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Evaluation of a Multi-Molecule Molecular Communication Testbed Based on Spectral Sensing

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Abstract—This work presents a novel flow-based molecular communication (MC) testbed using spectral sensing and ink concentration estimation to enable real-time multi-molecule (MUMO) transmission. MUMO communication opens up crucial opportunities for increased throughput as well as implementing more complex coding, modulation, and resource allocation strategies for MC testbeds. A concentration estimator using noninvasive spectral sensing at the receiver is proposed based on a simple absorption model. We conduct in-depth channel impulse response (CIR) measurements and a preliminary communication performance evaluation. Additionally, a simple analytical model is used to check the consistency of the CIRs. The results indicate that by utilizing MUMO transmission, on-off-keying, and a simple difference detector, the testbed can achieve up to 3 bits per second for near-error-free communication, which is on par with comparable testbeds that utilize more sophisticated coding or detection methods. Our platform lays the ground for implementing MUMO communication and evaluating various physical layer and networking techniques based on multiple molecule types in future MC testbeds in real time.

Index Terms—molecular communication, testbeds, multimolecule transmission, spectral sensing

I. Introduction

Molecular communication (MC) is based on the transfer of molecules for information exchange. It is envisioned to enable various revolutionary use cases in nanotechnology and medicine by providing nano-scale, energy-efficient, and biocompatible communication, for example, inside the human body. While research into theoretical models and simulation frameworks has advanced significantly since the emergence of MC around two decades ago, there are still only relatively few experimental implementations to validate MC systems.

However, some concepts for MC testbeds have been proposed in the past [1]. Among the first approaches for a practical implementation of MC were air-based unconstrained setups, where a spray-based transmitter (TX) would propel information molecules, often an alcohol solution, towards a receiving sensor [2], [3]. These testbeds are inexpensive, quick to set up, and have successfully been used for more sophisticated communication techniques such as MIMO [4] and network coding [5]. The second large category consists

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of flow-based experimental setups, often used as abstract representations of future in-body communications, for example, in the bloodstream. It is common to constrain the system to circular tubes with diameters of millimeter scale, induce flow, and transmit information using various types of pumps and information molecules, respectively. Farsad et al. presented an experimental MC platform based on transmitting acid and base as information molecules using multiple peristaltic pumps and inserting a pH probe into the channel as a receiver (RX) [6]. The testbed achieves data rates of up to 4 bits-persecond (bps) with a recurrent neural network detector. In [7], Brand et al. implemented a closed-loop flow-based testbed using a switchable fluorescent protein that can enter different states to convey information to the RX. The proposed *media* modulation concept increases efficiency since the same set of information molecules can be used for many consecutive transmissions. Two LED arrays for activating and deactivating the protein are used as TX and eraser, while a fluorescence measurement with a spectrometer occurs at the RX. The paper demonstrates a bit rate of 0.1 bps over a distance of 6 cm using a difference detector. The experimental MC system built by Wicke et al. uses special magnetic nanoparticles as information molecules, a peristaltic pump as a TX, and a non-invasive susceptometer as an RX [8]. In [9], several improvements of the testbed in [8] are described in detail, including micropumps at the TX and a convolutional neural network detector, yielding a bit rate of approximately 6 bps. In [8], different channel lengths between 5 cm and 40 cm are compared, while the performance bound in [9] is acquired for a channel length of 5 cm. In [10], real-time chemical reactions are utilized to implement an MC testbed capable of transmitting over multiple meters with a precision-syringepump TX. Experiments are conducted with symbol periods of around 30 min, but the platform demonstrates working MC communication with practical chemical reactions across relatively long distances.

All flow-based testbeds described above focus solely on single-link MC between one TX and one RX. While some platforms are potentially extendable to multiple participants, it is important to choose less invasive options for TX and RX because, for example, peristaltic pumps in the transmission channel, pH probes, or voltage measurements can add disturbances to the channel, which make adding more nodes

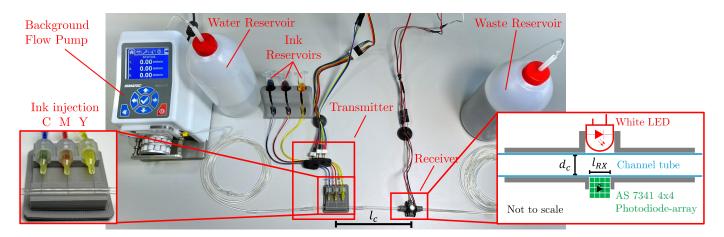


Fig. 1. Annotated image of the entire testbed structure. From a water reservoir, the background flow is generated by a peristaltic pump through tubes of diameter d_c . The liquid then flows past the TX which consists of three micropumps capable of injecting cyan, magenta, and yellow ink into the channel. After a channel of length l_c the mixture flows past the non-invasive RX with a spectral sensor of length l_{RX} , and lastly arrives in a waste reservoir.

difficult. The realization of more complex MC use cases relies on the communication between multiple low-capability nodes [11]. Therefore, the experimental investigation of systems with multiple TXs or RXs is crucial. Wang et al. have proposed a protocol and novel multiple access (MA) scheme for a multi-user MC system and conducted an evaluation on an experimental platform based on salinity with up to 4 TXs [12]. The presented system achieves a per TX throughput of up to 1 bps. The authors also emulate a parallel multi-molecule (MUMO) transmission but run it as two separate evaluations and do not implement it in real-time, suggesting a practical MUMO implementation as important future work.

The previously described MC testbeds are limited to single-molecule (SIMO) transmission. While in [10], multiple molecules are used for the chemical reactions, there is a single channel based on SIMO transmission. Pan et al. presented an experimental MC platform using an RGB color sensor RX and colored ink to transmit information from a single TX based on a peristaltic pump and electrical valves [13]. While the authors describe a MUMO operation of the testbed as theoretically possible, communication is demonstrated only with one color at a maximum rate of 0.1 bps.

In this paper, we present a novel, inexpensive, and easy-to-setup MC testbed capable of differentiating multiple molecules in real time. The latter enables MUMO communication, thereby increasing throughput and opening up numerous possibilities for implementing more complex coding, modulation, and resource management schemes. We utilize a low-cost micropump TX and non-invasive spectral RX design, which enables flexible adaptation of the physical network parameters and is easy to extend towards multiple TXs or RXs due to its repeatable 3D-printed modular structure. The information molecules in the form of printer ink are also inexpensive and safe to handle. We propose a linear ink concentration estimator capable of extracting three different simultaneously present colors (cyan, magenta, yellow) from a spectral measurement. Extensive channel impulse response (CIR) measurements of both SIMO and MUMO transmissions

are presented, and the impact of MUMO communication on the channel is discussed. Additionally, we apply a simple analytical model to confirm the consistency of our results. Lastly, we conduct a performance evaluation of the MUMO communication system using a simple difference detector and achieve up to 3 bps at a bit error rate (BER) of 10^{-2} over a distance of $> 20 \, \mathrm{cm}$, showing the practical communication potential of the testbed in the context of the related work discussed above.

II. TESTBED DESIGN

In the following, we describe the structure of our MC testbed by briefly focusing on the channel, TX and information molecules, and the RX. A photograph of the entire testbed structure is depicted in Figure 1.

The channel consists of a cylindrical tube made from a transparent bio-compatible polymer (Tygon) with a diameter of $d_{\rm c}=2\cdot r_{\rm c}=1.6\,{\rm mm}$. Background flow $Q_0=10\,{\rm \frac{mL}{min}}$ is provided by a peristaltic pump (Reglo ICC) with 12 rollers per channel for minimum pulsatility of the flow speed. It pumps water from the reservoir on the left-hand side through the channel. Then, TX and RX interact with the channel as described in the following sections, and the resulting wastewater is collected in a waste reservoir on the other side.

A. Information Molecules and Transmitter

In our testbed, we use colored ink as information molecules. To facilitate MUMO transmission using inexpensive materials, we have opted to use cyan (C), magenta (M), and yellow (Y) printer ink. These colors affect different parts of the visible light spectrum, and therefore, their impact is differentiable at the RX, as we will show later.

The TX consists of three micropumps, which use piezoelectric components to inject small amounts of liquid into the channel. Each pump is connected to an ink reservoir contained in a 3D-printed housing on one end. On the other end, the TX interfaces with the channel through a bluntend syringe needle for each color with an outer and inner diameter of 0.23 mm and 0.11 mm, respectively. Another 3Dprinted housing ensures consistent alignment of the needles with the channels such that they are inserted perpendicular to the background flow direction. The distance between each needle along the channel is 1.3 cm, and the TX midpoint is $l_{\rm c}=23.5\,{\rm cm}$ from the RX midpoint. The needle injecting cyan is furthest upstream, magenta in the middle, and yellow the furthest downstream. To minimize the influence of downstream needles on the ink injected by the upstream needles, it is crucial that their diameter is very small and that the insertion depth of the needles is decreased along the flow direction. For example, the cyan needle is inserted furthest so that the cyan ink passes by the other needles. One TX, consisting of three pumps, is connected to a single driver chip connected to an Arduino Nano. The pumps are controlled through an input voltage level that determines the flow speed at the outlet. The peristaltic pump causes significant pressure to be applied to the syringe needle outlets. Backflow in the form of water enters the TX if no voltage is applied to the micropumps while the background flow is running, a phenomenon observed in MC testbeds before [9]. Therefore, we define an idle voltage level $V_{\rm idle}$, which is always applied to the TX micropumps.

B. Receiver

The heart of the RX is an Adafruit AS7341 spectral sensor. It consists of a 4-by-4 sensor array that measures eight overlapping channels with center wavelengths 415 nm, 445 nm, 480 nm, 515 nm, 555 nm, 590 nm, 630 nm, and 680 nm. To maximize sensitivity and minimize outside interference, we have created a custom 3D-printed housing, as depicted and annotated on the right-hand side of Figure 1, into which the channel tube can be inserted. A white LED illuminates the channel from the side opposite the sensor. The sensor and LED are connected and controlled by an Arduino Nano, which samples the sensor readings at a frequency of approximately 19 Hz.

III. INK CONCENTRATION ESTIMATION

Many detection algorithms in MC rely on molecule concentrations as input signals. Since the spectral sensor does not directly provide the cyan, magenta, and yellow ink concentrations, $c_{\rm c}(t)$, $c_{\rm m}(t)$, and $c_{\rm y}(t)$, the testbed relies on estimating the individual color concentrations from the mixed spectrum measured at the RX. In our setup, deviations in the received spectrum are caused by light absorption of the inks passing through the RX. The Beer-Lambert law describes the linear relationship between concentration $c_i(t)$ ($i \in \{c, m, y\}$) and absorbance $a_i^j(t)$ due to ink i in the j-th spectral channel of the RX as

$$a_i^j(t) = \log_{10}\left(\frac{I_0^j}{I^j(t)}\right) = b_i^j c_i(t),$$
 (1)

where b_i^j is the absorption coefficient of ink i in channel j, $I^j(t)$ is the time-varying light intensity measured in the j-th channel, and I_0^j is a reference measurement with no ink in the

channel [14]¹. In the presence of multiple inks, the measured absorbance in channel j, $a^{j}(t)$, is the sum of the individual absorbances $a_{i}^{j}(t)$ [15] and therefore, in our case:

$$a^{j}(t) = a_{c}^{j}(t) + a_{m}^{j}(t) + a_{v}^{j}(t).$$
 (2)

Since we measure 8 distinct channels at the RX, as described in Section II, we can define the absorbance vector

$$\mathbf{a}(t) = [a^1(t), a^2(t), \cdots, a^8(t)]^T,$$
 (3)

the absorption coefficient vector \mathbf{b}_i for ink i, and an absorption matrix \mathbf{B}

$$\mathbf{b}_i = \begin{bmatrix} b_i^1, b_i^2, \cdots, b_i^8 \end{bmatrix}^T$$
 and $\mathbf{B} = [\mathbf{b}_c, \mathbf{b}_m, \mathbf{b}_y]$. (4)

Using Eqs. (1) and (2), the previous definitions, and the concentration vector $\mathbf{c}(t) = \left[c_{\mathrm{c}}(t), c_{\mathrm{m}}(t), c_{\mathrm{y}}(t)\right]^{T}$, we arrive at the following system of linear equations:

$$\mathbf{a}(t) = \mathbf{B} \cdot \mathbf{c}(t). \tag{5}$$

Due to the linearity of the system, by sending $N_{\rm p}$ calibration pulses with known peak concentration $c_{i,{\rm peak}}=1$ from each ink individually and averaging the peak absorption, we can determine the \mathbf{b}_i as

$$\mathbf{b}_{i} = \frac{1}{N_{p}} \sum_{k=1}^{N_{p}} \max_{t \in T_{k}} \left(\mathbf{a}(t) \right), \tag{6}$$

where T_k is a time interval, in which the k-th pulse is received. To calculate the estimated ink concentration vector $\hat{\mathbf{c}}(t)$ from an absorbance measurement $\mathbf{a}(t)$, we employ ordinary least squares estimation. According to [16, Chapter 4], for the overdetermined system in Eq. (5), the least squares optimal estimator is equal to the Moore-Penrose pseudoinverse \mathbf{B}^{\dagger} and therefore

$$\hat{\mathbf{c}}(t) = \mathbf{B}^{\dagger} \mathbf{a}(t) = \left(\mathbf{B}^T \mathbf{B}\right)^{-1} \mathbf{B}^T \mathbf{a}(t). \tag{7}$$

IV. CHANNEL IMPULSE RESPONSE MODELING

The propagation of the ink information molecules in our testbed is primarily based on advection via bulk background flow of water. Therefore, we will briefly analyze the expected flow properties in our testbed given its parameters and introduce a simple analytical model.

Firstly, we try to categorize the fluid flow in the system as either laminar or turbulent, as this significantly impacts the subsequent modeling [17]. The Reynolds number Re is a dimensionless quantity used to differentiate the laminar and turbulent case and is defined as [18, Chapter 1]

$$Re = \frac{d_c v_{\text{avg}}}{v}, \tag{8}$$

with the channel diameter $d_{\rm c}$ and the average background flow velocity $v_{\rm avg}=\frac{Q_0}{\pi r_{\rm c}^2}$ as the characteristic length and velocity scales of the system, respectively, and the kinematic viscosity of the fluid ν . The transition from laminar to

¹The reference measurement is obtained by averaging the light intensity in the absence of ink in each channel over several seconds.

turbulent flow in a smooth circular pipe occurs at approximately Re = 2100 [18, Chapter 4]. Setting $d_{\rm c}=1.6\,{\rm mm}$, $Q_0=10\,{\rm mL\over min}$, and using the kinematic viscosity of water $\nu_{\rm water}=1.01\times 10^{-6}\,{\rm m^2\over s}$, we get Re = $131\ll 2100$ for our testbed, which we therefore assume to operate in the laminar regime. For laminar flow in a straight, circular pipe, we can assume a *Poiseuille* flow velocity profile [18, Chapter 4]. The flow velocity v is a function of the radius ρ , radially symmetric around the center of the channel ($\rho=0$), and defined as follows:

$$v(\rho) = v_{\text{max}} \left(1 - \frac{\rho^2}{r_{\text{c}}^2} \right), \ \rho \in [0, r_{\text{c}}],$$
 (9)

where $v_{\rm max}=2\cdot v_{\rm avg}$ is the maximum flow velocity in the middle and $r_{\rm c}=\frac{d_{\rm c}}{2}=0.8\,{\rm mm}$ is the channel radius.

Secondly, we determine the relative influence of diffusion on the fluid transport, which can be described by the dimensionless Péclet number $\mathrm{Pe} = \frac{r_c v_{\mathrm{avg}}}{D}$ [17], with the diffusion coefficient D. We use dye-based fully dissolved ink and therefore use the diffusion coefficient of water in water, $D \lesssim 2.299 \times 10^{-9} \, \frac{\mathrm{m}^2}{\mathrm{s}}$ [19], as an upper limit approximation. We can, therefore, calculate $\mathrm{Pe} \gtrsim 2.88 \times 10^4 \gg 1$ and conclude that flow dominates over diffusion, which can be neglected as a result.

For the purposes of a simplified analytical model, we assume the RX to be passively observing the molecules over the channel cross-section along a section with length $l_{\rm RX}$ centered at a distance $l_{\rm c}$ from the TXs. Additionally, we assume that the TXs emit an instantaneous pulse of molecules distributed over the cross-section at the injection point. Using the model proposed by Wicke et al. in [8], we define a radially symmetric initial distribution based on the Beta distribution

$$f(s) = \frac{\Gamma(\alpha + \beta)}{\Gamma(\alpha)\Gamma(\beta)} s^{\alpha - 1} (1 - s)^{\beta - 1}; \ s = \frac{\rho^2}{r_c^2} \in [0, 1], \quad (10)$$

with the Gamma function $\Gamma(\cdot)$ and two variable parameters α and β . We then assume that $l_{\rm c}\gg l_{\rm RX}{}^2$, and therefore obtain the CIR [8]

$$h(t) = \begin{cases} C \frac{\Gamma(\alpha+\beta)}{\Gamma(\alpha)\Gamma(\beta)} \left(1 - \frac{l_c}{v_{\max}t}\right)^{\alpha-1} \left(\frac{l_c}{v_{\max}t}\right)^{\beta} & t \ge \frac{l_c}{v_{\max}} \\ 0 & \text{else}, \end{cases}$$
(11)

with a dimensionless scaling factor C. Mirroring [8], Eqs. (10) and (11) will be used to fit α and β towards the measured CIR and thereby check the consistency of the CIR and initial distribution across trials and between different colors.

V. RESULTS

In the following section, we will present evaluation results based on measurements from our testbed as described in Section II and the methods and models introduced in Sections III and IV. This includes a characterization of the CIR and an analysis of the communication capabilities using a random

TABLE I EVALUATION PARAMETERS

Parameter	Symbol	Value(s)
Idle pump voltage	V_{idle}	40 V
Bit-1 pump voltage	V_1	$110\mathrm{V}$
Bit-0 pump voltage	V_0	$40\mathrm{V}$
Symbol period	$T_{ m sym}$	$\{0.5, 1, 2, 10\}$ s
Injection time	$T_{ m inj}$	$\{0.05, 0.1\}$ s
Background flow	Q_0	$10 \frac{\text{mL}}{\text{min}}$
Channel diameter	$d_{ m c}$	$1.6\mathrm{mm}$
Channel length	$l_{ m c}$	$23.5\mathrm{cm}$

bit sequence. The parameters used throughout the evaluation can be seen in Table I.

A. Channel Impulse Response Evaluation

We will use experimental data from the testbed to generate average CIR curves for SIMO and MUMO transmission, respectively. Therefore, we have gathered measurements of 210 pulses in total, 30 trials each of the seven possible molecule combinations from 3 colors, i.e. cyan (C), magenta (M), and yellow (Y), individually, and CM, CY, MY, and CMY. The pulses were generated with an injection time of $T_{\rm inj}=0.1\,{\rm s}$ and separated by $T_{\rm sym}=10\,{\rm s}$ to avoid intersymbol interference (ISI). The peak spectral deviations from the SIMO trials were then used to calculate the averaged absorption vectors ${\bf b}_c$, ${\bf b}_m$, and ${\bf b}_y$, as shown in Eq. (6) with $N_{\rm p}=30$. Subsequently, the absorption matrix ${\bf B}$ is calculated for the concentration estimator as defined in Eq. (4). The estimator converts the spectral measurements from each pulse into estimated molecule concentrations.

Figure 2 depicts the results of the averaged CIR analysis. The solid C, M, and Y line depicts the averaged CIR from the respective SIMO pulses, and the darker shaded area around the curve represents the maximum deviation from the average for the SIMO pulses. We can observe that they exhibit very similar characteristics in terms of their peak and fall-off time between all three colors, except for minor shape variations. Additionally, the deviations are quite close to the average. All considered CIRs return to zero after approximately 5 seconds.

The thin dash-dotted red line in each plot corresponds to the analytical model introduced in Section IV fitted towards the shape of the SIMO CIR of each color using exhaustive search and finding the minimum mean-squared error solution for α and β . We can observe a good fit of the CIR shape for the optimal solution in each case, capturing the characteristics of the rise and peak of the CIR well while mostly deviating in the tail section. This could be explained by the inaccurate assumption of instantaneous release of the molecules, as small amounts of ink continue to leak out of the injection needle after $T_{\rm inj}$ has passed. This would cause an underestimation of the analytical model in the latter part of the CIR due to molecules arriving later than expected. The resulting initial distributions and the values for the optimal α and β are shown in Figure 3. Here, we can see that the fitted initial distributions are similar for all three colors, with M and Y exhibiting near identical results, while C deviates more and, as a result,

 $^{^2}$ This is valid since $l_{\rm c}=23.5\,{\rm cm}$ and the size of the sensor array inlet is $l_{\rm RX}=0.9\,{\rm mm}$. The effective $l_{\rm RX,eff}$ of the observed section will be larger, due to effects such as scattering, but still much smaller than $l_{\rm c}$.

exhibits a slightly sharper CIR peak. The differences here are presumably due to actual deviations in the placement of the injection needles in the channel as described in Section II and to imperfections in the physical injection process, such as leakage after the pulse and disturbances of the pulse from other needles in the channel.

Figure 2 also depicts the averaged CIR for the MUMO transmission as a dotted line, encompassing all measured pulses from one color as they are transmitted in parallel to 0, 1, or 2 other colors. The lighter-shaded area around the curve represents the maximum deviation from that average in either direction. In general, the average shape of the curve is retained in the MUMO case for all colors. However, we can observe some changes. Firstly, the variations around the average increase significantly, and secondly, the average curve is decreased from the SIMO average by about 20% at the peak across all three colors. There are several contributing factors to this effect. Since the colors are not perfectly mixed when they arrive at the RX, the random layering and occlusion of some colors over others could impact how strongly the color is seen on the spectrum due to the directional illumination setup of the RX from top to bottom. This would cause a decrease in the measured concentration for some pulses. Additionally, the synchronization and alignment of the pulses across different batches of measurements for each color combination are approximate. The differing distances of the injection needles also cause varying travel times of the colors from the TX. Therefore, the MUMO averaged pulses include some inherent misalignment, evident in the yellow CIR but present for all colors, which broadens and lowers the peak. Overall, we believe the physical setup and its inaccuracies are the leading cause of the differences between SIMO and MUMO CIR.

Figure 2 also includes the MUMO bit-0 as a dashed line, i.e., the measured response in each color when any other colors are currently being received. Here, the concentration estimate stays close to zero in all cases with no cross-talk from other colors. In general, the MUMO CIRs for bit-1 and bit-0 exhibit a level of consistency high enough for reliable communication, as we will show in Section V-B.

B. Communication System Performance

A random balanced sequence of 3 times 100 bits, one 100-bit sequence for each color, was generated for conducting a preliminary examination of the communication capabilities of the testbed. The TX uses on-off-keying (OOK) to transmit the bits such that the bit from ink i in symbol period k is $s_{i,k} \in \{0,1\}$. At the RX, we implemented a simple difference detector to enable operation under the influence of ISI. To determine the estimated symbol $\hat{s}_{i,k}$, the detector identifies two samples: the sample acquired at the first sample time t_0^k within symbol period k, $c_i(t_0^k)$, and the sample with the highest value within the symbol $\max_{t \in T_k} c_i(t)$. If we define

$$\Delta_{i,k} = \max_{t \in T_k} c_i(t) - c_i(t_0^k)$$
, the decision rule is

$$\hat{s}_{i,k} = \begin{cases} 1 & \Delta_{i,k} \ge \tau \\ 0 & \Delta_{i,k} < \tau \end{cases}$$
 (12)

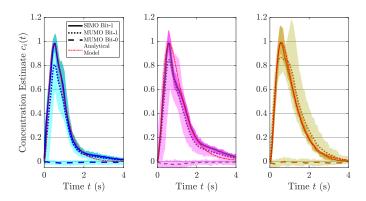


Fig. 2. Average CIR for SIMO and MUMO transmission of cyan, magenta, and yellow molecules as the estimated color concentration over time. The plot shows the SIMO CIR average, with the dark-shaded area representing the maximum deviation from the average. The best fit analytical CIR defined in Eq. (11) is shown in red. Additionally, the average MUMO transmission bit-1 and bit-0 response are shown with the lighter shaded area depicting the maximum deviation from the average.

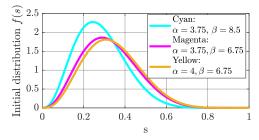


Fig. 3. Best fit initial Beta distributions f(s) as defined in Eq. 10 for the SIMO CIRs shown in Figure 2.

The detection process is applied simultaneously for all three colors using the same τ value. A preamble of 4 bit-1s is sent before the sequence to aid synchronization. The system BER within the 312 total transmitted bits across all colors is used for the performance evaluation.

The choice of threshold τ is fundamental in determining the resulting BER. Therefore, Figure 4 depicts the resulting system BER for $\tau \in [0,1]$. The evaluation was conducted for three different parameter settings of $\{T_{\rm sym}, T_{\rm inj}\}$, specifically $\{2\,{\rm s}, 0.1\,{\rm s}\}$, $\{1\,{\rm s}, 0.05\,{\rm s}\}$, and $\{0.5\,{\rm s}, 0.05\,{\rm s}\}$ with a resulting bit rate of 1.5 bps, 3 bps, and 6 bps, respectively. The results show almost error-free communication for $T_{\rm sym}=2\,{\rm s}$ and 1 s for the optimal choice of τ , with 1 error and 5 errors across all 312 bits, respectively. In both cases, ISI already plays a significant role, as we have seen in Figure 2, that the CIR takes $> 2\,{\rm s}$ to drop back to zero. For $T_{\rm sym}=0.5\,{\rm s}$, the BER rises to about 23% as ISI and variations in the CIR are difficult to overcome for the naive difference detector.

Figure 5 shows an excerpt of the raw light intensity $I^j(t)$ in each channel and the estimated concentrations $c_i(t)$ during a random bit sequence transmission with $T_{\rm sym}=2\,{\rm s}$. This side-by-side comparison showcases how the concentration estimator transforms the measured I(t) into the concentration estimate by utilizing the (anti-)correlations between the different ink colors. Additionally, we can see the significant impact of ISI on the shape of the pulses and that the difference

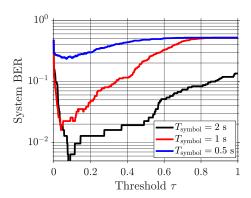


Fig. 4. System BER for a sample sequence of 312 bits sent via MUMO transmission using a range of detection thresholds τ , see Eq. (12). Results for three different values of the symbol period $T_{\rm sym} = \{0.5, 1, 2\}$ s are depicted, representing a bit rate of 6, 3, and 1.5 bps, respectively.

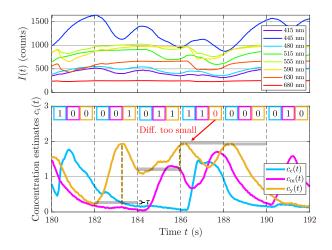


Fig. 5. Selected time frame from the received signal during the random bit sequence transmission with $T_{\mathrm{sym}}=2\,\mathrm{s}$. The plot on top depicts raw measured light intensities $I^j(t)$ for all channels, and the plot below depicts the corresponding estimated concentrations $c_i(t)$. Received bits in each symbol period are shown for the three inks at the top of the lower plot, with the single error of the entire sequence (yellow bit-1 transmitted, bit-0 detected) highlighted in red. For 4 selected symbol periods, the threshold τ and detected difference for the yellow signal are shown.

detector can deal with the vast majority of distortions. The singular error of the 312-bit sequence, caused by particularly strong ISI during a symbol with transmissions from all three colors, is highlighted in red.

VI. CONCLUSION

This work introduced a novel flow-based MC testbed design using spectral sensing and concentration estimation to differentiate between multiple color molecules, enabling real-time MUMO transmission and reception. We presented inexpensive and easily operable TX and information molecules and a non-invasive RX. Due to custom 3D-printed injection and detection casings, easy extension towards MC networks will be enabled. CIR results were presented, showing limited effects of the MUMO on the signal. The preliminary performance evaluation indicates the potential of our real-time MUMO platform to achieve single-digit bps throughput even with a straightforward detector design. Related work

has shown significant throughput gains using more sophisticated schemes [6], [9], and we plan to combine this with the demonstrated MUMO gains in future work, as well as incorporate the broad possibilities of coding, modulation, and resource management schemes using multiple molecule types to improve communication performance further. An evaluation of multiple TXs and RXs is a crucial next step, and we plan to evaluate multi-user schemes on the testbed. The real-time capabilities of the platform also enable feedback and control loops that could be crucial for evaluating various health-related sensing and actuation use cases.

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