



# Complete Genome Sequences of Hydrogenotrophic Denitrifiers

Clara Duffner,<sup>a,b</sup> Susanne Kublik,<sup>b</sup> Bärbel Fösel,<sup>b</sup> Michael Schloter,<sup>a,b</sup> Stefanie Schulz<sup>b</sup>

<sup>a</sup>Chair of Soil Science, TUM School of Life Sciences Weihenstephan, Technical University of Munich, Freising, Germany

<sup>b</sup>Research Unit Comparative Microbiome Analysis, Helmholtz Zentrum München Deutsches Forschungszentrum für Gesundheit und Umwelt, Neuherberg, Germany

**ABSTRACT** Hydrogenotrophic denitrifiers are important bacteria for nitrate removal in wastewater and aquifers. Here, we report the complete genome sequences of three hydrogenotrophic denitrifiers, namely, *Dechloromonas denitrificans* strain D110, *Ferribacterium limneticum* strain F76, and *Hydrogenophaga taeniospiralis* strain H3, all of which were isolated from a nitrate-polluted aquifer in Bavaria (Germany).

We isolated three hydrogenotrophic denitrifiers after enrichment from sediment (H3) and groundwater (D110 and F76) samples from a highly nitrate-polluted aquifer in the Hohenthann region of southeast Germany (1). Samples were obtained as described (2). Enrichment and isolation were performed under anoxic conditions with a 60% H<sub>2</sub>/10% CO<sub>2</sub>/30% N<sub>2</sub> atmosphere. Cultivation was done on mineral medium agar plates after all available nitrate and nitrite had been reduced. The mineral medium contained basal medium (3), 30 mM NaHCO<sub>3</sub> buffer, 1.5 mM NaNO<sub>3</sub>, 0.2% trace element solution (4), 0.1% selenite tungsten solution, and 0.1% vitamin solution (5). The isolates obtained were phylogenetically assigned by Sanger sequencing of the full 16S rRNA gene (primer pair 27f/1492r [6]) and aligned against the NCBI rRNA and internal transcribed spacer (ITS) databases with the nucleotide Basic Local Alignment Search Tool (BLASTn) (v2.10.0).

The complete genome sequences of *Dechloromonas denitrificans* D110, *Ferribacterium limneticum* F76, and *Hydrogenophaga taeniospiralis* H3 were obtained using single-molecule real-time (SMRT) cells and the Sequel system (Pacific Biosciences [PacBio], CA, USA). The DNA was extracted from 4.5 × 10<sup>9</sup> cells that had been grown in R2A medium at 30°C using the anion-exchange-based Genomic-tip (20/G) kit (Qiagen, Hilden, Germany). The multiplexed microbial libraries were prepared according to the procedure and checklist of the SMRTbell Express template preparation kit v2.0 (product number 101-696-100, v6 [March 2020]; PacBio). The genomic DNA was sheared to 9.5- to 12.6-kb-long fragments using g-TUBEs (Covaris, MA, USA) and further processed without additional size selection. The libraries, with a maximum expected genome size of 33 Mb, were loaded onto two SMRT cells at concentrations of 3 pM and 6 pM. Libraries were immobilized on the SMRT cells (2 h), preextended (2 h), and then sequenced on the Sequel system using v3.0 chemistry with a movie time of 10 h. The data were demultiplexed and the genomes were assembled using the HGAP4 pipeline embedded in SMRT Link v8.0.0.80529 (PacBio), with a seed coverage of 30×.

Details on the aligned subreads are summarized in Table 1. The genomes of *D. denitrificans* D110 and *F. limneticum* F76 comprised only one large contig each, which were circularized successfully with Circlator software v1.5.5 (7). The large contig of the *H. taeniospiralis* H3 genome was also circularized, while two additional small contigs of 5,700 and 2,750 bp could not be circularized.

According to CheckM software v1.1.2 (8), all three genome sequences were at least 99.2% complete, with a maximum contamination of 0.93% in the genome of *H. taeniospiralis* H3. The GC contents were similar for all three genomes, i.e., 62% for *D. denitrificans* D110, 60% for *F. limneticum* F76, and 67% for *H. taeniospiralis* H3. The genomes

**Editor** Julia A. Maresca, University of Delaware

**Copyright** © 2022 Duffner et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Stefanie Schulz, stefanie.schulz@helmholtz-muenchen.de.

The authors declare no conflict of interest.

**Received** 21 October 2021

**Accepted** 10 December 2021

**Published** 13 January 2022

**TABLE 1** SMRT Link-polished assembly results for the sequenced isolates, as well as the number of CDSs, 16S rRNAs, and tRNAs determined with NCBI PGAP

| Isolate                      | No. of realigned subreads | Mean subread length (bp) | Mean coverage (fold) | No. of contigs | Maximum contig length (bp)  | No. of CDSs | No. of 16S rRNAs | No. of tRNAs |
|------------------------------|---------------------------|--------------------------|----------------------|----------------|-----------------------------|-------------|------------------|--------------|
| <i>D. denitrificans</i> D110 | 525,543                   | 4,805                    | 511                  | 1              | 4,619,273                   | 4,301       | 4                | 63           |
| <i>F. limneticum</i> F76     | 266,526                   | 4,629                    | 259                  | 1              | 4,419,139                   | 4,128       | 3                | 58           |
| <i>H. taeniospiralis</i> H3  | 233,679                   | 5,387                    | 206                  | 3              | 5,275,671, 5,700, and 2,750 | 4,935       | 3                | 50           |

were phylogenetically assigned with the Type Strain Genome Server (TYGS) from DSMZ (9) and functionally annotated by the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) ([https://www.ncbi.nlm.nih.gov/genome/annotation\\_prok](https://www.ncbi.nlm.nih.gov/genome/annotation_prok)). The latter predicted 4,301 coding sequences (CDSs) in the genome of *D. denitrificans* D110, 4,128 CDSs in *F. limneticum* F76, and 4,935 CDSs in *H. taeniospiralis* H3 (Table 1). All genomes included genes coding for denitrification reductases, hydrogenases, and ribulose-1,5-bisphosphate carboxylase-oxygenase (RubisCO), which are all required for hydrogenotrophic denitrification.

**Data availability.** The assembly and annotation, as well as the raw reads, for the three genomes are available via BioProject [PRJNA727717](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA727717). The NCBI assembly accession numbers for the genomes are [GCA\\_020510585.1](https://www.ncbi.nlm.nih.gov/assembly/GCA_020510585.1) for *F. limneticum* F76, [GCA\\_020510685.1](https://www.ncbi.nlm.nih.gov/assembly/GCA_020510685.1) for *D. denitrificans* D110, and [GCA\\_020510445.1](https://www.ncbi.nlm.nih.gov/assembly/GCA_020510445.1) for *H. taeniospiralis* H3. The respective raw reads can be found under the accession numbers [SRR16235514](https://www.ncbi.nlm.nih.gov/assembly/SRR16235514), [SRR16235513](https://www.ncbi.nlm.nih.gov/assembly/SRR16235513), and [SRR16235512](https://www.ncbi.nlm.nih.gov/assembly/SRR16235512).

## ACKNOWLEDGMENT

C.D. received funding from the Deutsche Forschungsgemeinschaft (DFG) through the Technical University of Munich International Graduate School of Science and Engineering (IGSSE) (grant GSC 81).

## REFERENCES

- Wild L, Mayer B, Einsiedl F. 2018. Decadal delays in groundwater recovery from nitrate contamination caused by low O<sub>2</sub> reduction rates. *Water Resour Res* 54:9996–10012. <https://doi.org/10.1029/2018WR023396>.
- Duffner C, Holzapfel S, Wunderlich A, Einsiedl F, Schlöter M, Schulz S. 2021. *Dechloromonas* and close relatives prevail during hydrogenotrophic denitrification in stimulated microcosms with oxic aquifer material. *FEMS Microbiol Ecol* 97:fiab004. <https://doi.org/10.1093/femsec/fiab004>.
- Widdel F, Bak F. 1992. Gram-negative mesophilic sulfate-reducing bacteria, p 3352–3378. In Balows A, Trüper HG, Dworkin M, Harder W, Schleifer K-H (ed), *The prokaryotes: a handbook on the biology of bacteria: ecophysiology, isolation, identification, applications*. Springer, New York, NY. [https://doi.org/10.1007/978-1-4757-2191-1\\_21](https://doi.org/10.1007/978-1-4757-2191-1_21).
- Widdel F, Kohring G-W, Mayer F. 1983. Studies on dissimilatory sulfate-reducing bacteria that decompose fatty acids. *Arch Microbiol* 134:286–294. <https://doi.org/10.1007/BF00407804>.
- Balch WE, Fox GE, Magrum LJ, Woese CR, Wolfe RS. 1979. Methanogens: reevaluation of a unique biological group. *Microbiol Rev* 43:260–296. <https://doi.org/10.1128/mr.43.2.260-296.1979>.
- Lane DJ. 1991. 16S/23S rRNA sequencing, p 115–175. In Stackebrandt E, Goodfellow M (ed), *Nucleic acid techniques in bacterial systematics*. John Wiley and Sons, New York, NY.
- Hunt M, Silva ND, Otto TD, Parkhill J, Keane JA, Harris SR. 2015. Circlator: automated circularization of genome assemblies using long sequencing reads. *Genome Biol* 16:294. <https://doi.org/10.1186/s13059-015-0849-0>.
- Parks DH, Imelfort M, Skennerton CT, Hugenholtz P, Tyson GW. 2015. CheckM: assessing the quality of microbial genomes recovered from isolates, single cells, and metagenomes. *Genome Res* 25:1043–1055. <https://doi.org/10.1101/gr.186072.114>.
- Meier-Kolthoff JP, Göker M. 2019. TYGS is an automated high-throughput platform for state-of-the-art genome-based taxonomy. *Nat Commun* 10:2182. <https://doi.org/10.1038/s41467-019-10210-3>.