR (2009) Poaceae genomes: going from unattainable to becoming a model clade for comparative plant genomics. Plant Physiol **149**: 111–116

Brininger C, Spradlin S, Cobani L, Evilia C (2018) The more adaptive to change, the more likely you are to survive: protein adaptation in extremophiles. Semin Cell Dev Biol **84**: 158–169

Brodribb T, Brodersen CR, Carriqui M, Tonet V, Dominguez CR, McAdam S (2021) Linking xylem network failure with leaf tissue death. New Phytol **232**: 68–79

Brunner I, Herzog C, Dawes MA, Arend M, Sperisen C (2015) How tree roots respond to drought. Front Plant Sci **6**: 547

Buckley TN, John GP, Scoffoni C, Sack L (2015) How does leaf anatomy influence water transport outside the xylem? Plant Physiol **168**: 1616–1635

Chapman SC, Chakraborty S, Dreccer MF, Howden SM (2012) Plant adaptation to climate change—opportunities and priorities in breeding. Crop Pasture Sci **63**: 251–268

Chaudhary A, Chen X, Lesniewska B, Boikine R, Gao J, Wolf S, Schneitz K (2021) Cell wall damage attenuates root hair patterning and tissue morphogenesis mediated by the receptor kinase STRUBBELIG. Development **148**: dev199425

Chaumont F, Tyerman SD (2014) Aquaporins: highly regulated channels controlling plant water relations. Plant Physiol **164**: 1600–1618

Chen C, Riley WJ, Prentice IC, Keenan TF (2022) CO_2 fertilization of terrestrial photosynthesis inferred from site to global scales. Proc Nal Acad Sci USA **119**: e2115627119

Chen T, Wu H, Wu J, Fan X, Li X, Lin Y (2017) Absence of OsβCA1 causes a CO_2 deficit and affects leaf photosynthesis and the stomatal response to CO_2 in rice. Plant J **90**: 344–357

Chen Y, Li W, Turner JA, Anderson CT (2021) PECTATE LYASE LIKE12 patterns the guard cell wall to coordinate turgor pressure and wall mechanics for proper stomatal function in Arabidopsis. Plant Cell **33**: 3134–3150

Chen ZJ, Mas P (2019) Interactive roles of chromatin regulation and circadian clock function in plants. Genome Biol **20**: 62

Choat B, Brodribb TJ, Brodersen CR, Duursma RA, Lopez R, Medlyn BE (2018) Triggers of tree mortality under drought. Nature **558**: 531–539

Chung BYW, Balcerowicz M, Di Antonio M, Jaeger KE, Geng F, Franaszek K, Marriott P, Brierley I, Firth AE, Wigge PA (2020) An RNA thermoswitch regulates daytime growth in Arabidopsis. Nat Plants **6**: 522–532

Cecchini NM, Monteoliva MI, Alvarez ME (2011) Proline dehydrogenase contributes to pathogen defense in Arabidopsis. Plant Physiol **155**: 1947–1959

Chong L, Xu R, Huang P, Guo P, Zhu M, Du H, Sun X, Ku L, Zhu JK, Zhu Y (2022) The tomato OST1–VOZ1 module regulates drought-mediated flowering. Plant Cell **34**: 2001–2018

Christenhusz MJM, Byng JW (2016) The number of known plants species in the world and its annual increase. Phytotaxa **261**: 201–217

Christmann A, Grill E, Huang J (2013) Hydraulic signals in long-distance signaling. Curr Opin Plant Biol **16**: 293–300

Christmann A, Weiler EW, Steudle E, Grill E (2007) A hydraulic signal in root-to-shoot signalling of water shortage. Plant J **52**: 167–174

Christmas MJ, Breed MF, Lowe AJ. (2016) Constraints to and conservation implications for climate change adaptation in plants. Conserv Genet **17**: 305–320

Conner JK (2003) Artificial selection: a powerful tool for ecologists. Ecology **84**: 1650–1660

Corlett RT (2016) Plant diversity in a changing world: status, trends, and conservation needs. Plant Divers **38**: 10–16

Covington MF, Maloof JN, Straume M, Kay SA, Harmer SL (2008) Global transcriptome analysis reveals circadian regulation of key pathways in plant growth and development. Genome Biol **9**: R130

Crepin N, Rolland F (2019) SnRK1 activation, signaling, and networking for energy homeostasis. Curr Opin Plant Biol **51**: 29–36

Cruz DF, De Meyer S, Ampe J, Sprenger H, Herman D, Van Hautegem T, De Block J, Inzé D, Nelissen H, Maere S (2020) Using single-plant-omics in the field to link maize genes to functions and phenotypes. Mol Syst Biol **16**: e9667

Cutler SR, Rodriguez PL, Finkelstein RR, Abrams SR (2010) Abscisic acid: emergence of a core signaling network. Annu Rev Plant Biol **61**: 651–679

Cuneo IF, Barrios-Masias F, Knipfer T, Uretsky J, Reyes C, Lenain P, Brodersen CR, Walker MA, McElrone AJ (2021) Differences in grapevine rootstock sensitivity and recovery from drought are linked to fine root cortical lacunae and root tip function. New Phytol **229**: 272–283

D'Ambrosio JM, Couto D, Fabro G, Scuffi D, Álvarez ME, Lamattina L, Munnik T, Andersson MX, Zipfel C, Laxalt AM (2017) PLC2 regulates MAMP-triggered immunity by modulating ROS production in Arabidopsis. Plant Physiol **175**: 970–981

Daszkowska-Golec A (2022) ABA is important not only under stress—revealed by the discovery of new ABA transporters. Trends Plant Sci **27**: 423–425

Dayer S, Herrera JC, Dai ZW, Burlett R, Lamarque LJ, Delzon S, Bortolami G, Cochard H, Gambetta GA (2020) The sequence and thresholds of leaf hydraulic traits underlying grapevine varietal differences in drought tolerance. J Exp Bot **71**: 4333–4344

De Jong F, Munnik T (2021) Attracted to membranes: lipid-binding domains in plants. Plant Physiol **185**: 707–723

De Kauwe MG, Medlyn BE, Tissue DT (2021) To what extent can rising [CO_2] ameliorate plant drought stress? New Phytol **231**: 2118–2124

Desai JS, Lawas LMF, Valente AM, Leman AR, Grinevich DO, Jagadish SVK, Doherty CJ (2021) Warm nights disrupt transcriptome rhythms in field-grown rice panicles. Proc Natl Acad Sci USA **118**: e2025899118

Des Marais DL, Auchincloss LC, Sukamtoh E, McKay JK, Logan T, Richards JH, Juenger TE (2014) Variation in MPK12 affects water use efficiency in Arabidopsis and reveals a link to ABA response. Proc Natl Acad Sci USA **111**: 2836–2841

Dickson EJ, Hille B (2019) Understanding phosphoinositides: rare, dynamic, and essential membrane phospholipids. Biochem J **476**: 1–23

Ding L, Milhiet T, Couvreur V, Nelissen H, Meziane A, Parent B, Aesaert S, Van Lijsebettens M, Inze D, Tardieu F, et al. (2020) Modification of the expression of the aquaporin ZmPIP2;5 affects water relations and plant growth. Plant Physiol **182**: 2154–2165

Ding L, Milhiet T, Parent B, Meziane A, Tardieu F, Chaumont F (2021) The plasma membrane aquaporin ZmPIP2;5 enhances the sensitivity of stomatal closure to water deficit. Plant Cell Environ **45**: 1146–1156

Di Martino C, Pizzuto R, Pallotta ML, De Santis A, Passarella S (2006) Mitochondrial transport in proline catabolism in plants: the existence of two separate translocators in mitochondria isolated from durum wheat seedlings. Planta **223**: 1123–1133

Dittrich M, Mueller HM, Bauer H, Peirats-Llobet M, Rodriguez PL, Geilfus CM, Carpentier SC, Al Rasheid KAS, Kollist H, Merilo E, et al. (2019) The role of Arabidopsis ABA receptors from the PYR/PYL/RCAR family in stomatal acclimation and closure signal integration. Nat Plants **5**: 1002–1011

Doheny-Adams T, Hunt L, Franks PJ, Beerling DJ, Gray JE (2012) Genetic manipulation of stomatal density influences stomatal size, plant growth and tolerance to restricted water supply. Phil Trans R Soc Lond B Biol Sci **367**: 547–555

Domec JC, King JS, Carmichael MJ, Overby AT, Wortemann R, Smith WK, Miao G, Noormets A, Johnson DM (2021) Aquaporins, and not changes in root structure, provide new insights into physiological responses to drought, flooding, and salinity. J Exp Bot **72**: 4489–4501

Domec JC, Noormets A, King JS, Sun G, McNulty SG, Gavazzi MJ, Boggs JL, Treasure EA (2009) Decoupling the influence of leaf and root hydraulic conductances on stomatal conductance and its sensitivity to vapour pressure deficit as soil dries in a drained loblolly pine plantation. Plant Cell Environ **32**: 980–991

Donovan LA, Maherali H, Caruso CM, Huber H, de Kroon H (2011) The evolution of the worldwide leaf economics spectrum. Trends Ecol Evol **26**: 88–95

Doumane M, Caillaud MC, Jaillais Y (2022) Experimental manipulation of phosphoinositide lipids: from cells to organisms. Trends Cell Biol **32**: 445–461

Doumane M, Lebecq A, Colin L, Fangain A, Stevens FD, Bareille J, Hamant O, Belkhadir Y, Munnik T, Jaillais Y, et al. (2021) Inducible depletion of PI(4,5)P2 by the synthetic iDePP system in Arabidopsis. Nat Plants **7**: 587–597

Du H, Huang F, Wu N, Li X, Hu H, Xiong L (2018) Integrative regulation of drought escape through ABA-dependent and -independent pathways in rice. Mol Plant **11**: 584–597

Dubois GA, Jaillais Y (2021) Anionic phospholipid gradients: an uncharacterized frontier of the plant endomembrane network. Plant Physiol **185**: 577–592

Duddek P, Carminati A, Koebernick N, Ohmann L, Lovric G, Delzon S, Rodriguez-Dominguez CM, King A, Ahmed MA (2022) The impact of drought-induced root and root hair shrinkage on root-soil contact. Plant Physiol **189**: 1232–1236

Duncan S, Holm S, Questa J, Irwin J, Grant A, Dean C (2015) Seasonal shift in timing of vernalization as an adaptation to extreme winter. Elife **4**: e06620

Dwivedi SL, Reynolds MP, Ortiz R (2021) Mitigating tradeoffs in plant breeding. iScience **24**: 102965

Edlefsen NE (1941) Some thermodynamic aspects of the use of soil-moisture by plants. Eos Trans Am Geophys Union **22**: 917–940

Ejiri M, Sawazaki Y, Shiono K (2020) Some accessions of amazonian wild rice (*Oryza glumaepatula*) constitutively form a barrier to radial oxygen loss along adventitious roots under aerated conditions. Plants **9**: 880

Ejiri M, Shiono K (2019) Prevention of radial oxygen loss is associated with exodermal suberin along adventitious roots of annual wild species of *Echinochloa*. Front Plant Sci **10**: 254

Endo A, Sawada Y, Takahashi H, Okamoto M, Ikegami K, Koiwai H, Seo M, Toyomasu T, Mitsuhashi W, Shinozaki K, et al. (2008) Drought induction of Arabidopsis 9-cis-epoxycarotenoid dioxygenase occurs in vascular parenchyma cells. Plant Physiol **147**: 1984–1993

Ermakova M, Osborn H, Groszmann M, Bala S, Bowerman A, Mcgaughey S, Byrt C, Alonso-Cantabrana H, Tyerman S, Furbank RT, et al. (2021) Expression of a CO_2-permeable aquaporin enhances mesophyll conductance in the C-4 species Setaria viridis. Elife **10**: 16

Escalante-Perez M, Krol E, Stange A, Geiger D, Al-Rasheid KA, Hause B, Neher E, Hedrich R (2011) A special pair of phytohormones controls excitability, slow closure, and external stomach formation in the Venus flytrap. Proc Natl Acad Sci USA **108**: 15492–15497

Eshel G, Araus V, Undurraga S, Soto DC, Moraga C, Montecinos A, Moyano T, Maldonado J, Díaz FP, Varala K, et al. (2021) Plant ecological genomics at the limits of life in the Atacama Desert. Proc Natl Acad Sci USA **118**: e2101177118

Fabro G, Kovacs I, Pavet V, Szabados L, Alvarez ME (2004) Proline accumulation and AtP5CS2 gene activation are induced by plant–pathogen incompatible interactions in Arabidopsis. Mol Plant-Microbe Interact **17**: 343–350

Fabro G, Rizzi YS, Alvarez ME (2016) Arabidopsis proline dehydrogenase contributes to flagellin-mediated PAMP-triggered immunity by affecting RBOHD. Mol Plant-Microbe Interact **29**: 620–628

Fang Y, Xiong L (2015) General mechanisms of drought response and their application in drought resistance improvement in plants. Cell Mol Life Sci **72**: 673–689

Farmer EE, Gao YQ, Lenzoni G, Wolfender JL, Wu Q (2020) Wound- and mechanostimulated electrical signals control hormone responses. New Phytol **227**: 1037–1050

Fernandez-Pozo N, Menda N, Edwards JD, Saha S, Tecle IY, Strickler SR, Bombarely A, Fisher-York T, Pujar A, Foerster H, et al. (2015) The Sol Genomics Network (SGN)–from genotype to phenotype to breeding. Nucleic Acids Res **43**: D1036–D1041

Feng W, Kita D, Peaucelle A, Cartwright HN, Doan V, Duan Q, Liu MC, Maman J, Steinhorst L, Schmitz-Thom I, et al. (2018) The FERONIA receptor kinase maintains cell-wall integrity during salt stress through Ca^{2+} signaling. Curr Biol **28**: 666–675 e665

Fitzpatrick CR, Copeland J, Wang PW, Guttman DS, Kotanen PM, Johnson MTJ (2018) Assembly and ecological function of the root microbiome across angiosperm plant species. Proc Natl Acad Sci USA **115**: E1157–E1165

Flütsch S, Wang Y, Takemiya A, Vialet-Chabrand SR, Klejchová M, Nigro A, Hills A, Lawson T, Blatt MR, Santelia D (2020) Guard cell starch degradation yields glucose for rapid stomatal opening in Arabidopsis. Plant Cell **32**: 2325–2344

Forlani G, Bertazzini M, Zarattini M, Funck D, Ruszkowski M, Nocek B (2015) Functional properties and structural characterization of rice D^1-pyrroline-5-carboxylate reductase. Front Plant Sci **6**: 565

Franks PJ, Adams MA, Amthor JS, Barbour MM, Berry JA, Ellsworth DS, Farquhar GD, Ghannoum O, Lloyd J, McDowell N (2013) Sensitivity of plants to changing atmospheric CO_2 concentration: from the geological past to the next century. New Phytol **197**: 1077–1094

Franks SJ (2011) Plasticity and evolution in drought avoidance and escape in the annual plant Brassica rapa. New Phytol 190: 249–257

Fukao T, Xu K, Ronald PC, Bailey-Serres J (2006) A variable cluster of ethylene response factor-like genes regulates metabolic and developmental acclimation responses to submergence in rice. Plant Cell 18: 2021–2034

Fukao T, Yeung E, Bailey-Serres J (2011) The submergence tolerance regulator SUB1A mediates crosstalk between submergence and drought tolerance in rice. Plant Cell 23: 412–427

Fujii Y, Tanaka H, Konno N, Ogasawara Y, Hamashima N, Tamura S, Hasegawa S, Hayasaki Y, Okajima K, Kodama Y (2017) Phototropin perceives temperature based on the lifetime of its photoactivated state. Proc Natl Acad Sci USA 114: 9206–9211

Funck D, Baumgarten L, Stift M, von Wiren N, Schonemann L (2020) Differential contribution of P5CS isoforms to stress tolerance in Arabidopsis. Front Plant Sci 11: 565134

Funck D, Winter G, Baumgarten L, Forlani G (2012) Requirement of proline synthesis during Arabidopsis reproductive development. BMC Plant Biol 12: 191

Galbiati F, Chiozzotto R, Locatelli F, Spada A, Genga A, Fornara F (2016) Hd3a, RFT1 and Ehd1 integrate photoperiodic and drought stress signals to delay the floral transition in rice. Plant Cell Environ 39: 1982–1993

Gerth K, Lin F, Menzel W, Krishnamoorthy P, Stenzel I, Heilmann M, Heilmann I (2017) Guilt by association: a phenotype-based view of the plant phosphoinositide network. Annu Rev Plant Biol 68: 349–374

Gibbs JW (1961) The Scientific Papers of J. Willard Gibbs. Volume 1: Thermodynamics. Dover, New York, NY

Giberti S, Funck D, Forlani G (2014) Δ^1-pyrroline-5-carboxylate reductase from Arabidopsis thaliana: stimulation or inhibition by chloride ions and feedback regulation by proline depend on whether NADPH or NADH acts as cosubstrate. New Phytol 202: 911–919

González-Guzmán M, Cellini F, Fotopoulos V, Balestrini R, Arbona V (2022) New approaches to improve crop tolerance to biotic and abiotic stresses. Physiol Plant 174: e13547

Granger RA (1995) Fluid Mechanics. Dover, New York, NY

Groen SC, Joly-Lopez Z, Platts AE, Natividad M, Fresquez Z, Mauck WM, Quintana MR, Cabral CLU, Torres RO, Satija R, et al. (2020) The strength and pattern of natural selection on gene expression in rice. Nature 578: 572–576

Groen SC, Joly-Lopez Z, Platts AE, Natividad M, Fresquez Z, Mauck WM, Quintana MR, Cabral CLU, Torres RO, Satija R, et al. (2022) Evolutionary systems biology reveals patterns of rice adaptation to drought-prone agro-ecosystems. Plant Cell 34: 759–783

Grondin A, Mauleon R, Vadez V, Henry A (2016) Root aquaporins contribute to whole plant water fluxes under drought stress in rice (Oryza sativa L.). Plant Cell Environ 39: 347–365

Grundy J, Stoker C, Carré IA (2015) Circadian regulation of abiotic stress tolerance in plants. Front Plant Sci 6: 648

Gujas B, Cruz TMD, Kastanaki E, Vermeer JEM, Munnik T, Rodriguez-Villalon A (2017) Perturbing phosphoinositide homeostasis oppositely affects root vascular differentiation in Arabidopsis thaliana roots. Development 144: 3578–3589

Hamant O, Inoue D, Bouchez D, Dumais J, Mjolsness E (2019) Are microtubules tension sensors? Nat Commun 10: 2360

Hammond WM, Adams HD (2019) Dying on time: traits influencing the dynamics of tree mortality risk from drought. Tree Physiol 39: 906–909

Hammond WM, Williams AP, Abatzoglou JT, Adams HD, Klein T, Lopez R, Saenz-Romero C, Hartmann H, Breshears DD, Allen CD (2022) Global field observations of tree die-off reveal hotter-drought fingerprint for Earth's forests. Nat Commun 13: 1761

Hancock AM, Brachi B, Faure N, Horton MW, Jarymowycz LB, Sperone FG, Toomajian C, Roux F, Bergelson J (2011) Adaptation to climate across the Arabidopsis thaliana genome. Science 334: 83–86

Hanson AD, Rathinasabapathi B, Rivoal J, Burnet M, Dillon MO, Gage DA (1994) Osmoprotective compounds in the plumbaginaceae—a natural experiment in metabolic engineering of stress tolerance. Proc Natl Acad Sci USA 91: 306–310

Hare PD, Cress WA (1997) Metabolic implications of stress-induced proline accumulation in plants. Plant Growth Regul 21: 79–102

Hashiguchi A, Komatsu S (2016) Impact of post-translational modifications of crop proteins under abiotic stress. Proteomes 4: 42

Hashimoto M, Negi J, Young J, Israelsson M, Schroeder JI, Iba K (2006) Arabidopsis HT1 kinase is a negative regulator of stomatal closing. Nature Cell Biol 8: 391–397

Hattori Y, Nagai K, Furukawa S, Song X-J, Kawano R, Sakakibara H, Wu J, Matsumoto T, Yoshimura A, Kitano H, et al. (2009) The ethylene response factors SNORKEL1 and SNORKEL2 allow rice to adapt to deep water. Nature 460: 1026–1030

Hayes S, Pantazopoulou CK, van Gelderen K, Reinen E, Tween AL, Sharma A, de Vries M, Prat S, Schuurink RC, Testerink C, et al. (2019) Soil salinity limits plant shade avoidance. Curr Biol 29: 1669–1676.e4

Hayes S, Schachtschabel J, Mishkind M, Munnik T, Arisz SA (2021) Hot topic: thermosensing in plants. Plant Cell Environ 44: 2018–2033

Hebbelmann I, Selinski J, Wehmeyer C, Goss T, Voss I, Mulo P, Kangasjärvi S, Aro EM, Oelze ML, Dietz KJ, et al. (2011) Multiple strategies to prevent oxidative stress in Arabidopsis plants lacking the malate valve enzyme NADP-malate dehydrogenase. J Exp Bot 63: 1445–1459

He CJ, Morgan PW, Drew MC (1996) Transduction of an ethylene signal is required for cell death and lysis in the root cortex of maize during aerenchyma formation induced by hypoxia. Plant Physiol 112: 463–472

Henderson SW, Nourmohammadi S, Ramesh SA, Yool AJ (2022) Aquaporin ion conductance properties defined by membrane environment, protein structure, and cell physiology. Biophys Rev 14: 181–198

Hernandez-Santana V, Rodriguez-Dominguez CM, Fernandez JE, Diaz-Espejo A (2016) Role of leaf hydraulic conductance in the regulation of stomatal conductance in almond and olive in response to water stress. Tree Physiol 36: 725–735

Hiyama A, Takemiya A, Munemasa S, Okuma E, Sugiyama N, Tada Y, Murata Y, Shimazaki KI (2017) Blue light and CO_2 signals converge to regulate light-induced stomatal opening. Nat Commun 8: 1284

Hoffmann AA, Sgrò CM (2011) Climate change and evolutionary adaptation. Nature 470: 479–485

Hostetler AN, Govindarajulu R, Hawkins JS (2021a) QTL mapping in an interspecific sorghum population uncovers candidate regulators of salinity tolerance. Plant Stress 2: 100024

Hostetler AN, Khangura RS, Dilkes BP, Sparks EE (2021b) Bracing for sustainable agriculture: the development and function of brace roots in members of Poaceae. Curr Opin Plant Biol 59: 101985

Hsu PK, Takahashi T, Munemasa S, Merilo E, Laanemets K, Waadt R, Pater D, Kollist H, Schroeder JI (2018) Abscisic acid-independent stomatal CO_2 signal transduction pathway and convergence of CO_2 and ABA signaling downstream of OST1 Kinase. Proc Natl Acad Sci USA 115: E9971–E9980

Hsu PK, Takahashi Y, Merilo E, Costa A, Zhang L, Kernig K, Lee Katie H, Schroeder JI (2021) Raf-like kinases and receptor-like (pseudo)kinase GHR1 are required for stomatal vapor pressure difference response. Proc Natl Acad Sci USA 118: e2107280118

Hu H, Boisson-Dernier A, Israelsson-Nordström M, Böhmer M, Godoski J, Ries A, Kuhn JM, Schroeder JI (2010) Carbonic anhydrases are upstream regulators of CO_2-controlled stomatal movements in guard cells. Nat Cell Biol 12: 87–93

Hu J, Zhuang Y, Li X, Li X, Sun C, Ding Z, Xu R, Zhang D (2022) Time-series transcriptome comparison reveals the gene regulation network under salt stress in soybean (*Glycine max*) roots. BMC Plant Biol **22**: 157

Hwang K, Susila H, Nasim Z, Jung JY, Ahn JH (2019) Arabidopsis ABF3 and ABF4 transcription factors act with the NF–YC complex to regulate SOC1 expression and mediate drought-accelerated flowering. Mol Plant **12**: 489–505

Iannacone R, Cellini F, Morelli G, Ruberti I (2012) Translational biology approaches to improve abiotic stress tolerance in crops. Improv Crop Resist Abiotic Stress **1**: 207–239

Iosip AL, Bohm J, Scherzer S, Al-Rasheid KAS, Dreyer I, Schultz J, Becker D, Kreuzer I, Hedrich R (2020) The Venus flytrap trigger hair-specific potassium channel KDM1 can reestablish the K + gradient required for hapto-electric signaling. PLoS Biol **18**: e3000964

Ischebeck T, Seiler S, Heilmann I (2010) At the poles across kingdoms: phosphoinositides and polar tip growth. Protoplasma **240**: 13–31

Jaglo-Ottosen KR, Gilmour SJ, Zarka DG, Schabenberger O, Thomashow MF (1998) Arabidopsis CBF1 overexpression induces COR genes and enhances freezing tolerance. Science **280**: 104–106

Jakobson L, Vaahtera L, Toldsepp K, Nuhkat M, Wang C, Wang YS, Horak H, Valk E, Pechter P, Sindarovska Y, et al. (2016) Natural variation in Arabidopsis Cvi-0 accession reveals an important role of MPK12 in guard cell CO_2 signaling. PLoS Biol **14**: e2000322

Javot H, Lauvergeat V, Santoni V, Martin-Laurent F, Güçlü J, Vinh J, Heyes J, Franck KI, Schäffner AR, Bouchez D, et al. (2003) Role of a single aquaporin isoform in root water uptake. Plant Cell **15**: 509–522

Justino GC, Sodek L (2013) Recovery of nitrogen fixation after short-term flooding of the nodulated root system of soybean. J Plant Physiol **170**: 235–241

Jordan DR, Hunt CH, Cruickshank AW, Borrell AK, Henzell RG (2012) The relationship between the stay-green trait and grain yield in elite Sorghum hybrids grown in a range of environments. Crop Sci **52**: 1153–1161

Jung JH, Domijan M, Klose C, Biswas S, Ezer D, Gao M, Khattak AK, Box MS, Charoensawan V, Cortijo S, et al. (2016) Phytochromes function as thermosensors in Arabidopsis. Science **354**: 886–889

Jung JH, Barbosa AD, Hutin S, Kumita JR, Gao MJ, Derwort D, Silva CS, Lai XL, Pierre E, Geng F, et al. (2020) A prion-like domain in ELF3 functions as a thermosensor in Arabidopsis. Nature **585**: 256–260

Kalladan R, Lasky JR, Chang TZ, Sharma S, Juenger TE, Verslues PE (2017) Natural variation identifies genes affecting drought-induced abscisic acid accumulation in *Arabidopsis thaliana*. Proc Natl Acad Sci USA **114**: 11536–11541

Kamiyama Y, Hirotani M, Ishikawa S, Minegishi F, Katagiri S, Rogan CJ, Takahashi F, Nomoto M, Ishikawa K, Kodama Y, et al. (2021) Arabidopsis group C Raf-like protein kinases negatively regulate abscisic acid signaling and are direct substrates of SnRK2. Proc Natl Acad Sci USA **118**: e2100073118

Kapilan R, Vaziri M, Zwiazek JJ (2018) Regulation of aquaporins in plants under stress. Biol Res **51**: 4

Kashi Y, King D (2006) Simple sequence repeats as advantageous mutators in evolution. Trends Genet **22**: 253–259

Kawa D, Julkowska MM, Sommerfeld HM, Ter Horst A, Haring MA, Testerink C (2016) Phosphate-dependent root system architecture responses to salt stress. Plant Physiol **172**: 690–706

Kawecki TJ, Ebert D (2004) Conceptual issues in local adaptation. Ecol Lett **7**: 1225–1241

Kellermeier F, Armengaud P, Seditas TJ, Danku J, Salt DE, Amtmann A (2014) Analysis of the root system architecture of Arabidopsis provides a quantitative readout of crosstalk between nutritional signals. Plant Cell **26**: 1480–1496

Kenney AM, McKay JK, Richards JH, Juenger TE (2014) Direct and indirect selection on flowering time, water-use efficiency (WUE, δ (13)C), and WUE plasticity to drought in Arabidopsis thaliana. Ecol Evol **4**: 4505–4521

Kesari R, Lasky JR, Villamor JG, Marais DLD, Chen YJC, Liu TW, Lin W, Juenger TE, Verslues PE (2012) Intron-mediated alternative splicing of Arabidopsis P5CS1 and its association with natural variation in proline and climate adaptation. Proc Natl Acad Sci USA **109**: 9197–9202

Kim SC, Wang X (2020) Phosphatidic acid: an emerging versatile class of cellular mediators. Essays Biochem **64**: 533–546

Kinoshita T, Ono N, Hayashi Y, Morimoto S, Nakamura S, Soda M, Kato Y, Ohnishi M, Nakano T, Inoue S, et al. (2011) FLOWERING LOCUS T regulates stomatal opening. Curr Biol **21**: 1232–1238

Kirscht A, Kaptan SS, Bienert GP, Chaumont F, Nissen P, De Groot BL, Kjellbom P, Gourdon P, Johanson U (2016) Crystal structure of an ammonia-permeable aquaporin. PLoS Biol **14**: 19

Kitomi Y, Hanzawa E, Kuya N, Inoue H, Hara N, Kawai S, Kanno N, Endo M, Sugimoto K, Yamazaki T, et al. (2020) Root angle modifications by the DRO1 homolog improve rice yields in saline paddy fields. Proc Natl Acad Sci USA **117**: 21242–21250

Klepsch M, Zhang Y, Kotowska MM, Lamarque LJ, Nolf M, Schuldt B, Torres-Ruiz JM, Qin DW, Choat B, Delzon S, et al. (2018) Is xylem of angiosperm leaves less resistant to embolism than branches? Insights from microCT, hydraulics, and anatomy. J Exp Bot **69**: 5611–5623

Klusener B, Boheim G, Liss H, Engelberth J, Weiler EW (1995) Gadolinium-sensitive, voltage-dependent calcium release channels in the endoplasmic reticulum of a higher plant mechanoreceptor organ. EMBO J **14**: 2708–2714

Knipfer T, Danjou M, Vionne C, Fricke W (2021) Salt stress reduces root water uptake in barley (*Hordeum vulgare* L.) through modification of the transcellular transport path. Plant Cell Environ **44**: 458–475

Knipfer T, Reyes C, Earles JM, Berry ZC, Johnson DM, Brodersen CR, McElrone AJ (2019) Spatiotemporal coupling of vessel cavitation and discharge of stored xylem water in a tree sapling. Plant Physiol **179**: 1658–1668

Koenig D, Weigel D (2015) Beyond the thale: comparative genomics and genetics of Arabidopsis relatives. Nat Rev Genet **16**: 285–298

Kolbe AR, Brutnell TP, Cousins AB, Studer A (2018) Carbonic anhydrase mutants in *Zea mays* have altered stomatal responses to environmental signals. Plant Physiol **177**: 980–989

Kooyers NJ (2015) The evolution of drought escape and avoidance in natural herbaceous populations. Plant Sci **234**: 155–162

Kourghi M, Nourmohammadi S, Pei JV, Qiu J, Mcgaughey S, Tyerman SD, Byrt CS, Yool AJ (2017) Divalent cations regulate the ion conductance properties of diverse classes of aquaporins. Int J Mol Sci **18**: 2323

Koziol L, Rieseberg LH, Kane N, Bever JD (2012) Reduced drought tolerance during domestication and the evolution of weediness results from tolerance-growth trade-offs. Evolution **66**: 3803–3814

Kretzschmar T, Pelayo MAF, Trijatmiko KR, Gabunada LFM, Alam R, Jimenez R, Mendioro MS, Slamet-Loedin IH, Sreenivasulu N, Bailey-Serres J, et al. (2015) A trehalose-6-phosphate phosphatase enhances anaerobic germination tolerance in rice. Nat Plants **1**: 15124

Kudo M, Kidokoro S, Yoshida T, Mizoi J, Kojima M, Takebayashi Y, Sakakibara H, Fernie AR, Shinozaki K, Yamaguchi-Shinozaki K (2019) A gene-stacking approach to overcome the trade-off between drought stress tolerance and growth in Arabidopsis. Plant J **97**: 240–256

Kumar MN, Jane WN, Verslues PE (2013) Role of the putative osmosensor Arabidopsis histidine kinase 1 (AHK1) in dehydration avoidance and low water potential response. Plant Physiol **161**: 942–953

Kuroda R, Kato M, Tsuge T, Aoyama T (2021) Arabidopsis phosphatidylinositol 4-phosphate 5-kinase genes PIP5K7, PIP5K8, and PIP5K9 are redundantly involved in root growth adaptation to osmotic stress. Plant J 106: 913–927

Kuroha T, Nagai K, Gamuyao R, Wang DR, Furuta T, Nakamori M, Kitaoka T, Adachi K, Minami A, Mori Y, et al. (2018) Ethylene-gibberellin signaling underlies adaptation of rice to periodic flooding. Science 361: 181–186

Kuromori T, Seo M, Shinozaki K (2018) ABA transport and plant water stress responses. Trends Plant Sci 23: 513–522

Kuromori T, Sugimoto E, Shinozaki K (2014) Intertissue signal transfer of abscisic acid from vascular cells to guard cells. Plant Physiol 164: 1587–1592

Kusano H, Testerink C, Vermeer JEM, Tsuge T, Oka A, Shimada H, Munnik T, Aoyama T (2008) The Arabidopsis phosphatidylinositol phosphate 5-kinase PIP5K3 is a key regulator for root hair tip growth. Plant Cell 20: 367–380

Lamers J, van der Meer T, Testerink C (2020) How plants sense and respond to stressful environments. Plant Physiol 182: 1624–1635

Leakey ADB, Ferguson JN, Pignon CP, Wu A, Jin Z, Hammer GL, Lobell DB (2019) Water use efficiency as a constraint and target for improving the resilience and productivity of C3 and C4 crops. Ann Rev Plant Biol 70: 781–808

Lee CM, Thomashow MF (2012) Photoperiodic regulation of the C-repeat binding factor (CBF) cold acclimation pathway and freezing tolerance in Arabidopsis thaliana. Proc Natl Acad Sci USA 109: 15054–15059

Leebens-Mack JH, Barker MS, Carpenter EJ, Deyholos MK, Gitzendanner MA, Graham SW, Grosse I, Melkonian M, Mirarab S, Porsch M, et al. (2019) One thousand plant transcriptomes and the phylogenomics of green plants. Nature 574: 679–685

Legris M, Klose C, Burgie ES, Rojas CC, Neme M, Hiltbrunner A, Wigge PA, Schafer E, Vierstra RD, Casal JJ (2016) Phytochrome B integrates light and temperature signals in Arabidopsis. Science 354: 897–900

LeNoble ME, Spollen WG, Sharp RE (2004) Maintenance of shoot growth by endogenous ABA: genetic assessment of the involvement of ethylene suppression. J Exp Bot 55: 237–245

Li XM, Delzon S, Torres-Ruiz J, Badel E, Burlett R, Cochard H, Jansen S, King A, Lamarque LJ, Lenoir N, et al. (2020) Lack of vulnerability segmentation in four angiosperm tree species: evidence from direct X-ray microtomography observation. Ann For Sci 77: 37

Lin C, Ogorek LLP, Pedersen O, Sauter M (2021) Oxygen in the air and oxygen dissolved in the floodwater both sustain growth of aquatic adventitious roots in rice. J Exp Bot 72: 1879–1890

Lin Z, Li Y, Zhang Z, Liu X, Hsu CC, Du Y, Sang T, Zhu C, Wang Y, Satheesh V, et al. (2020) A RAF-SnRK2 kinase cascade mediates early osmotic stress signaling in higher plants. Nat Commun 11: 613

Lind KR, Siemianowski O, Yuan B, Sizmur T, VanEvery H, Banerjee S, Cademartiri L (2021) Evidence for root adaptation to a spatially discontinuous water availability in the absence of external water potential gradients. Proc Natl Acad Sci USA 118: e2012892118

Ling Y, Mahfouz MM, Zhou S (2021) Pre-mRNA alternative splicing as a modulator for heat stress response in plants. Trends Plant Sci 26: 1153–1170

Liu H, Wei JP, Yang T, Mu WX, Song B, Yang T, Fu Y, Wang XB, Hu GH, Li WS, et al. (2019) Molecular digitization of a botanical garden: high-depth whole-genome sequencing of 689 vascular plant species from the Ruili Botanical Garden. Gigascience 8: giz007

Livneh B, Hoerling MP (2016) The physics of drought in the US central great plains. J Climate 29: 6783–6804

Longkumer T, Chen CY, Biancucci M, Bhaskara GB, Verslues PE (2022) Spatial differences in stoichiometry of EGR phosphatase and Microtubule-associated Stress Protein 1 control root meristem activity during drought stress. Plant Cell 34: 742–758

Lorenzo-Orts L, Couto D, Hothorn M (2020) Identity and functions of inorganic and inositol polyphosphates in plants. New Phytol 225: 637–652

Ludwig W, Hayes S, Trenner J, Delker C, Quint M (2021) On the evolution of plant thermomorphogenesis. J Exp Bot 72: 7345–7358

Lynch JP (2007) TURNER REVIEW No. 14. Roots of the second green revolution. Aust J Bot 55: 493

Lynch JP (2015) Root phenes that reduce the metabolic costs of soil exploration: opportunities for 21st century agriculture. Plant Cell Environ 38: 1775–1784

Lynch M, Walsh B (1998) Genetics and analysis of quantitative traits. Sinauer Associates, Inc. Sunderland, Massachusetts, USA

Ma L, Zhang H, Sun L, Jiao Y, Zhang G, Miao C, Hao F (2011) NADPH oxidase AtrbohD and AtrbohF function in ROS-dependent regulation of Na + /K + homeostasis in Arabidopsis under salt stress. J Exp Bot 63: 305–317

Mackay TFC (2001) The genetic architecture of quantitative traits. Ann Rev Genet 35: 303–339

MacTavish R, Anderson JT (2020) Resource availability alters fitness trade-offs: implications for evolution in stressful environments. Am J Bot 107: 308–318

Maggio A, Bressan RA, Zhao Y, Park J, Yun DJ (2018) It's hard to avoid avoidance: uncoupling the evolutionary connection between plant growth, productivity and stress "tolerance". Int J Mol Sci 19: 3671

Mairhofer S, Zappala S, Tracy S, Sturrock C, Bennett M, Mooney S, Pridmore T (2011) RooTrak: automated recovery of 3D plant root architecture in soil from X-ray micro computed tomography using visual tracking. Plant Physiol 158: 561–569

Mano Y, Nakazono M (2021) Genetic regulation of root traits for soil flooding tolerance in genus Zea. Breed Sci 71: 30–39

Mano Y, Omori F (2013) Flooding tolerance in interspecific introgression lines containing chromosome segments from teosinte (Zea nicaraguensis) in maize (Zea mays subsp. mays). Ann Bot 112: 1125–1139

Markham KK, Greenham K (2021) Abiotic stress through time. New Phytol 231: 40–46

Marten H, Hyun T, Gomi K, Seo S, Hedrich R, Roelfsema MR (2008) Silencing of NtMPK4 impairs CO$_2$-induced stomatal closure, activation of anion channels and cytosolic Ca^{2+} signals in Nicotiana tabacum guard cells. Plant J 55: 698–708

Martignago D, Siemiatkowska B, Lombardi A, Conti L (2020) Abscisic acid and flowering regulation: many targets, different places. Int J Mol Sci 21: 9700

Martiniere A, Fiche JB, Smokvarska M, Mari S, Alcon C, Dumont X, Hematy K, Jaillais Y, Nollmann M, Maurel C (2019) Osmotic stress activates two reactive oxygen species pathways with distinct effects on protein nanodomains and diffusion. Plant Physiol 179: 1581–1593

Martin-StPaul N, Delzon S, Cochard H (2017) Plant resistance to drought depends on timely stomatal closure. Ecol Lett 20: 1437–1447

Maurel C, Tournaire-Roux C, Verdoucq L, Santoni V (2021) Hormonal and environmental signaling pathways target membrane water transport. Plant Physiol 187: 2056–2070

Maurel C, Verdoucq L, Rodrigues O (2016) Aquaporins and plant transpiration. Plant Cell Environ 39: 2580–2587

McCulloh KA, Johnson DM, Meinzer FC, Woodruff DR (2014) The dynamic pipeline: hydraulic capacitance and xylem hydraulic safety in four tall conifer species. Plant Cell Environ 37: 1171–1183

McDowell NG, Sapes G, Pivovaroff A, Adams HD, Allen CD, Anderegg WRL, Arend M, Breshears DD, Brodribb T, Choat B, et al. (2022) Mechanisms of woody-plant mortality under rising drought, CO$_2$ and vapour pressure deficit. Nat Rev Earth Environ 3: 294–308

McElrone AJ, Manuck CM, Brodersen CR, Patakas A, Pearsall KR, Williams LE (2021) Functional hydraulic sectoring in grapevines as evidenced by sap flow, dye infusion, leaf removal and micro-computed tomography. AoB Plants 13: plab003

Mckay JK, Richards JH, Mitchell-Olds T (2003) Genetics of drought adaptation in *Arabidopsis thaliana*: I. Pleiotropy contributes to genetic correlations among ecological traits. Mol Ecol 12: 1137–1151

Medlyn BE, Barton CVM, Broadmeadow MSJ, Ceulemans R, De Angelis P, Forstreuter M, Freeman M, Jackson SB, Kellomaki S, Laitat E, et al. (2001) Stomatal conductance of forest species after long-term exposure to elevated CO_2 concentration: a synthesis. New Phytol 149: 247–264

Miao C, Xiao L, Hua K, Zou C, Zhao Y, Bressan RA, Zhu J-K (2018) Mutations in a subfamily of abscisic acid receptor genes promote rice growth and productivity. Proc Natl Acad Sci 115: 6058–6063

Miflin B (2000) Crop improvement in the 21st century. J Exp Bot 51: 1–8

Miller G, Honig A, Stein H, Suzuki N, Mittler R, Zilberstein A (2009) Unraveling Δ^1-pyrroline-5-carboxylate-proline cycle in plants by uncoupled expression of proline oxidation enzymes. J Biol Chem 284: 26482–26492

Miller G, Stein H, Honig A, Kapulnik Y, Zilberstein A (2005) Responsive modes of *Medicago sativa* proline dehydrogenase genes during salt stress and recovery dictate free proline accumulation. Planta 222: 70–79

Miller ML, Roddy AB, Brodersen CR, McElrone AJ, Johnson DM (2020) Anatomical and hydraulic responses to desiccation in emergent conifer seedlings. Am J Bot 107: 1177–1188

Miryeganeh M (2021) Plants' epigenetic mechanisms and abiotic stress. Genes (Basel) 12: 1106

Mishkind M, Vermeer JEM, Darwish E, Munnik T (2009) Heat stress activates phospholipase D and triggers PIP_2 accumulation at the plasma membrane and nucleus. Plant J 60: 10–21

Moe-Lange J, Gappel NM, Machado M, Wudick MM, Sies CSA, Schott-Verdugo SN, Bonus M, Mishra S, Hartwig T, Bezrutczyk M, et al. (2021) Interdependence of a mechanosensitive anion channel and glutamate receptors in distal wound signaling. Sci Adv 7: eabg4298

Mom R, Muries B, Benoit P, Robert-Paganin J, Réty S, Venisse JS, Padua A, Label P, Auguin D (2021) Voltage-gating of aquaporins, a putative conserved safety mechanism during ionic stresses. FEBS Lett 595: 41–57

Mommer L, Kirkegaard J, van Ruijven J (2016) Root-root interactions: towards a rhizosphere framework. Trends Plant Sci 21: 209–217

Monroe JG, Powell T, Price N, Mullen JL, Howard A, Evans K, Lovell JT, McKay JK (2018) Drought adaptation in *Arabidopsis thaliana* by extensive genetic loss-of-function. eLife 7: e41038

Morales A, De Boer HJ, Douma JC, Elsen S, Engels S, Glimmerveen T, Sajeev N, Huber M, Luimes M, Luitjens E, et al. (2021) Effects of sub-lethal single, simultaneous, and sequential abiotic stresses on phenotypic traits of *Arabidopsis thaliana*. AoB Plants 14: plac029

Mori IC, Rhee J, Shibasaka M, Sasano S, Kaneko T, Horie T, Katsuhara M (2014) CO_2 transport by PIP2 aquaporins of barley. Plant Cell Physiol 55: 251–257

Morris EC, Griffiths M, Golebiowska A, Mairhofer S, Burr-Hersey J, Goh T, von Wangenheim D, Atkinson B, Sturrock CJ, Lynch JP, et al. (2017) Shaping 3D root system architecture. Curr Biol 27: R919–R930

Mott KA, Sibbernsen ED, Shope JC (2008) The role of the mesophyll in stomatal responses to light and CO_2. Plant Cell Environ 31: 1299–1306

Munnik T (2001) Phosphatidic acid—an emerging plant lipid second messenger. Trends Plant Sci 6: 227–233

Munnik T (2014) PI–PLC: phosphoinositide-phospholipase C in plant signaling. *In* X Wang, ed, Phospholipases in Plant Signaling. Vol 20—Signaling and Communication in Plants. Springer-Verlag, Berlin Heidelberg, Germany, pp 27–54

Munnik T, Mongrand S, Zársky V, Blatt M (2021) Dynamic membranes—the indispensable platform for plant growth, signaling, and development. Plant Physiol 185: 547–549

Munnik T, Testerink C (2009) Plant phospholipid signalling—'in a nutshell'. J Lipid Res 50: 260–265

Munnik T, Vermeer JEM (2010) Osmotic stress-induced phosphoinositide and inositol phosphate signalling in plants. Plant Cell Environ 33: 655–669

Murahama M, Yoshida T, Hayashi F, Ichino T, Sanada Y, Wada K (2001) Purification and characterization of Delta(1)-pyrroline-5-carboxylate reductase isoenzymes, indicating differential distribution in spinach (Spinacia oleracea L.) leaves. Plant Cell Physiol 42: 742–750

Murthy SE, Dubin AE, Whitwam T, Jojoa-Cruz S, Cahalan SM, Mousavi SAR, Ward AB, Patapoutian A (2018) OSCA/TMEM63 are an evolutionarily conserved family of mechanically activated ion channels. Elife 7: e41844

Nadakuduti SS, Enciso-Rodríguez F (2020) Advances in genome editing with CRISPR systems and transformation technologies for plant DNA manipulation. Front Plant Sci 11: 637159

Nagano AJ, Sato Y, Mihara M, Antonio BA, Motoyama R, Itoh H, Nagamura Y, Izawa T (2012) Deciphering and prediction of transcriptome dynamics under fluctuating field conditions. Cell 151: 1358–1369

Nagai K, Mori Y, Ishikawa S, Furuta T, Gamuyao R, Niimi Y, Hobo T, Fukuda M, Kojima M, Takebayashi Y, et al. (2020) Antagonistic regulation of the gibberellic acid response during stem growth in rice. Nature 584: 109–114

Nelissen H, Sprenger H, Demuynck K, De Block J, Van Hautegem T, De Vliegher A, Inzé D (2020) From laboratory to field: yield stability and shade avoidance genes are massively differentially expressed in the field. Plant Biotechnol J 18: 1112–1114

Noack LC, Bayle V, Armengot L, Rozier F, Mamode-Cassim A, Stevens FD, Caillaud MC, Munnik T, Mongrand S, Pleskot R, et al. (2022) A nanodomain-anchored scaffolding complex is required for the function and localization of phosphatidylinositol 4-kinase alpha in plants. Plant Cell 34: 302–332

Noack LC, Jaillais Y (2020) Functions of anionic lipids in plants. Annu Rev Plant Biol 71: 71–102

Nguyen CT, Kurenda A, Stolz S, Chetelat A, Farmer EE (2018) Identification of cell populations necessary for leaf-to-leaf electrical signaling in a wounded plant. Proc Natl Acad Sci USA 115: 10178–10183

Nuccio ML, Paul M, Bate NJ, Cohn J, Cutler SR (2018) Where are the drought tolerant crops? An assessment of more than two decades of plant biotechnology effort in crop improvement. Plant Sci 273: 110–119

Oh GGK, O'Leary BM, Signorelli S, Millar AH (2022) Alternative oxidase (AOX) 1a and 1d limit proline-induced oxidative stress and aid salinity recovery in Arabidopsis. Plant Physiol 188: 1521–1536

O'Malley RC, Huang SSC, Song L, Lewsey MG, Bartlett A, Nery JR, Galli M, Gallavotti A, Ecker JR (2016) Cistrome and epicistrome features shape the regulatory DNA landscape. Cell 165: 1280–1292

Orosa-Puente B, Leftley N, von Wangenheim D, Banda J, Srivastava AK, Hill K, Truskina J, Bhosale R, Morris E, Srivastava M, et al. (2018) Root branching toward water involves posttranslational modification of transcription factor ARF7. Science 362: 1407–1410

Patel D, Basu M, Hayes S, Majláth I, Hetherington FM, Tschaplinski TJ, Franklin KA (2013) Temperature-dependent shade avoidance involves the receptor-like kinase ERECTA. Plant J 73: 980–992

Pedersen O, Sauter M, Colmer TD, Nakazono M (2021) Regulation of root adaptive anatomical and morphological traits during low soil oxygen. New Phytol 229: 42–49

Platre MP, Noack LC, Doumane M, Bayle V, Simon MLA, Maneta-Peyret L, Fouillen L, Stanislas T, Armengot L, Pejchar P, et al.

(2018) A combinatorial lipid code shapes the electrostatic landscape of plant endomembranes. Dev Cell 45: 465–480

Panter PE, Muranaka T, Cuitun-Coronado D, Graham CA, Yochikawa A, Kudoh H, Dodd AN (2019) Circadian regulation of the plant transcriptome under natural conditions. Front Genet 10: 1239

Park HJ, Kim WY, Yun DJ (2016) A new insight of salt stress signaling in plant. Mol Cells 39: 447–459

Pascut FC, Couvreur V, Dietrich D, Leftley N, Reyt G, Boursiac Y, Calvo-Polanco M, Casimiro I, Maurel C, Salt DE, et al. (2021) Non-invasive hydrodynamic imaging in plant roots at cellular resolution. Nat Commun 12: 4682

Peret B, Li GW, Zhao J, Band LR, Voss U, Postaire O, Luu DT, Da Ines O, Casimiro I, Lucas M, et al. (2012) Auxin regulates aquaporin function to facilitate lateral root emergence. Nat Cell Biol 14: 991

Pivovaroff AL, Sack L, Santiago LS (2014) Coordination of stem and leaf hydraulic conductance in southern California shrubs: a test of the hydraulic segmentation hypothesis. New Phytol 203: 842–850

Postaire O, Tournaire-Roux C, Grondin A, Boursiac Y, Morillon R, Schäffner AR, Maurel C (2010) A PIP1 aquaporin contributes to hydrostatic pressure-induced water transport in both the root and rosette of Arabidopsis. Plant Physiol 152: 1418–1430

Prado K, Cotelle V, Li G, Bellati J, Tang N, Tournaire-Roux C, Martinière A, Santoni V, Maurela C (2019) Oscillating aquaporin phosphorylation and 14-3-3 proteins mediate the circadian regulation of leaf hydraulics. Plant Cell 31: 417–429

Prasch CM, Sonnewald U (2013) Simultaneous application of heat, drought, and virus to Arabidopsis plants reveals significant shifts in signaling networks. Plant Physiol 162: 1849–1866

Prats KA, Brodersen CR (2021) Desiccation and rehydration dynamics in the epiphytic resurrection fern Pleopeltis polypodioides. Plant Physiol 187: 1501–1518

Procko C, Murthy S, Keenan WT, Mousavi SAR, Dabi T, Coombs A, Procko E, Baird L, Patapoutian A, Chory J (2021) Stretch-activated ion channels identified in the touch-sensitive structures of carnivorous Droseraceae plants. Elife 10: e64250

Provart NJ, Alonso J, Assmann SM, Bergmann D, Brady SM, Brkljacic J, Browse J, Chapple C, Colot V, Cutler S et al. (2016) 50 years of Arabidopsis research: highlights and future directions. New Phytol 209: 921–944

Qiu JE, Mcgaughey SA, Groszmann M, Tyerman SD, Byrt CS (2020) Phosphorylation influences water and ion channel function ofAtPIP2;1. Plant Cell and Environ 43: 2428–2442

Quint M, Delker C, Franklin KA, Wigge PA, Halliday KJ, Van Zanten M (2016) Molecular and genetic control of plant thermomorphogenesis. Nat Plants 2: 15190

Raschke K (1975) Stomatal action. Ann Rev Plant Physiol 26: 309–340

Raschke K, Shabalhang M, Wolf R (2003) The slow and the quick anion conductance in whole guard cells: their voltage-dependent alternation, and the modulation of their activities by abscisic acid and CO_2. Planta 217: 639–650

Rasmussen S, Barah P, Suarez-Rodriguez MC, Bressendorff S, Friis P, Costantino P, Bones AM, Nielsen HB, Mundy J (2013) Transcriptome responses to combinations of stresses in Arabidopsis. Plant Physiol 161: 1783–1794

Reichardt S, Piepho HP, Stintzi A, Schaller A (2020) Peptide signaling for drought-induced tomato flower drop. Science 367: 1482

Reiser V, Raitt DC, Saito H (2003) Yeast osmosensor Sln1 and plant cytokinin receptor Cre1 respond to changes in turgor pressure. J Cell Biol 161: 1035–1040

Ren YB, Miao M, Meng Y, Cao JS, Fan TT, Yue JY, Xiao FM, Liu YS, Cao SQ (2018) DFR1-mediated inhibition of proline degradation pathway regulates drought and freezing tolerance in Arabidopsis. Cell Rep 23: 3960–3974

Reynoso MA, Borowsky AT, Pauluzzi GC, Yeung E, Zhang J, Formentin E, Velasco J, Cabanlit S, Duvenjian C, Prior MJ, et al.

(2022) Gene regulatory networks shape developmental plasticity of root cell types under water extremes. Dev Cell 57: 1177–1192.e6

Reynoso MA, Kajala K, Bajic M, West DA, Pauluzzi G, Yao AI, Hatch K, Zumstein K, Woodhouse M, Rodriguez-Medina J, et al. (2019) Evolutionary flexibility in flooding response circuitry in angiosperms. Science 365: 1291–1295

Riboni M, Galbiati M, Tonelli C, Conti L (2013) GIGANTEA enables drought escape response via abscisic acid-dependent activation of the florigens and SUPPRESSOR OF OVEREXPRESSION OF CONSTANS1. Plant Physiol 162: 1706–1719

Riboni M, Robustelli Test A, Galbiati M, Tonelli C, Conti L (2016) ABA-dependent control of GIGANTEA signalling enables drought escape via up-regulation of FLOWERING LOCUS T in Arabidopsis thaliana. J Exp Bot 67: 6309–6322

Rivero RM, Mittler R, Blumwald E, Zandalinas SI (2022) Developing climate-resilient crops: improving plant tolerance to stress combination. Plant J 109: 373–389

Riggi M, Bourgoint C, Macchione M, Matile S, Loewith R, Roux A (2019) TORC2 controls endocytosis through plasma membrane tension. J Cell Biol 218: 2265–2276

Rizhsky L, Liang H, Shuman J, Shulaev V, Davletova S, Mittler R (2004) When defense pathways collide. The response of Arabidopsis to a combination of drought and heat stress. Plant Physiol 134: 1683–1696

Robbins NE, 2nd, Dinneny JR (2015) The divining root: moisture-driven responses of roots at the micro- and macro-scale. J Exp Bot 66: 2145–2154

Robbins NE, 2nd, Dinneny JR (2018) Growth is required for perception of water availability to pattern root branches in plants. Proc Natl Acad Sci USA 115: E822–E831

Rockwell FE, Holbrook NM, Stroock AD (2014) The competition between liquid and vapor transport in transpiring leaves. Plant Physiol 164: 1741–1758

Rodrigues O, Reshetnyak G, Grondin A, Saijo Y, Leonhardt N, Maurel C, Verdoucq L (2017) Aquaporins facilitate hydrogen peroxide entry into guard cells to mediate ABA- and pathogen-triggered stomatal closure. Proc Natl Acad Sci USA 114: 9200–9205

Rodriguez-Dominguez CM, Brodribb TJ (2020) Declining root water transport drives stomatal closure in olive under moderate water stress. New Phytol 225: 126–134

Roff DA, Fairbairn DJ (2007) The evolution of trade-offs: where are we? J Evol Biol 20: 433–447

Romero-Montepaone S, Sellaro R, Esteban Hernando C, Costigliolo-Rojas C, Bianchimano L, Ploschuk EL, Yanovsky MJ, Casal JJ (2021) Functional convergence of growth responses to shade and warmth in Arabidopsis. New Phytol 231: 1890–1905

Sabbioni G, Funck D, Forlani G (2021) Enzymology and regulation of D^1-pyrroline-5-carboxylate synthetase 2 from rice. Front Plant Sci 12: 672702

Sack L (2004) Responses of temperate woody seedlings to shade and drought: do trade-offs limit potential niche differentiation? Oikos 107: 110–127

Sack L, Cowan PD, Jaikumar N, Holbrook NM (2003) The 'hydrology' of leaves: co-ordination of structure and function in temperate woody species. Plant Cell Environ 26: 1343–1356

Sack L, John GP, Buckley TN (2018) ABA accumulation in dehydrating leaves is associated with decline in cell volume, not turgor pressure. Plant Physiol 176: 489–495

Saltz JB, Hessel FC, Kelly MW (2017) Trait correlations in the genomics era. Trends Ecol Evol 32: 279–290

Sanchez DH (2013) Physiological and biotechnological implications of transcript-level variation under abiotic stress. Plant Biol 15: 925–930

Sanchez SE, Kay SA (2016) The plant circadian clock: from a simple timekeeper to a complex developmental manager. Cold Spring Harb Perspect Biol 8: a027748

Sawyer LA, Hennessy JM, Peixoto AA, Rosato E, Parkinson H, Costa R, Kyriacou CP (1997) Natural variation in a Drosophila clock gene and temperature compensation. Science **278**: 2117–2120

Schenk HJ, Espino S, Goedhart CM, Nordenstahl M, Cabrera HIM, Jones CS (2008) Hydraulic integration and shrub growth form linked across continental aridity gradients. Proc Natl Acad Sci USA **105**: 11248–11253

Schenk HJ, Jansen S, Holtta T (2021) Positive pressure in xylem and its role in hydraulic function. New Phytol **230**: 27–45

Scholz P, Pejchar P, Fernkorn M, Škrabálková E, Pleskot R, Blersch K, Munnik T, Potocký M, Ischebeck T (2022) DIACYLGLYCEROL KINASE 5 regulates polar tip growth of tobacco pollen tubes. New Phytol **233**: 2185–2202

Schulze ED, Hall AE (1982) Stomatal responses, water loss and CO_2 assimilation rates of plants in contrasting environments. *In* OL Lange, PS Nobel, CB Osmond, H Ziegler, eds, Physiological Plant Ecology II: Water Relations and Carbon Assimilation. Springer Berlin Heidelberg, Berlin, Heidelberg, pp 181–230

Scoffoni C, Sack L (2017) The causes and consequences of leaf hydraulic decline with dehydration. J Exp Bot **68**: 4479–4496

Scoffoni C, Vuong C, Diep S, Cochard H, Sack L (2014) Leaf shrinkage with dehydration: coordination with hydraulic vulnerability and drought tolerance. Plant Physiol **164**: 1772–1788

Scoffoni C, Albuquerque C, Brodersen CR, Townes SV, John GP, Bartlett MK, Buckley TN, McElrone AJ, Sack L (2017) Outside-xylem vulnerability, not xylem embolism, controls leaf hydraulic decline during dehydration. Plant Physiol **173**: 1197–1210

Senthil-Kumar M, Mysore KS (2012) Ornithine-delta-aminotransferase and proline dehydrogenase genes play a role in non-host disease resistance by regulating pyrroline-5-carboxylate metabolism-induced hypersensitive response. Plant Cell Environ **35**: 1329–1343

Seo PJJ, Mas P (2015) STRESSing the role of the plant circadian clock. Plant Sci **20**: 230–237

Sewelam N, Oshima Y, Mitsuda N, Ohme-Takagi M (2014) A step towards understanding plant responses to multiple environmental stresses: a genome-wide study. Plant Cell Environ **37**: 2024–2035

Sharma M, Kumar P, Verma V, Sharma R, Bhargava B, Irfan M (2022) Understanding plant stress memory response for abiotic stress resilience: molecular insights and prospects. Plant Physiol Biochem **179**: 10–24

Sharma S, Villamor JG, Verslues PE (2011) Essential role of tissue-specific proline synthesis and catabolism in growth and redox balance at low water potential. Plant Physiol **157**: 292–304

Shaw RG, Etterson JR (2012) Rapid climate change and the rate of adaptation: insight from experimental quantitative genetics. New Phytol **195**: 752–765

Shinde S, Villamor JG, Lin WD, Sharma S, Verslues PE (2016) Proline coordination with fatty acid synthesis and redox metabolism of chloroplast and mitochondria. Plant Physiol **172**: 1074–1088

Shindo C, Aranzana MJ, Lister C, Baxter C, Nicholls C, Nordborg M, Dean C (2005) Role of FRIGIDA and FLOWERING LOCUS C in determining variation in flowering time of Arabidopsis. Plant Physiol **138**: 1163–1173

Shiono K, Yoshikawa M, Kreszies T, Yamada S, Hojo Y, Matsuura T, Mori IC, Schreiber L, Yoshioka T (2022) Abscisic acid is required for exodermal suberization to form a barrier to radial oxygen loss in the adventitious roots of rice (*Oryza sativa*). New Phytol **233**: 655–669

Shipley B, Meziane D (2002) The balanced-growth hypothesis and the allometry of leaf and root biomass allocation. Funct Ecol **16**: 326–331

Signorelli S (2016) The fermentation analogy: a point of view for understanding the intriguing role of proline accumulation in stressed plants. Front Plant Sci **7**: 1339

Simmons CR, Lafitte HR, Reimann KS, Brugière N, Roesler K, Albertsen MC, Greene TW, Habben JE (2021) Successes and insights of an industry biotech program to enhance maize agronomic traits. Plant Sci **307**: 110899

Simon ML, Platre MP, Assil S, van Wijk R, Chen WY, Chory J, Dreux M, Munnik T, Jaillais Y (2014) A multi-colour/multi-affinity marker set to visualize phosphoinositide dynamics in Arabidopsis. Plant J **77**: 322–337

Sinervo B, Svensson E (2002) Correlational selection and the evolution of genomic architecture. Heredity **89**: 329–338

Slatyer RO, Taylor SA (1960) Terminology in plant– and soil–water relations. Nature **187**: 922–924

Smith-Martin CM, Skelton RP, Johnson KM, Lucani C, Brodribb TJ (2020) Lack of vulnerability segmentation among woody species in a diverse dry sclerophyll woodland community. Funct Ecol **34**: 777–787

Soma F, Takahashi F, Suzuki T, Shinozaki K, Yamaguchi-Shinozaki K (2020) Plant Raf-like kinases regulate the mRNA population upstream of ABA-unresponsive SnRK2 kinases under drought stress. Nat Commun **11**: 1373

Stuart TS (1968) Revival of respiration and photosynthesis in dried leaves of polypodium polypodiodes. Planta **83**: 185

Suda H, Mano H, Toyota M, Fukushima K, Mimura T, Tsutsui I, Hedrich R, Tamada Y, Hasebe M (2020) Calcium dynamics during trap closure visualized in transgenic Venus flytrap. Nat Plants **6**: 1219–1224

Sun Y, Oh DH, Duan L, Ramachandran P, Ramirez A, Bartlett A, Dassanayake M, Dinneny JR (2022) Divergence in a stress regulatory network underlies differential growth control. Nat Plants **8**: 549–560

Székely G, Ábrahám E, Cséplő Á, Rigó G, Zsigmond L, Csiszár J, Ayaydin F, Strizhov N, Jásik J, Schmelzer E, et al. (2008) Duplicated P5CS genes of Arabidopsis play distinct roles in stress regulation and developmental control of proline biosynthesis. Plant J **53**: 11–28

Szoke A, Miao GH, Hong ZL, Verma DPS (1992) Subcellular location of Δ^1- pyrroline-5-carboxylate reductase in root nodule and leaf of soybean. Plant Physiol **99**: 1642–1649

Takahashi F, Suzuki T, Osakabe Y, Betsuyaku S, Kondo Y, Dohmae N, Fukuda H, Yamaguchi-Shinozaki K, Shinozaki K (2018) A small peptide modulates stomatal control via abscisic acid in long-distance signalling. Nature **556**: 235

Tarashansky AJ, Musser JM, Khariton M, Li P, Arendt D, Quake SR, Wang B (2021) Mapping single-cell atlases throughout Metazoa unravels cell type evolution. Elife **10**: e66747

Tejos R, Sauer M, Vanneste S, Palacios-Gomez M, Li H, Heilmann M, van Wijk R, Vermeer JEM, Heilmann I, Munnik T, et al. (2014) Bipolar plasma membrane distribution of phosphoinositides and their requirement for auxin-mediated cell polarity and patterning in Arabidopsis. Plant Cell **26**: 2114–2128

Thenkabail PS, Teluguntla PG, Xiong J, Oliphant A, Congalton RG, Ozdogan M, Gumma MK, Tilton JC, Giri C, Milesi C, et al. (2021) Global cropland-extent product at 30 pm resolution (GCEP30) derived from Landsat satellite time-series data for the year 2015 using multiple machine-learning algorithms on Google Earth Engine cloud: U.S. Geological Survey Professional Paper 1868, p 63

Tõldsepp K, Zhang J, Takahashi Y, Sindarovska Y, Hõrak H, Ceciliato P, Koolmeister K, Wang Y, Vaahtera L, Jakobson L, et al. (2018) Mitogen-activated protein kinases MPK4 and MPK12 are key components mediating CO_2-induced stomatal movements. Plant J **96**: 1018–1035

Tomkins M, Hughes A, Morris RJ (2021) An update on passive transport in and out of plant cells. Plant Physiol **187**: 1973–1984

Törnroth-Horsefield S, Wang Y, Hedfalk K, Johanson U, Karlsson M, Tajkhorshid E, Neutze R, Kjellbom P (2006) Structural mechanism of plant aquaporin gating. Nature **439**: 688–694

Tournaire-Roux C, Sutka M, Javot H, Gout E, Gerbeau P, Luu DT, Bligny R, Maurel C (2003) Cytosolic pH regulates root water

transport during anoxic stress through gating of aquaporins. Nature **425**: 393–397

Tran LSP, UraoT, QinF, Maruyama K, Kakimoto T, Shinozaki K, Yamaguchi-Shinozaki K (2007) Functional analysis of AHK1/ATHK1 and cytokinin receptor histidine kinases in response to abscisic acid, drought, and salt stress in Arabidopsis. Proc Natl Acad Sci USA **104**: 20623–20628

Tsuda M, Tyree MT (1997) Whole-plant hydraulic resistance and vulnerability segmentation in Acer saccharinum. Tree Physiol **17**: 351–357

Tsuda M, Tyree MT (2000) Plant hydraulic conductance measured by the high pressure flow meter in crop plants. J Exp Bot **51**: 823–828

Tyerman SD, Mcgaughey SA, Qiu JE, Yool AJ, Byrt CS (2021) Adaptable and multifunctional ion-conducting aquaporins. *In* SS Merchant, ed, Annual Review of Plant Biology, Vol 72. Palo Alto, CA, pp 703–736

Tyree MT, Zimmermann MH (2002) Xylem Structure and the Ascent of Sap. Springer, Heidelberg, Germany

Toyota M, Spencer D, Sawai-Toyota S, Jiaqi W, Zhang T, Koo AJ, Howe GA, Gilroy S (2018) Glutamate triggers long-distance, calcium-based plant defense signaling. Science **361**: 1112–1115

Uga Y, Sugimoto K, Ogawa S, Rane J, Ishitani M, Hara N, Kitomi Y, Inukai Y, Ono K, Kanno N, et al. (2013) Control of root system architecture by DEEPER ROOTING 1 increases rice yield under drought conditions. Nat Genet **45**: 1097–1102

Umezawa T, Nakashima K, Miyakawa T, Kuromori T, Tanokura M, Shinozaki K, Yamaguchi-Shinozaki K (2010) Molecular basis of the core regulatory network in abscisic acid responses: sensing, signaling, and transport. Plant Cell Physiol **51**: 1821–1839

Umezawa T, Sugiyama N, Mizoguchi M, Hayashi S, Myouga F, Yamaguchi-Shinozaki K, Ishihama Y, Hirayama T, Shinozaki K (2009) Type 2C protein phosphatases directly regulate abscisic acid-activated protein kinases in Arabidopsis. Proc Natl Acad Sci USA **106**: 17588–17593

Undurraga SF, Press MO, Legendre M, Bujdoso N, Bale J, Wang H, Davis SJ, Verstrepen KJ, Queitsch C (2012) Background-dependent effects of polyglutamine variation in the *Arabidopsis thaliana* gene ELF3. Proc Natl Acad Sci USA **109**: 19363–19367

Ursache R, Andersen TG, Marhavý P, Geldner N (2018) A protocol for combining fluorescent proteins with histological stains for diverse cell wall components. Plant J **93**: 399–412

van Dongen JT, Licausi F (2015) Oxygen sensing and signaling. Annu Rev Plant Biol **66**: 345–367

Van den Honert TH (1948) Water transport in plants as a catenary process. Trans Faraday Soc **44**: 146–153

van Leeuwen W, Vermeer JEM, Gadella Jr TWJ, Munnik, T (2007) Visualisation of phosphatidylinositol 4,5-bisphosphate in the plasma membrane of suspension-cultured tobacco BY-2 cells and whole Arabidopsis seedlings. Plant J **52**: 1014–1026

van Wijk R, Zhang Q, Zarza X, Lamers M, Reyes-Marquez F, Guardia A, Scuffi D, García-Mata C, Ligterink W, Haring MA, et al. (2018) Role for Arabidopsis PLC7 in stomatal movement, seed mucilage attachment, and leaf serration. Front Plant Sci **9**: 1721

Vermeer JEM, van Wijk R, Goedhart J, Geldner N, Chory J, Gadella Jr TWJ, Munnik T (2017) Imaging diacylglycerol in the cytosolic leaflet of plant membranes. Plant Cell Physiol **58**: 1196–1207

Verbraeken L, Wuyts N, Mertens S, Cannoot B, Maleux K, Demuynck K, De Block J, Merchie J, Dhondt S, Bonaventure G, et al. (2021) Drought affects the rate and duration of organ growth but not inter-organ growth coordination. Plant Physiol **186**: 1336–1353

Verslues PE, Agarwal M, Katiyar-Agarwal S, Zhu J, Zhu JK (2006) Methods and concepts in quantifying resistance to drought, salt and freezing, abiotic stresses that affect plant water status. Plant J **45**: 523–539

Vlad F, Rubio S, Rodrigues A, Sirichandra C, Belin C, Robert N, Leung J, Rodriguez PL, Lauriere C, Merlot S (2009) Protein phosphatases 2C regulate the activation of the Snf1-related kinase OST1 by abscisic acid in Arabidopsis. Plant Cell **21**: 3170–3184

Wallace JG, Rodgers-Melnick E, Buckler ES (2018) On the road to Breeding 4.0: unraveling the good, the bad, and the boring of crop quantitative genomics. Ann Rev Genet **52**: 421–444

Walsh B, Blows MW (2009) Abundant genetic variation + strong selection = multivariate genetic constraints: a geometric view of adaptation. Ann Rev Ecol Evol Syst **40**: 41–59

Wang C, Hu H, Qin X, Zeise B, Xu D, Rappel WJ, Boron WF, Schroeder JI (2016) Reconstitution of CO_2 regulation of SLAC1 anion channel and function of CO_2-permeable PIP2;1 aquaporin as CARBONIC ANHYDRASE4 interactor. Plant Cell **28**: 568–582

Wang P, Zhao Y, Li Z, Hsu C-C, Liu X, Fu L, Hou Y-J, Du Y, Xie S, Zhang C et al. (2018a) Reciprocal regulation of the TOR kinase and ABA receptor balances plant growth and stress response. Mol Cell **69**: 100–112.e6

Wang P, Xue L, Batelli G, Lee S, Hou YJ, van Oosten MJ, Zhang H, Tao WA, Zhu JK (2013a) Quantitative phosphoproteomics identifies SnRK2 protein kinase substrates and reveals the effectors of abscisic acid action. Proc Natl Acad Sci USA **110**: 11205–11210

Wang S, Zhang Y, Ju W, Chen JM, Ciais P, Cescatti A, Sardans J, Janssens IA, Wu M, Berry JA, et al. (2020a) Recent global decline of CO(2) fertilization effects on vegetation photosynthesis. Science **370**: 1295–1300

Wang Y, Li L, Ye T, Lu Y, Chen X, Wu Y (2013b) The inhibitory effect of ABA on floral transition is mediated by ABI5 in Arabidopsis. J Exp Bot **64**: 675–684

Wang Y, Zhao ZJ, Liu F, Sun LR, Hao FS (2020b) Versatile roles of aquaporins in plant growth and development. Int J Mol Sci **21**: 28

Wang Z, Wang F, Hong Y, Yao J, Ren Z, Shi H, Zhu JK (2018b) The flowering repressor SVP confers drought resistance in Arabidopsis by regulating abscisic acid catabolism. Mol Plant **11**: 1184–1197

Wang ZY, Xiong L, Li W, Zhu JK, Zhu J (2011) The plant cuticle is required for osmotic stress regulation of abscisic acid biosynthesis and osmotic stress tolerance in Arabidopsis. Plant Cell **23**: 1971–1984

Weiss M, Weiss M (2019) An assessment of threats to the American power grid. Energy Sustain Soc **9**: 18

Willi Y, Van Buskirk J (2022) A review on trade-offs at the warm and cold ends of geographical distributions. Phil Trans R Soc B Biol Sci **377**: 20210022

Wilson ME, Basu MR, Bhaskara GB, Verslues PE, Haswell ES (2014) Plastid hypo-osmotic stress activates cellular osmotic stress responses. Plant Physiol **165**: 119–128

Xu J, Misra G, Sreenivasulu N, Henry A (2021) What happens at night? Physiological mechanisms related to maintaining grain yield under high night temperature in rice. Plant Cell Environ **44**: 2245–2261

Yamauchi T, Tanaka A, Inahashi H, Nishizawa NK, Tsutsumi N, Inukai Y, Nakazono M (2019) Fine control of aerenchyma and lateral root development through AUX/IAA- and ARF-dependent auxin signaling. Proc Natl Acad Sci USA **116**: 20770–20775

Yamauchi T, Tanaka A, Mori H, Takamure I, Kato K, Nakazono M (2016) Ethylene-dependent aerenchyma formation in adventitious roots is regulated differently in rice and maize. Plant Cell Environ **39**: 2145–2157

Yamauchi T, Yoshioka M, Fukazawa A, Mori H, Nishizawa NK, Tsutsumi N, Yoshioka H, Nakazono M (2017) An NADPH oxidase RBOH functions in rice roots during lysigenous aerenchyma formation under oxygen-deficient conditions. Plant Cell **29**: 775–790

Yang J, Cao Y, Ma L (2021) Co-transcriptional RNA processing in plants: exploring from the perspective of polyadenylation. Int J Mol Sci **22**: 3300

Yang Y, Li Y, Sancar A, Oztas O (2020) The circadian clock shapes the Arabidopsis transcriptome by regulating alternative splicing and alternative polyadenylation. J Biol Chem **295**: 7608–7619

Yang Z, Liu J, Poree F, Schaeufele R, Helmke H, Frackenpohl J, Lehr S, von Koskull-Doring P, Christmann A, Schnyder H, et al. (2019) Abscisic acid receptors and coreceptors modulate plant water use efficiency and water productivity. Plant Physiol **180**: 1066–1080

Yamaguchi-Shinozaki K, Shinozaki K (2006) Transcriptional regulatory networks in cellular responses and tolerance to dehydration and cold stresses. Annu Rev Plant Biol **57**: 781–803

Ye H, Song L, Chen H, Valliyodan B, Cheng P, Ali L, Vuong T, Wu C, Orlowski J, Buckley B, et al. (2018) A major natural genetic variation associated with root system architecture and plasticity improves waterlogging tolerance and yield in soybean. Plant Cell Environ **41**: 2169–2182

Yoshida T, Christmann A, Yamaguchi-Shinozaki K, Grill E, Fernie AR (2019a) Revisiting the basal role of ABA—roles outside of stress. Trends Plant Sci **24**: 625–635

Yoshida T, Fujita Y, Maruyama K, Mogami J, Todaka D, Shinozaki K, Yamaguchi-Shinozaki K (2014) Four Arabidopsis AREB/ABF transcription factors function predominantly in gene expression downstream of SnRK2 kinases in abscisic acid signalling in response to osmotic stress. Plant Cell Environ **38**: 35–49

Yoshida T, Obata T, Feil R, Lunn JE, Fujita Y, Yamaguchi-Shinozaki K, Fernie AR (2019b) The role of abscisic acid signaling in maintaining the metabolic balance required for Arabidopsis growth under nonstress conditions. Plant Cell **31**: 84–105

Zandalinas SI, Fritschi FB, Mittler R (2021) Global warming, climate change, and environmental pollution: recipe for a multifactorial stress combination disaster. Trends Plant Sci **26**: 588–599

Zarza X, van Wijk R, Shabala L, Hunkeler A, Lefebvre M, Rodriguez-Villalón A, Shabala S, Tiburcio AF, Heilmann I, Munnik T (2020) Lipid kinases PIP5K7 and PIP5K9 are required for polyamine-triggered K^+ efflux in Arabidopsis roots. Plant J **104**: 416–432

Zeppel MJB, Harrison SP, Adams HD, Kelley DI, Li G, Tissue DT, Dawson TE, Fensham R, Medlyn BE, Palmer A, et al. (2015) Drought and resprouting plants. New Phytol **206**: 583–589

Zhang H, Zhu J, Gong Z, Zhu JK (2022) Abiotic stress responses in plants. Nat Rev Genet **23**: 104–119

Zhang J, De-Oliveira-Ceciliato P, Takahashi Y, Schulze S, Dubeaux G, Hauser F, Azoulay-Shemer T, Tõldsepp K, Kollist H, Rappel WJ, et al. (2018a) Insights into the molecular mechanisms of CO_2-mediated regulation of stomatal movements. Curr Biol **28**: R1356–R1363

Zhang J, Wang N, Miao Y, Hauser F, McCammon JA, Rappel WJ, Schroeder JI (2018b) Identification of SLAC1 anion channel residues required for regulation of stomatal movements. Proc Natl Acad Sci USA **115**: 11129–11137

Zhang L, Becker DF (2015) Connecting proline metabolism and signaling pathways in plant senescence. Front Plant Sci **6**: 552

Zhang L, Takahashi Y, Hsu PK, Kollist H, Merilo E, Krysan PJ, Schroeder JI (2020) FRET kinase sensor development reveals SnRK2/OST1 activation by ABA but not by MeJA and high CO_2 during stomatal closure. eLife **9**: e56351

Zhang Q, van Wijk R, Shahbaz M, Roels W, van Schooten B, Vermeer J, Zarza X, Guardia A, Scuffi D, García-Mata C, et al. (2018c) Arabidopsis phospholipase C3 is involved in lateral root initiation and ABA responses in seed germination and stomatal closure. Plant Cell Physiol **59**: 469–486

Zhang Q, van Wijk R, Zarza X, Shahbaz M, van Hooren M, Guardia A, Scuffi D, García-Mata C, van den Ende W, Hoffmann-Benning S, et al. (2018d) Knock-down of Arabidopsis PLC5 reduces primary root growth and secondary root formation while overexpression improves drought tolerance and causes stunted root hair growth. Plant Cell Physiol **59**: 2004–2019

Zhang S, Feng M, Chen W, Zhou XF, Lu JY, Wang YR, Li YH, Jiang CZ, Gan SS, Ma N, et al. (2019) In rose, transcription factor PTM balances growth and drought survival via PIP2;1 aquaporin. Nat Plants **5**: 290–299

Zheng HJ, Wu AZ, Zheng CC, Wang YF, Cai R, Shen XF, Xu RR, Liu P, Kong LJ, Dong ST (2009) QTL mapping of maize (*Zea mays*) stay-green traits and their relationship to yield. Plant Breed **128**: 54–62

Zheng Y, Cabassa-Hourton C, Planchais S, Lebreton S, Savoure A (2021) The proline cycle as an eukaryotic redox valve. J Exp Bot **72**: 6856–6866

Zhou X, Lu J, Zhang Y, Guo J, Lin W, Van Norman JM, Qin Y, Zhu X, Yang Z (2021) Membrane receptor-mediated mechano-transduction maintains cell integrity during pollen tube growth within the pistil. Dev Cell **56**: 1030–1042

Zhu JK (2002) Salt and drought stress signal transduction in plants. Annu Rev Plant Biol **53**: 247–273

Zhu SD, Liu H, Xu QY, Cao KF, Ye Q (2016a) Are leaves more vulnerable to cavitation than branches? Funct Ecol **30**: 1740–1744

Zhu Y, Klasfeld S, Jeong CW, Jin R, Goto K, Yamaguchi N, Wagner D (2020) TERMINAL FLOWER 1-FD complex target genes and competition with FLOWERING LOCUS T. Nat Commun **11**: 5118

Zhu Z, Piao S, Myneni RB, Huang M, Zeng Z, Canadell JG, Ciais P, Sitch S, Friedlingstein P, Arneth A (2016b) Greening of the Earth and its drivers. Nat Climate Change **6**: 791–795