



# Assessing breast density using the chemical-shift encoding-based proton density fat fraction in 3-T MRI

Tabea Borde<sup>1</sup> · Mingming Wu<sup>1</sup> · Stefan Ruschke<sup>1</sup> · Christof Boehm<sup>1</sup> · Jonathan Stelter<sup>1</sup> · Kilian Weiss<sup>2</sup> · Stephan Metz<sup>1</sup> · Marcus Richard Makowski<sup>1</sup> · Dimitrios C. Karampinos<sup>1</sup> · Eva Maria Fallenberg<sup>1</sup>

Received: 19 May 2022 / Revised: 22 November 2022 / Accepted: 29 November 2022 / Published online: 20 December 2022  
© The Author(s) 2022

## Abstract

**Objectives** There is a clinical need for a non-ionizing, quantitative assessment of breast density, as one of the strongest independent risk factors for breast cancer. This study aims to establish proton density fat fraction (PDFF) as a quantitative biomarker for fat tissue concentration in breast MRI and correlate mean breast PDFF to mammography.

**Methods** In this retrospective study, 193 women were routinely subjected to 3-T MRI using a six-echo chemical shift encoding-based water-fat sequence. Water-fat separation was based on a signal model accounting for a single  $T_2^*$  decay and a pre-calibrated 7-peak fat spectrum resulting in volumetric fat-only, water-only images, PDFF- and  $T_2^*$ -values. After semi-automated breast segmentation, PDFF and  $T_2^*$  values were determined for the entire breast and fibroglandular tissue. The mammographic and MRI-based breast density was classified by visual estimation using the American College of Radiology Breast Imaging Reporting and Data System categories (ACR A-D).

**Results** The PDFF negatively correlated with mammographic and MRI breast density measurements (Spearman rho:  $-0.74$ ,  $p < .001$ ) and revealed a significant distinction between all four ACR categories. Mean  $T_2^*$  of the fibroglandular tissue correlated with increasing ACR categories (Spearman rho:  $0.34$ ,  $p < .001$ ). The PDFF of the fibroglandular tissue showed a correlation with age (Pearson rho:  $0.56$ ,  $p = .03$ ).

**Conclusion** The proposed breast PDFF as an automated tissue fat concentration measurement is comparable with mammographic breast density estimations. Therefore, it is a promising approach to an accurate, user-independent, and non-ionizing breast density assessment that could be easily incorporated into clinical routine breast MRI exams.

## Key Points

- *The proposed PDFF strongly negatively correlates with visually determined mammographic and MRI-based breast density estimations and therefore allows for an accurate, non-ionizing, and user-independent density measurement.*
- *In combination with  $T_2^*$ , the PDFF can be used to track structural alterations in the composition of breast tissue for an individualized risk assessment for breast cancer.*

**Keywords** Breast · Breast density · Magnetic resonance imaging

---

Dimitrios C. Karampinos and Eva Maria Fallenberg contributed equally.

✉ Tabea Borde  
tabea.borde@tum.de

<sup>1</sup> Department of Diagnostic and Interventional Radiology, Klinikum rechts der Isar, TUM School of Medicine, Technical University of Munich, Ismaninger Str. 22, 81675 Munich, Germany

<sup>2</sup> Philips GmbH, Hamburg, Germany

## Abbreviations

ACR	American College of Radiology
BI-RADS®	Breast Imaging Reporting and Data System
FOV	Field of view
ICC	Intraclass correlation coefficient
PDFF	Proton density fat fraction
SENSE	Sensitivity encoding

## Introduction

Breast cancer accounts for almost 25% of all diagnosed cancers in women and is the leading cause of cancer-related deaths among women worldwide [1]. In both screen-detected and symptomatic patients, high breast density has been identified as one of the strongest independent risk factors and is associated with a twofold increase in breast cancer incidence rates [2–4].

In clinical practice, the breast density is assessed in mammograms by visual estimation of the content of radiopaque fibroglandular parenchyma within the breast and classified according to the American College of Radiology (ACR) categories under the Breast Imaging Reporting and Data System® (BI-RADS®, 5<sup>th</sup> edition) in 4 grades: (A) almost entirely fatty, (B) scattered fibroglandular densities, (C) heterogeneously dense, which may obscure small masses, and (D) extremely dense, which lowers the sensitivity of mammography [5]. However, several limitations lead to a non-consistent intra- and inter-reader variability of mammography-based breast density measurements. First, the technical execution requires a compression of the breast which is then exposed to low-dose ionizing radiation. Slight deviations in tissue compression or radiation exposure calibration may already confound breast density measurements [6]. Furthermore, the resulting two-dimensional projection of the volumetric body has a limited capacity to provide objective proportions of the breast composition [4]. Altogether, these factors combined with the non-neglectable, detrimental exposure to ionizing radiation, especially at younger age, challenge the accuracy and reliability of mammography in assessing breast density and cancer risk prediction.

Magnetic resonance imaging (MRI) presents a suitable alternative for an objective, volumetric quantification of the structural composition of the breast without the use of ionizing radiation. Standard T<sub>1</sub>-weighted imaging techniques have been primarily used to separate adipose and fibroglandular breast tissue [7, 8]. Recent studies using chemical shift encoding-based water-fat separation (Dixon imaging) improved the contrast in separating fat from water proton signals in the breast. These approaches yielded favorable results in the differentiation of adipose and fibroglandular breast tissue and the measurement of a signal-weighted breast fat fraction but are dependent on the imaging protocol and the breast density segmentation processing pipeline [9–11]. A promising quantitative biomarker for the tissue fat concentration is the water-fat MRI-based proton density fat fraction (PDFF). The PDFF is defined as the ratio of density of triglyceride protons to the total density of triglyceride and water protons and therefore accurately reflects the concentration of fat within that tissue. The PDFF values derive directly from automatically calculated PDFF maps that are insensitive to changes in acquisition parameters which renders the PDFF a comprehensive and clinically practical biomarker [12–14]. In previous studies,

the PDFF has been successfully correlated to different structural tissues such as muscle, pancreas, and liver [15–17] and has been only recently applied for breast density assessment [18, 19].

Therefore, the purpose of this study is to establish the proton density fat fraction as a quantitative biomarker for fat tissue concentration in breast MRI by correlating its outcome to mammography as the most widely used breast density measurement in clinical practice.

## Material and methods

### Study population

This institutional review board–approved retrospective study initially included 300 pre- and postmenopausal women out of which 193 women (median age 48 years, range 29–81 years) had received digital mammography within 1–15 months (median 9) prior to the conductance of the MRI scan from August to November 2020 (Fig. 1). Indication for breast imaging was either screening in high-risk patients, surveillance of cancer patients, or further work up of unclear findings.

### Breast density measurements

Qualitative breast density measurements in two-dimensional mammograms, currently the most widely used breast density estimation in clinical practice, were used as the clinical standard reference. Mammography was performed by using full-field digital mammographic units (GE Healthcare). The visual evaluation of the composition of breast tissue was performed in the craniocaudal and mediolateral-oblique view and classified into the American College of Radiology (ACR) Breast Imaging Reporting and Data System® (BI-RADS®) categories, 5<sup>th</sup>

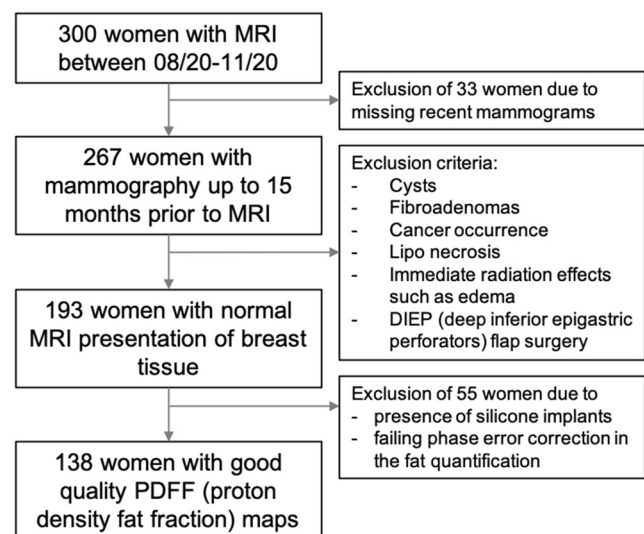


Fig. 1 Study flow chart

edition [20], by the reporting radiologist (4 years of experience in mammography) as well as an independent reader (1 year of experience in mammography) blinded to the study in one session. The inter-reader reliability was assessed using Cohen's kappa.

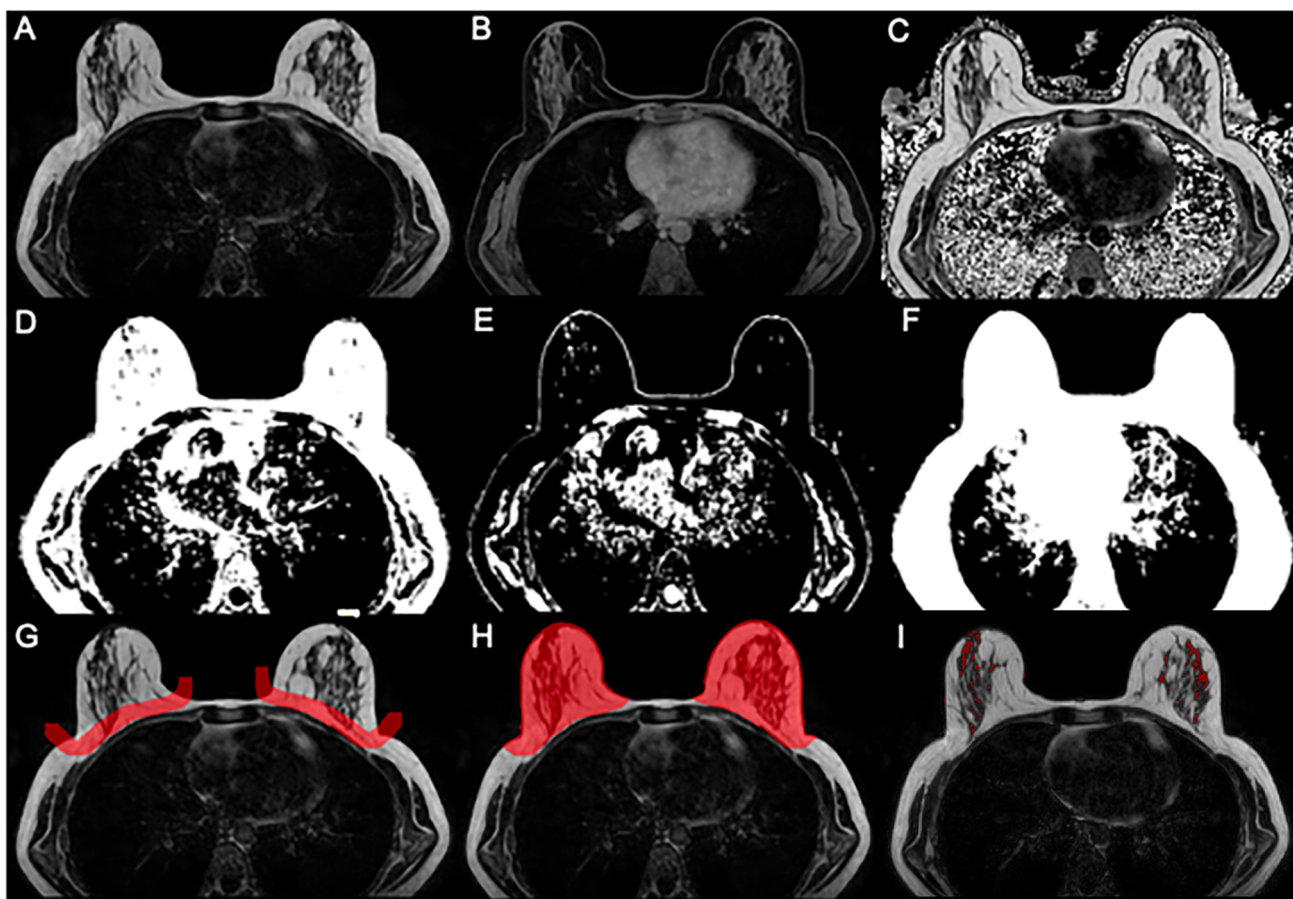
Furthermore, the same visual evaluation was performed in MR T1-weighted images according to the ACR-BIRADS categories by the same reporting and independent radiologists blinded to the results of the mammographic readings.

## MR measurements

Each woman was examined in prone position using a dedicated 7-channel breast radiofrequency (RF) coil in a 3.0-T MRI scanner (Philips Ingenia, Philips Healthcare). In addition to conventional T<sub>1</sub>- and T<sub>2</sub>-weighted sequences, a six-echo 3D spoiled gradient-echo sequence employing bipolar imaging readouts in axial plane was routinely included with the following parameters: 1.7 mm isotropic acquisition voxel size, field of view (FOV, mm): AP = 220, RL = 440, FH = 190, TR/TE/

$\Delta$ TE = 8.8/1.43/1.1, flip angle = 3°, scan time: 2 min 11 s per stack. A combination of sensitivity encoding (SENSE) and compressed sensing was employed based on the vendor's implementation (Compressed SENSE, Philips Healthcare) with an acceleration factor equal to 3. This proton density Dixon sequence was part of the clinical routine protocol to provide artifact-free 3D fat-suppressed images before contrast injection.

After phase error correction, water-fat separation was performed online using the water-fat separation algorithm of the vendor (mDixon Quant, Philips Healthcare). Water-fat separation was performed based on a signal model accounting for a single T<sub>2</sub>\* decay and a pre-calibrated 7-peak fat spectrum accounting for the presence of multiple peaks in the fat spectrum. Resulting volumetric fat-only, water-only images, PDFF maps (Fig. 2), and T<sub>2</sub>\* values were generated. The PDFF maps were automatically computed as the ratio of the fat signal over the sum of fat and water signals. The visual estimation of the breast density in conventional MR in native T1-weighted images



**Fig. 2** Segmentation workflow. **A** Axial MRI fat-separated image. **B** Axial MRI water-separated image. **C** PDFF map. **D–F** Fat, water, and foreground mask as a result of k-means clustering. **G** Manual segmentation delineating the border between the pectoralis muscle and

breast parenchyma. **H** Automated completion of segmentation masks of left and right mask. **I** Final automated segmentation of the fibroglandular tissue

according to the BI-RADS classification was performed by a radiologist with 10 years and a radiologist with 1 year of experience in MR breast readings blinded to the study in one session.

## MR image processing

Breast density analysis was performed on breasts unaffected by cancer, fibroadenomas, cysts, lipo-necrosis after treatment or immediate radiation effects, and without breast augmentation or implantation material to minimize interference with the breast density analysis. Semi-automated breast segmentation was performed by manually delineating the border between the pectoralis muscle and the breast parenchyma on every second slice in fat-only images using the open-source segmentation program ITK-Snap ([www.itksnap.org](http://www.itksnap.org), version 3.8.0, Fig. 2G) [21]. Using the Image processing toolbox (MATLAB R2020b, The MathWorks), k-means clustering was performed, and the FOV was divided into water, fat, and background signal. The manually selected borders were connected with a dilation step and the centroids of the left and right breast were calculated by projecting the manual delineation in AP orientation. Both the centroid position and the border delineation helped separating the breast from the thorax using “bwselect” from the Image processing toolbox. Finally, the mean PDFF over the entire breast and the mean PDFF and  $T_2^*$  over only the fibroglandular tissue of the isolated breasts were automatically extracted. The fibroglandular tissue was segmented using the sharp delineation between fat and fibroglandular tissue as well as a fat-concentration cut-off of 20% as fibroglandular tissue empirically contains very low fat signal (0–10%). All segmentation steps were performed by the same operator. The workflow of the image segmentation is illustrated in Fig. 2.

## Statistical analysis

All statistical analyses were performed using the open-source software R (R Foundation for Statistical Computing, version 1.3.1056, 2020). The Shapiro-Wilk normality test revealed a non-Gaussian distribution of the nominal variables including age, and the PDFF and  $T_2^*$  of the entire breast and the fibroglandular tissue. Calculations of the left and right breast were compared using the intraclass correlation coefficient (ICC) followed by paired *t*-tests. To reduce bias, one breast in each woman was chosen according to the exclusion criteria for further analysis (Fig. 1). The inter-reader reliability of the mammographic and MRI-based breast density estimations was assessed using Cohen’s kappa. The Cohen’s kappa reference values were used established by McHugh et al [22]. To test the concordance between the categorical mammographic and nominal MRI-based breast density measurements, the Spearman correlation coefficient *r* was calculated. To

examine the relationship between the categorical, visually estimated mammographic and MRI-based breast density, the chi-squared test of independence was performed. The PDFF values were plotted in relation to the four ACR breast density categories. To evaluate their congruity, the Kruskal-Wallis test was performed followed by the Bonferroni-Holm correction criteria to adjust the *p* values in multiple comparisons due to their non-Gaussian distribution. To test for differences of average measures, paired *t*-tests were used. A two-tailed *p* value < 0.05 was considered statistically significant.

## Results

Evaluable PDFF maps were obtained in 138 women, excluding 55 patients due to different sources of artifacts. In detail, 21 datasets were excluded due to the presence of silicone implants (current Dixon processing does not account for silicone signals). In addition, 34 datasets were excluded due to a failing phase error correction in the fat quantification (Fig. 1). The failing phase error correction step resulted in erroneous fat fraction values over the entire field of view and was mostly likely related to an erroneous phase error estimation due to the lack of signal in the lungs within a large portion of the imaging field of view.

Patient and breast characteristics are displayed in Table 1. In this patient cohort, 91 women (66 %) presented with a reported history of breast cancer treated with surgical breast conserving therapy (*n* = 58), and/or a combination of radiation, chemotherapy, and additional antihormonal treatment (*n* = 46). All cancer patients revealed normal MRI follow-ups without any signs of recurrence. Separate calculations for right and left breasts showed similar results with a range of ICC of 0.997 to 0.999 (*p* < .001) for breast density and glandular tissue, and paired *t*-tests demonstrated no significant differences (*p* = .94). The chi-squared test revealed a significant relationship between visual mammographic and conventional MRI-based breast density readings (*p* < .001). The inter-reader reliability in mammographic breast density estimations was found to be low (*k* = 0.5, 95% confidence interval [CI] 0.38, 0.62) [22]. The inter-rater reliability in conventional MRI-based breast density estimations was moderate (*k* = 0.7, 95% CI 0.6, 0.79). Interestingly, the inter-rater reliability between mammographic and conventional MRI-based breast density readings revealed a minimal agreement (*k* = 0.3, 95% CI 0.17, 0.42). There was no difference in qualitative breast density between the craniocaudal view and left mediolateral-oblique view of the breast (*p* = .25).

Across all patients, the mean PDFF of the entire breast was 76.1 % and the mean PDFF of only the fibroglandular tissue was 7.7 %. Figure 3 shows an exemplary comparison of mammograms and PDFF maps of all four ACR categories. The PDFF of the entire breast strongly, negatively correlated with the mammographic breast density measurements (Spearman

**Table 1** Patient characteristics

Parameter	Overall patient cohort <i>N</i> (%)
No. of patients	138
Age (median [IQR])	49.5 (43–56)
Breast characteristics	
ACR	
A	13 (9.4)
B	52 (37.7)
C	53 (38.4)
D	20 (14.5)
PDFF (median [IQR])	81.83 (73.31–87.97)
Breast cancer in history	
Yes	
Surgery	58 (42.0)
Radiation therapy	40 (29)
Chemotherapy	28 (20.3)
Hormone therapy	24 (17.4)
No	
47 (34.1)	
Family history for breast cancer	
Positive	
26 (18.8)	
Negative	
77 (55.8)	
Gene mutation (including BRCA, FANCM, CHEK2)	
Yes	
29 (21.0)	
No	
49 (35.5)	

Patient and tumor characteristics. *IQR* interquartile range, *ACR* American College of Radiology categories, [A] almost entirely fatty, [B] scattered fibroglandular densities, [C] heterogeneously dense, and [D] extremely dense, *PDFF* proton density fat fraction, *BRCA* breast cancer gene, *FANCM* Fanconi anemia, complementation group M, *CHEK2* Checkpoint Kinase 2

rho:  $-0.69$ ,  $p < .001$ ) and revealed a significant distinction between all four ACR categories following the Bonferroni-Holm correction (Fig. 4, Table 2). The PDFF of the entire breast also correlated with MRI-based visual estimation of the breast density (Spearman rho:  $-0.56$ ,  $p < .001$ ). Furthermore, the PDFF of fibroglandular tissue showed a correlation with age (logistic regression coefficient  $-0.02$ ,  $p = .007$ , Pearson rho:  $0.56$ ,  $p = .03$ , Fig. 5). Mean  $T_2^*$  over the fibroglandular tissue also positively correlated with increasing ACR categories (Spearman rho:  $0.34$ ,  $p < .001$ ). Mean  $T_2^*$  of the fibroglandular tissue did not correlate with age ( $p = .72$ , Fig. 5) but showed a trend towards shorter  $T_2^*$  values with increasing fat concentration stratified by ACR categories ( $p < .001$ ).

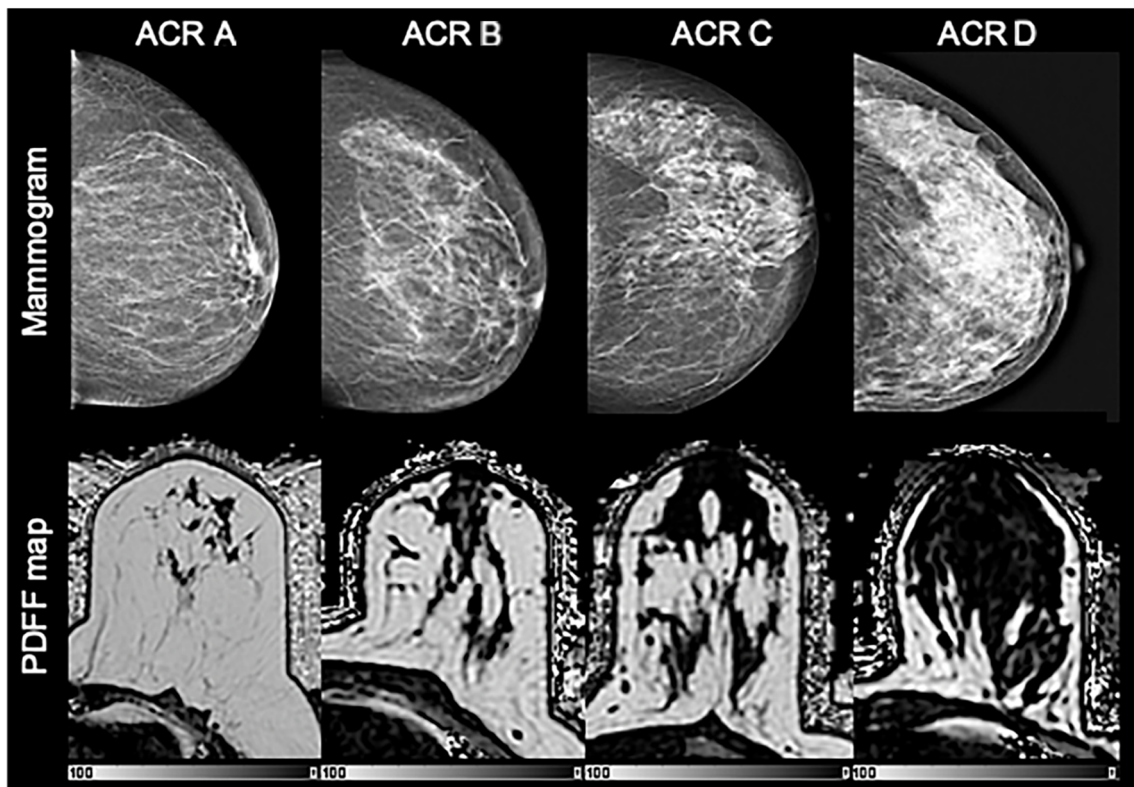
## Discussion

The main finding of this study is that the breast PDFF strongly negatively correlates with mammographic and conventional

MRI-based visual estimation of the breast density. The PDFF is an objective quantification of the breast density and in combination with  $T_2^*$  also provides information about the structural composition of the breast. The acquisition of the PDFF maps only requires a few minutes of MRI scan time which facilitates its applicability in clinical breast MR. Altogether, the PDFF allows for a robust, non-ionizing, quantitative alternative to measure the breast density and track structural alterations in the composition of the breast. Therefore, the PDFF can be used as a parameter for an individualized risk assessment for breast cancer.

Breast density estimation has emerged as a priority for assessing breast cancer risk as it is identified as one of the most prevalent risk factors for breast cancer and is implemented in most of the multifactorial risk assessment software tools [3, 23, 24]. Although radiographic mammography is the most widely used breast density measurement in clinical practice, its informative value is limited by a low intra- and inter-reader reliability. This discordance is influenced by the experience of the reporting radiologist and an incongruent technical execution [6, 25, 26]. Furthermore, the resulting two-dimensional projection of the volumetric body has a limited capacity to provide objective proportions of the breast composition, especially in dense breasts [4].

The clinical gold standard to estimate breast density remains mammography mainly due to its wide availability and relatively fast conductance. With the introduction of the BI-RADS classification system, the visual estimation of the breast density in  $T_1$ -weighted MRI images was internationally standardized but is also still subject to moderate inter-rater reliability issues. The proposed, water-fat chemical-shift-based PDFF overcomes the 2D projection limitations of conventional mammography and presents a reliable, objective discriminator of tissue fat and water content. Previous works assessing breast density with MRI have relied on thresholding of  $T_1$ -weighted and Dixon images [8, 10, 11, 27]. Tagliafico et al proposed a semi-automated method with an intensity threshold for dense breast parenchyma which is related to the entire breast volume [8]. Furthermore, Thomson et al used a defined threshold as a proposed MRI-based breast density measure calculated as the ratio of breast voxels with  $< 80\%$  apparent fat fraction [27]. These approaches are susceptible to partial volume effects because of the sharp thresholding assigned to each breast voxel which ultimately increases breast density estimation errors. Besides, these methods did not account for the  $T_1$  bias which potentially results in a relative amplification of the signal of fat compared to the signal of water. The present work uses a low flip angle gradient echo sequence for mapping the PDFF, which is independent of the underlying relaxation properties and therefore reduces  $T_1$  bias. In another approach, Ding et al employed a Dixon acquisition using an IDEAL-GRASE sequence to quantify

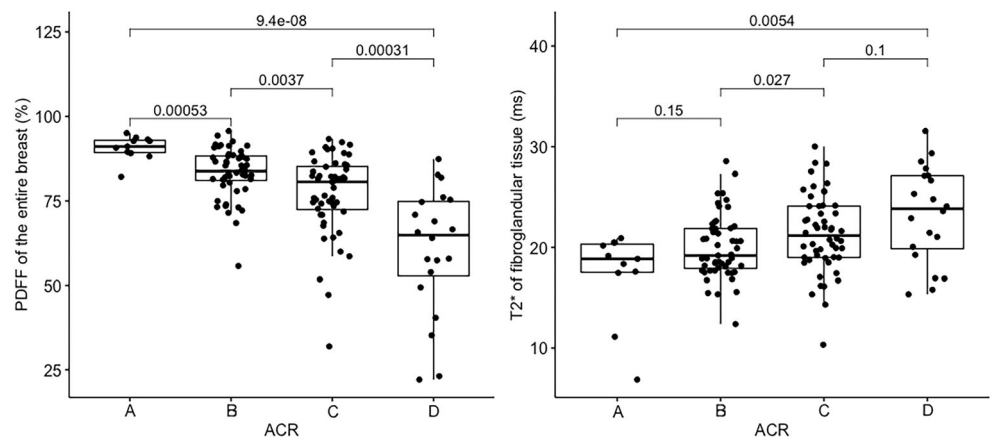


**Fig. 3** Mammograms with breast density estimation following the American College of Radiology Breast Imaging Reporting and Data System categories A-D with corresponding PDFF maps

breast density based on the fraction of fibroglandular tissue and actual water content in each voxel. However, this sequence is limited by an intrinsic fat-water signal bias and therefore requires additional calibrations to remove  $T_1$ - and  $T_2$ -weighting effects [11]. In this study, a 6-echo gradient echo acquisition was used accounting for a single  $T_2^*$  decay and a pre-calibrated 7-peak fat spectrum accounting for the presence of multiple peaks in the fat spectrum in order to reduce potential confounding imaging factors, as also recently proposed [18]. In addition, scanning time remained under 3 min which renders the PDFF imaging sequence easily incorporable into clinical routine breast MRI exams.

Ascertained PDFF is independent of field strength, scanner platform, and specific scanning parameters and highly correlates with parenchymal triglyceride concentration [12]. Consequently, the PDFF has been already successfully applied in other organs and reliably quantified even small changes in tissue fat concentration [15, 28–30]. Unlike the signal fat-fraction, the PDFF reflects the actual content of fat in the breast and therefore presents a potentially reliable, standardized biomarker of breast density, as recently proposed [18]. In this study, the PDFF especially of the fibroglandular tissue also correlated with age which potentially mirrors the structural change of the breast with age [31]. Most prominent

**Fig. 4** (A) Correlation of entire breast PDFF values to the four American College of Radiology Breast Imaging Reporting and Data System categories with adjusted  $p$  values revealing a significant negative correlation. (B) Significant correlation of  $T_2^*$  of fibroglandular tissue of the breast to ACR A-D



**Table 2** MRI parameter results stratified by the four American College of Radiology Breast Imaging Reporting and Data System categories (ACR A-D)

ACR	Mammography <i>N</i> (%)	Breast MR <i>N</i> (%)	PDFF entire breast Median % (IQR)	PDFF fibroglandular Median % (IQR)	T2* fibroglandular Median % (IQR)
ACR A	13 (9.4)	10 (7.2)	91.4 (89.8–93.2)	8.6 (7.6–9.5)	17.6 (15.2–20.7)
ACR B	52 (37.7)	52 (37.7)	85.1 (81.3–88.1)	8.5 (7.0–10.2)	19.9 (17.6–23.4)
ACR C	53 (38.4)	52 (37.7)	79.7 (73.8–84.8)	7.6 (6.1–9.3)	21.8 (19.8–24.4)
ACR D	20 (14.5)	24 (17.4)	64.0 (57.9–73.2)	5.8 (4.4–6.8)	23.1 (20.3–27.1)

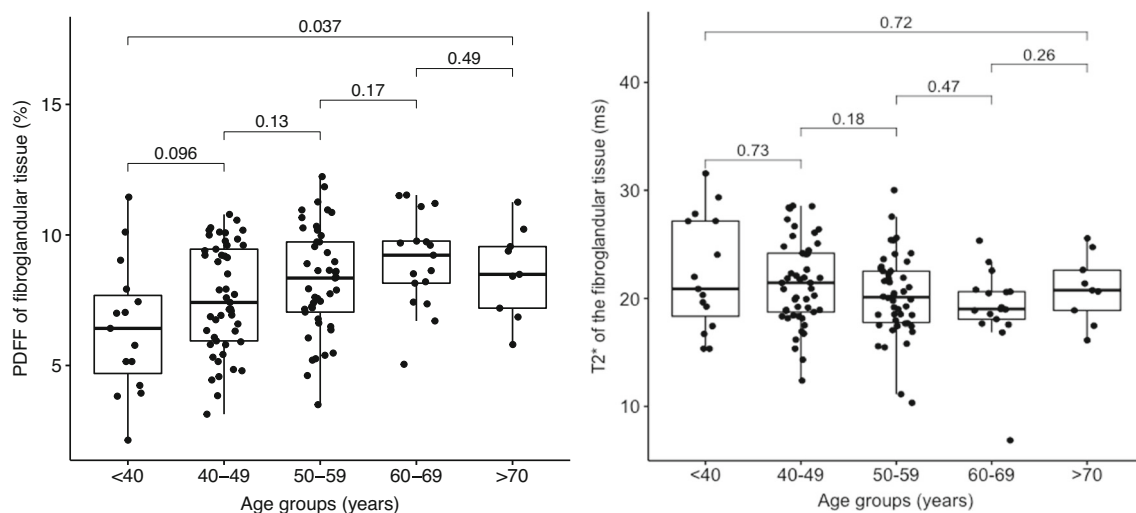
MRI parameter results. ACR American College of Radiology according to the Breast Imaging Reporting and Data System (BI-RADS, 5th edition), [A] almost entirely fatty, [B] scattered fibroglandular densities, [C] heterogeneously dense, which may obscure small masses, and [D] extremely dense, PDFF proton density fat fraction, IQR interquartile range

across the menopausal transition of 50–60 years, the fibroglandular tissue shows a natural decline in the amount of dense breast tissue with aging [32] that is also detectable with PDFF and T<sub>2</sub>\* measurements. The observed dependence of fibroglandular tissue T<sub>2</sub>\* with the ACR categorization might be also related to magnetic susceptibility differences between the water and fat components or to changes in the glandular component within the fibroglandular tissue [33]. The introduction of the PDFF (in combination with T<sub>2</sub>\*) might provide additional and valuable information in the course of an individual risk assessment in screening patients and follow-up MRIs. The denser and more heterogeneously composed the breast presents, the higher is the risk to develop breast cancer but the lower is the detectability rate on mammography. As demonstrated in the DENSE trial, the additional use of MRI in clinical screening exams in very dense breasts with normal results in mammography significantly decreased the occurrence of interval cancers [34]. With the supplemental integration of the PDFF, the breast density can be objectively classified for a robust and standardized, individual risk assessment especially in patients with very dense and heterogeneously composed breasts to enable tailored screening regimes. Furthermore, structural changes over time may be

objectively tracked for an individualized risk assessment or treatment response estimation after cancer occurrence or anti-hormonal treatment [35, 36].

The present work has some limitations. First, it is focused on the relationship between breast PDFF and the mammographic and conventional MRI-based density metric. No validation of the reported fat fraction to another MR-based fat concentration measure was performed. However, the employed PDFF methodology has already accounted for confounding effects known from the application of the methodology in other organs. Second, no breast PDFF reproducibility analysis was presently performed. High reproducibility of PDFF for liver fat content and breast fat content in volunteers has been already confirmed [13, 37], therefore PDFF for breast parenchyma fat concentration in patients would be also expected to be high. Third, the employed methodology was based on a bipolar readout acquisition which accounted for phase errors in the quantification but showed failing behaviors in a small number of datasets. Further work using monopolar readout gradient could alleviate such problems [38].

The proposed breast PDFF mapping is a promising approach to an accurate, user-independent, and non-ionizing tissue fat concentration measurement that is directly comparable



**Fig. 5** Correlation of mean PDFF values of the fibroglandular tissue to different age groups with adjusted *p* values revealing a positive correlation

with visual mammographic, and conventional MRI-based breast density estimations. The improved concordance of the poor reader reproducibility, its platform and scan parameter independence, and the simple integration of the PDFF in clinical routine breast MRI exams may provide clinicians with a valuable tool for an individualized cancer risk assessment accurate and a reliable evaluation of longitudinal, structural changes in breast density and composition.

**Acknowledgements** The present study was supported in part by the European Research Council (grant agreement No 677661, ProFatMRI). The authors also acknowledge research support from Philips Healthcare.

**Funding** Open Access funding enabled and organized by Projekt DEAL. The present was supported in part by the European Research Council (grant agreement No 677661, ProFatMRI).

## Declarations

**Guarantor** The scientific guarantor of this publication is Prof. Dr. Dimitrios Karampinos from the Technical University Munich.

**Conflict of interest** T.B., M.W., S.R., C.B., J.S., S.M., M.R.M., and E.M.F. have no conflicts of interest to declare. K.W. is currently employed at Philips GmbH. D.C.K. receives a research grant from the European Research Council (grant agreement No 677661, ProFatMRI).

**Statistics and biometry** One of the authors (M.W.) has significant statistical expertise and gave statistical advice. However, no complex statistical methods were necessary for this paper.

**Informed consent** Written informed consent was waived by the institutional review board.

**Ethical approval** Institutional review board approval was obtained. The retrospective analysis of the routinely acquired data was approved by the local ethics committee (number 61/21 S-KK).

## Methodology

- Retrospective
- Diagnostic or prognostic study
- Performed at one institution

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain

permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

## References

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A (2018) Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin.* 68(6):394–424
2. Vachon CM, van Gils CH, Sellers TA et al (2007) Mammographic density, breast cancer risk and risk prediction. *Breast Cancer Res.* 9(6):217
3. Boyd NF, Guo H, Martin LJ et al (2007) Mammographic density and the risk and detection of breast cancer. *N Engl J Med.* 356(3):227–236
4. Lee CI, Chen LE, Elmore JG (2017) Risk-based breast cancer screening: implications of breast density. *Med Clin North Am.* 101(4):725–741
5. D'Orsi CJ, Sickles EA, Mendelson EB, Morris EA (2013) ACR BI-RADS® Atlas, Breast Imaging Reporting and Data System: 5th edition. American College of Radiology, Reston, VA, USA
6. Sohn G, Lee JW, Park SW et al (2014) Reliability of the percent density in digital mammography with a semi-automated thresholding method. *J Breast Cancer.* 17(2):174–179
7. Khazen M, Warren RM, Boggis CR et al (2008) A pilot study of compositional analysis of the breast and estimation of breast mammographic density using three-dimensional T1-weighted magnetic resonance imaging. *Cancer Epidemiol Biomarkers Prev.* 17(9):2268–2274
8. Tagliafico A, Bignotti B, Tagliafico G et al (2014) Breast density assessment using a 3T MRI system: comparison among different sequences. *PLoS One.* 9(6):e99027
9. Clendenen TV, Zeleniuch-Jacquotte A, Moy L, Pike MC, Rusinek H, Kim S (2013) Comparison of 3-point Dixon imaging and fuzzy C-means clustering methods for breast density measurement. *J Magn Reson Imaging.* 38(2):474–481
10. Wengert GJ, Helbich TH, Vogl WD et al (2015) Introduction of an automated user-independent quantitative volumetric magnetic resonance imaging breast density measurement system using the Dixon sequence: comparison with mammographic breast density assessment. *Invest Radiol.* 50(2):73–80
11. Ding J, Stopeck AT, Gao Y et al (2018) Reproducible automated breast density measure with no ionizing radiation using fat-water decomposition MRI. *J Magn Reson Imaging.* 48(4):971–981
12. Reeder SB, Hu HH, Sirlin CB (2012) Proton density fat-fraction: a standardized MR-based biomarker of tissue fat concentration. *J Magn Reson Imaging.* 36(5):1011–1014
13. Yokoo T, Shieh-morteza M, Hamilton G et al (2011) Estimation of hepatic proton-density fat fraction by using MR imaging at 3.0 T. *Radiology.* 258(3):749–759
14. Meisamy S, Hines CD, Hamilton G et al (2011) Quantification of hepatic steatosis with T1-independent, T2-corrected MR imaging with spectral modeling of fat: blinded comparison with MR spectroscopy. *Radiology.* 258(3):767–775
15. Fukui H, Hori M, Fukuda Y et al (2019) Evaluation of fatty pancreas by proton density fat fraction using 3-T magnetic resonance imaging and its association with pancreatic cancer. *Eur J Radiol.* 118:25–31
16. Baum T, Inhuber S, Dieckmeyer M et al (2016) Association of quadriceps muscle fat with isometric strength measurements in healthy males using chemical shift encoding-based water-fat magnetic resonance imaging. *J Comput Assist Tomogr.* 40(3):447–451



17. Yokoo T, Bydder M, Hamilton G et al (2009) Nonalcoholic fatty liver disease: diagnostic and fat-grading accuracy of low-flip-angle multiecho gradient-recalled-echo MR imaging at 1.5 T. *Radiology*. 251(1):67–76
18. Henze Bancroft LC, Strigel RM, Macdonald EB et al (2022) Proton density water fraction as a reproducible MR-based measurement of breast density. *Magn Reson Med*. 87(4):1742–1757
19. Borde T, Wu M, Ruschke S, Boehm C, Weiss K, Metz S, Makowski RM, Karampinos CD Assessing breast density using the standardized proton density fat fraction based on chemical shift encoding-based water-fat separation. *ISMRM*; 17.-20.05.2021; Vancouver, CA.2021
20. Liberman L, Menell JH (2002) Breast Imaging Reporting and Data System (BI-RADS). *Radiol Clin North Am*. 40(3):409–430 v
21. Yushkevich PA, Piven J, Hazlett HC et al (2006) User-guided 3D active contour segmentation of anatomical structures: significantly improved efficiency and reliability. *Neuroimage*. 31(3):1116–1128
22. McHugh ML (2012) Interrater reliability: the kappa statistic. *Biochem Med (Zagreb)*. 22(3):276–282
23. Engmann NJ, Golmakani MK, Miglioretti DL, Sprague BL, Kerlikowske K, Breast Cancer Surveillance C (2017) Population-attributable risk proportion of clinical risk factors for breast cancer. *JAMA Oncol*. 3(9):1228–1236
24. Raghavendra A, Sinha AK, Le-Petross HT et al (2017) Mammographic breast density is associated with the development of contralateral breast cancer. *Cancer*. 123(11):1935–1940
25. Gur D, Sumkin JH, Hardesty LA et al (2004) Recall and detection rates in screening mammography. *Cancer*. 100(8):1590–1594
26. Alomaim W, O’Leary D, Ryan J, Rainford L, Evanoff M, Foley S (2019) Variability of breast density classification between US and UK radiologists. *J Med Imaging Radiat Sci*. 50(1):53–61
27. Thomson CA, Chow HHS, Wertheim BC et al (2017) A randomized, placebo-controlled trial of diindolylmethane for breast cancer biomarker modulation in patients taking tamoxifen. *Breast Cancer Res Treat*. 165(1):97–107
28. Lohofer FK, Kaissis GA, Muller-Leisse C et al (2019) Acceleration of chemical shift encoding-based water fat MRI for liver proton density fat fraction and T2\* mapping using compressed sensing. *PLoS One*. 14(11):e0224988
29. Ruschke S, Pokorney A, Baum T et al (2017) Measurement of vertebral bone marrow proton density fat fraction in children using quantitative water-fat MRI. *MAGMA*. 30(5):449–460
30. Karampinos DC, Holwein C, Buchmann S et al (2017) Proton density fat-fraction of rotator cuff muscles is associated with isometric strength 10 years after rotator cuff repair: a quantitative magnetic resonance imaging study of the shoulder. *Am J Sports Med*. 45(9):1990–1999
31. Checka CM, Chun JE, Schnabel FR, Lee J, Toth H (2012) The relationship of mammographic density and age: implications for breast cancer screening. *AJR Am J Roentgenol*. 198(3):W292–W295
32. Engmann NJ, Scott C, Jensen MR et al (2019) Longitudinal changes in volumetric breast density in healthy women across the menopausal transition. *Cancer Epidemiol Biomarkers Prev*. 28(8):1324–1330
33. Lee SH, Ryu HS, Jang MJ et al (2021) Glandular tissue component and breast cancer risk in mammographically dense breasts at screening breast US. *Radiology* 210367
34. Bakker MF, de Lange SV, Pijnappel RM et al (2019) Supplemental MRI screening for women with extremely dense breast tissue. *N Engl J Med*. 381(22):2091–2102
35. Sandberg ME, Li J, Hall P et al (2013) Change of mammographic density predicts the risk of contralateral breast cancer—a case-control study. *Breast Cancer Res*. 15(4):R57
36. Brentnall AR, Warren R, Harkness EF et al (2020) Mammographic density change in a cohort of premenopausal women receiving tamoxifen for breast cancer prevention over 5 years. *Breast Cancer Res*. 22(1):101
37. Middleton MS, Heba ER, Hooker CA et al (2017) Agreement between magnetic resonance imaging proton density fat fraction measurements and pathologist-assigned steatosis grades of liver biopsies from adults with nonalcoholic steatohepatitis. *Gastroenterology*. 153(3):753–761
38. Ruschke S, Eggers H, Kooijman H et al (2017) Correction of phase errors in quantitative water-fat imaging using a monopolar time-interleaved multi-echo gradient echo sequence. *Magn Reson Med*. 78(3):984–996

**Publisher’s note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.