

PET Imaging of Self-Assembled ^{18}F -Labelled Pd_2L_4 Metallacages for Anticancer Drug Delivery

Raúl Cosialls,^[a, c] Cristina Simó,^[b, c] Salvador Borrós,^[d] Vanessa Gómez-Vallejo,^[b] Claudia Schmidt,^[e] Jordi Llop,^{*,[b]} Ana B. Cuenca,^{*,[a, f]} and Angela Casini^{*,[e, g]}

Abstract: To advance the design of self-assembled metallosupramolecular architectures as new generation theranostic agents, the synthesis of ^{18}F -labelled $[\text{Pd}_2\text{L}_4]^{4+}$ metallacages is reported. Different spectroscopic and bio-analytical methods support the formation of the host-guest cage-cisplatin

complex. The biodistribution profiles of one of the cages, alone or encapsulating cisplatin have been studied by PET/CT imaging in healthy mice in vivo, in combination to ICP-MS ex vivo.

Introduction

Supramolecular systems are attracting increasing attention in the development of nanomaterials for different applications.^[1] Self-assembled porous metallacages are particularly attractive supramolecular coordination complexes featuring discrete mo-

lecular 3-dimensional (3D)-architectures with various appealing applications, including storage, separation, catalysis, recognition, as well as light emitting materials amongst others.^[2] In medicine, the biological properties of these well-defined *molecular vessels* have recently been gaining momentum for drug delivery of therapeutics^[3] and imaging agents,^[4] as well as for the development of novel *theranostic* platforms.^[5] Among the different metal assemblies, some of us have focused on the advantages of a particular type of palladium-based metallacage scaffold $[\text{Pd}_2\text{L}_4]^{4+}$ ($\text{L} = 3,5\text{-bis}(3\text{-ethynylpyridine})\text{phenyl}$) as potential delivery system for the well-established anticancer drug cisplatin.^[6] Thus, we developed the *exo*-functionalization of the ligands to add different bioactive components, including fluorescent tags^[7,8,9] and peptidic domains.^[10] It was also demonstrated that encapsulation of cisplatin in integrin targeted metallacages leads to reduced nephrotoxicity with respect to free cisplatin.^[11] Encapsulated cisplatin showed also higher in vitro cytotoxicity against cancer cells expressing the integrin receptors.^[11] Noteworthy, $[\text{Pd}_2\text{L}_4]^{4+}$ cages tethered to a blood brain barrier (BBB)-translocating peptide and encapsulating radioactive pertechnetate were recently studied for their biodistribution in mouse models, and demonstrated the stability of the host-guest (cage-pertechnetate) complex and its brain penetration capability.^[12]

While all the prominent proof-of-concept reports mentioned above can give a glimpse of a bright future for the use of metallacages as drug delivery systems, in vivo imaging studies of these supramolecular entities are still scarce.^[13,4] Most importantly, their design as novel theranostic platforms featuring both therapeutic and imaging modalities is still in its infancy. Besides, the ultimate question of the structural integrity of the metallacages upon in vivo injection remains unresolved. In this context, we considered that ^{18}F -labelling followed by in vivo Positron Emission Tomography (PET) imaging might offer interesting insights to that end. Additionally, and in contrast to optical imaging techniques, PET is fully translational into the clinical setting to assess whole body biodistribution. We considered the positron emitter fluorine-18 (^{18}F) as an appropriate radionuclide due to its wide availability and

[a] R. Cosialls, Dr. A. B. Cuenca

BISI-Bonds group

Dept. of Organic and Pharmaceutical Chemistry

Institut Químic de Sarrià, URL

Via Augusta 390, 08017 Barcelona (Spain)

E-mail: anabelen.cuenca@iqs.url.edu

[b] C. Simó, Dr. V. Gómez-Vallejo, Dr. J. Llop

CIC biomaGUNE

Basque Research and Technology Alliance (BRTA)

Paseo Miramón 182, 20014 San Sebastián, Gipuzkoa (Spain)

E-mail: jlllop@cicbiomagune.es

[c] R. Cosialls, C. Simó

Department of Organic and Inorganic Chemistry,

Universidad del País Vasco (UPV/EHU),

Barrio Sarriena s/n, 48940 Leioa, Bizkaia, (Spain)

[d] Prof. S. Borrós

Grup d'Enginyeria de Materials (GEMAT)

Institut Químic de Sarrià, URL

Via Augusta 390, 08017 Barcelona (Spain)

[e] Dr. C. Schmidt, Prof. A. Casini

Chair of Medicinal and Bioinorganic chemistry

Department of Chemistry

Technical University of Munich

Lichtenbergstr. 4, 85748 Garching b. München (Germany)

E-mail: angela.casini@tum.de

[f] Dr. A. B. Cuenca

Centro de Innovación en Química Avanzada (ORFEO-CINQA) (Spain)

[g] Prof. A. Casini

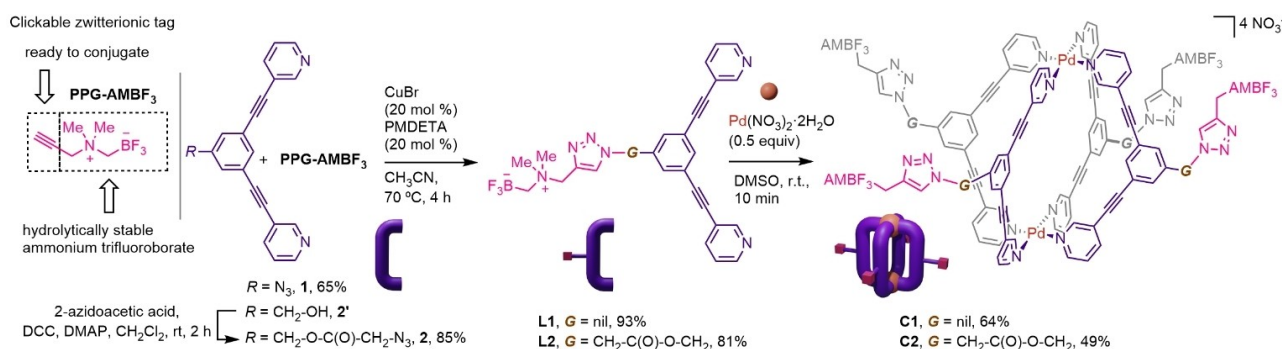
Munich Data Science Institute

Technical University of Munich

85748 Garching b. München (Germany)

Supporting information for this article is available on the WWW under <https://doi.org/10.1002/chem.202202604>

© 2022 The Authors. Chemistry - A European Journal published by Wiley-VCH GmbH. This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.



Scheme 1. Structure of the clickable zwitterionic ammonium trifluoroborate (AMBF₃) tag PPG-AMBF₃, and synthesis of ligands **L1** and **L2** and of the AMBF₃-modified cages **C1** and **C2**.

favorable properties (relatively long half-life and short positron range).

Results and Discussion

Among the possible ¹⁸F-labelling strategies, our attention was drawn away from the common ¹⁸F-carbon bond-forming processes, focusing instead on the [¹⁹F]-to-[¹⁸F]-boron isotopic exchange using ammonium trifluoroborate functionalities (AMBF₃). These types of prosthetic groups, introduced by the Perrin laboratory, allow for efficient, single-step, aqueous ¹⁸F-labelling using as the labelling agent aqueous [¹⁸F]fluoride directly produced in the cyclotron, thus avoiding time-consuming evaporation steps for solvent exchange.^[14a,b] As an efficient route to append the AMBF₃ fragment to the metallacage, we used a classical “click” approach, via a propargyl-bearing AMBF₃ reagent (PPG-AMBF₃,^[14c] Scheme 1) that could be conjugated to the azide-modified cage ligand by Cu-catalyzed alkyne-azide cycloaddition reaction (CuAAC). Hence, two azide-functionalized bis(pyridyl)ethynyl ligand precursors were initially considered, namely the derivative **1**^[9a] with the azide attached directly to the central phenylene unit, and compound **2** in which a spacer is introduced between the ligand portion and the azide (see Figure S1a–b). The latter was prepared in an 85% yield by Steglich-type esterification of the benzylic alcohol *exo*-functionalized ligand **2'**^[15] with 2-azidoacetic acid. Next, the AMBF₃-modified ligands **L1** and **L2**, were synthesized through a CuAAC reaction of azides **1** and **2** with PPG-AMBF₃ using the CuBr-PMDETA (*N,N,N',N',N'*-pentamethyldiethylenetriamine) as catalyst (Scheme 1).¹⁶ Data related to the NMR characterization of the ligands are reported in Figure S3–S4 in the Supporting Information.

With ligands **L1** and **L2** in hand, we proceeded to the self-assembly of the homoleptic cages **C1** and **C2** with palladium dinitrate precursor which reached completion within 10 min after mixing (Scheme 1). The assembly of both [Pd₂L₄]⁴⁺ cages was unequivocally confirmed by ¹H, DOSY NMR experiments, as well as by high-resolution electrospray mass spectrometry (HR-ESI-MS) (Figure S5–S9 in the Supporting Information).^[6]

Next, we verified that the newly assembled cages retained their ability to encapsulate cisplatin, an important feature for their potential as drug delivery systems. X-ray diffraction analysis has shown that Pd₂L₄ cages can encapsulate up to 2 cisplatin molecules.^[3c,6a] It should be noted that the cage's 3,5-bis(3-ethynylpyridine)phenyl) scaffold creates a hydrophobic cavity whereby cisplatin encapsulation should be favoured over occupancy of the cavity by water molecules in solution.^[17] Thus, the host-guest properties of **C1** and **C2** were studied by ¹H and ¹⁹⁵Pt NMR spectroscopy, as well as by HR-ESI-MS (Figure S10–S14). Metallacages **C1** or **C2** (1 equiv.) were dissolved in DMF-*d*₇ and then up to 3 equiv. of cisplatin were sequentially added stepwise, with each addition followed by a 10-min sonication step. The ¹H NMR spectrum recorded after the last addition revealed an identifiable downfield chemical shift of the *exo*-facing proton H_b (Δδ = 0.02 ppm) for both cages, previously observed for similar cage systems upon cisplatin encapsulation.^[3c,6b] In addition, an observable upfield chemical shift (Δδ = 0.02 ppm) was found for the endohedral cavity-facing H_a, a plausible sign of the guest presence (Figure 1 and Figure S10). Afterwards, ¹⁹⁵Pt NMR spectroscopy was also utilized to gain further insights into the cisplatin encapsulation in these cavities. Compared with free cisplatin, an upfield

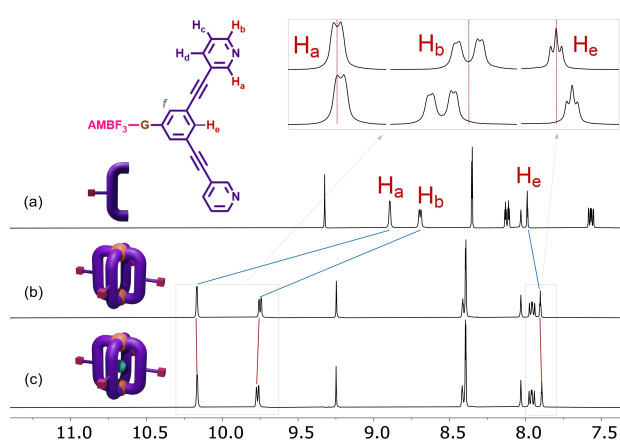


Figure 1. ¹H NMR spectrum (DMF-*d*₇, 298 K) of ligand **L1** (a), cage **C1**(NO₃)₄ (b) and (c) host-guest adduct [**C1**(NO₃)₄·(cisplatin)].

chemical shift of about -2 ppm was observed upon addition of 2 equiv. of cisplatin to a DMF-*d*₇ solution of complex **C1** (Figure S12), in line with previous studies,^[9] and corroborating the idea of cisplatin encapsulation.

HR-ESI-MS analysis of [**C1**(NO₃)₄⊃cisplatin] and [**C2**(NO₃)₄⊃cisplatin], respectively, provided further evidence of the cage-cisplatin encapsulation properties. For example, when a 1:2 mixture of **C1** and cisplatin in DMF was analysed, two clear peaks at $m/z=839.8452$ and $m/z=1290.7616$ appeared that could be unambiguously assigned to [Pd₂(L1)₄(NO₃)₄⊃cisplatin]³⁺ and [Pd₂(L1)₄(NO₃)₂⊃cisplatin]²⁺ host-guest complexes, respectively (see Figure S13a–d). Moreover, a peak at $m/z=1440.7548$ was attributed to [**C1**(NO₃)₂⊃(cisplatin)₂]²⁺ adducts. These species were also detected in 4% DMSO in water. Analogous behaviour was observed for cage **C2** (see Figure S14a–c).

The encapsulation of cisplatin in **C1** was also studied by ¹H DOSY NMR in DMF-*d*₇/D₂O (90:10). Cisplatin alone in DMF shows a broad signal at ca. 4.2 ppm (H from NH₃), with a diffusion coefficient ca. 6×10^{-6} m²/s.¹¹ Upon addition of 2 equiv. cisplatin to 1 equiv. metallacage, the typical signal of free cisplatin disappeared in the DOSY plot (although marginally present in the ¹H spectrum likely due to the fast exchange of free vs. encapsulated cisplatin species), while significant broadening of the cage signals were observed, accompanied by small shifts in their diffusion coefficients (Figure S15, spectrum c), suggesting that the cage cavity has been saturated to form the [**C1**⊃cisplatin] host-guest complex.¹¹ The cisplatin peak reappears only upon addition of a third equivalent of cisplatin to the sample (Figure S15, spectrum d), which should not undergo encapsulation.^[11] These data agree with the aforementioned HR-ESI-MS studies.

The feasibility of the ¹⁸F-isotopic labelling of *exo*-functionalized cages was then assessed. Initially, direct ¹⁹F-to-[¹⁸F] exchange (¹⁸F-EX) from the preassembled **C1** and **C2** was attempted as a more straightforward approach. To this end, a wet no carrier added solution of ¹⁸F-fluoride ion was generated and subsequently trapped on an anion exchange resin (QMA Cartridge). The ¹⁸F-fluoride was next eluted with isotonic saline and then added to a vial containing unlabelled metallacages **C1** and **C2** in a DMF/water-pyridazine-HCl buffer (pH=2) following previously established procedures.^[14a] The mixture was heated at 90 °C for 30–60 min. Unfortunately, the radio-HPLC analysis of the ¹⁸F-EX reaction evidenced major disassembly of the Pd²⁺ cages to the corresponding isotopically marked ¹⁸F-L1 and ¹⁸F-L2 ligands. Nevertheless, this experiment proved the efficient ¹⁸F-EX reaction of the ligand framework prior to the self-assembling process.

Given the fast kinetics of the assembly (few minutes), the final generation of the ¹⁸F-labelled-**C1** was straightforwardly achieved within the radioisotope half-life time. Thus, ¹⁸F-L1 was prepared as above with slight modification of the experimental conditions (T=85 °C; reaction time=45 min, see Supporting Information for details). After confirmation of the quantitative formation of ¹⁸F-L1 by radio-HPLC (Figure S21, chromatogram a), the labelled ¹⁸F-C1 cage was assembled by mixing ¹⁸F-L1 (2 equiv.) and 1 equiv. of Pd(NO₃)₂·2H₂O in DMSO for 30 min in

95% chromatographic yield (see Figure S21, chromatogram b). The same conditions were applied to achieve ¹⁸F-L2 and cage ¹⁸F-C2. However, we decided to pursue the *in vivo* study only with **L1** since chromatographic yield of ligand ¹⁸F-L2 proved lower (40%) than the corresponding ¹⁸F-L1. Afterwards, we proceeded with encapsulation of cisplatin in the radio-labelled cage. It should be noted that the radio-HPLC retention time of the parent cage ¹⁸F-C1 and the same cage incubated for 5 min with 2 equiv. of cisplatin were virtually indistinguishable (Figure S21, chromatogram c). However, inductively coupled plasma optical emission spectrometry (ICP-MS) analysis of the manually collected fractions of the latter sample did reveal presence of Pt for cisplatin loaded ¹⁸F-C1 (ca. 0.5 ng) but not in samples of free ¹⁸F-C1 and ¹⁸F-L1 (ca. 0.03 and 0.05 ng, respectively) used as controls; an observation that constitutes another indirect evidence of cisplatin encapsulation (Figure S22).

In parallel, the stability of cage **C1** over several hours was assessed through a series of ¹H NMR experiments in different conditions that included 100% DMSO-*d*₆ and 4% DMSO-*d*₆ in D₂O (Figure S16–S17). The stability of the [**C1**(NO₃)₄⊃cisplatin] complex was also monitored by ¹H NMR over time in 4% DMSO-*d*₆ in D₂O, and the results showed that the signals of the cage remain prominent only during the first hour (Figure S18). The origin of this enhanced cage instability in the presence of cisplatin is presently under investigation. Moreover, the stability of the free cage **C1** (0.15 mM injection concentration, 4% DMSO in H₂O) was also gauged by HR-ESI-MS. The resulting spectra showed the prominent presence of intact [**C1**(NO₃)_n]²⁺ species (Figure S19). Importantly, the presence of similar **C1**-related species was also observed in 4% DMSO in saline solution (up to 0.09% NaCl) (Figure S20).

Next, biodistribution of ¹⁸F-L1, ¹⁸F-C1 and cisplatin loaded ¹⁸F-C1 was investigated in healthy mice using PET imaging in combination with computed tomography (CT). PET acquisitions were started immediately after administration of labelled compounds and dynamic scans were acquired for 60 min. Quantification analysis of PET images were performed only in those organs clearly visualized on CT images (brain, heart, lungs, liver, kidneys, and bladder). Visual inspection of PET images obtained over the first 10 min after administration (Figure 2a) showed a very different profile for ¹⁸F-L1, with presence of radioactivity in heart, lungs, intestines and gall bladder, with respect to cage ¹⁸F-C1. The latter showed major accumulation in the liver and lower accumulation in the kidneys (both statistically significant in the time frame 0–4 min), as confirmed by image quantification (Figure 2b). Images also suggest lower accumulation of ¹⁸F-C1 in the gall bladder, although no quantification was carried out in this vesicle as, due to its small size, results could be subjected to severe partial volume effect. The observed differences in biodistribution provide evidence that the species arising upon injection of the supramolecular structure is distinct from the free ligand. In contrast, the biodistribution profile for ¹⁸F-L1 and ¹⁸F-[**C1**(NO₃)₄⊃cisplatin] showed more similarities. This result suggests that a possible disassembly of the cisplatin loaded ¹⁸F-C1 may occur after *in vivo* injection, in accordance with the above-mentioned NMR studies. Accumulation of radioactivity in the different

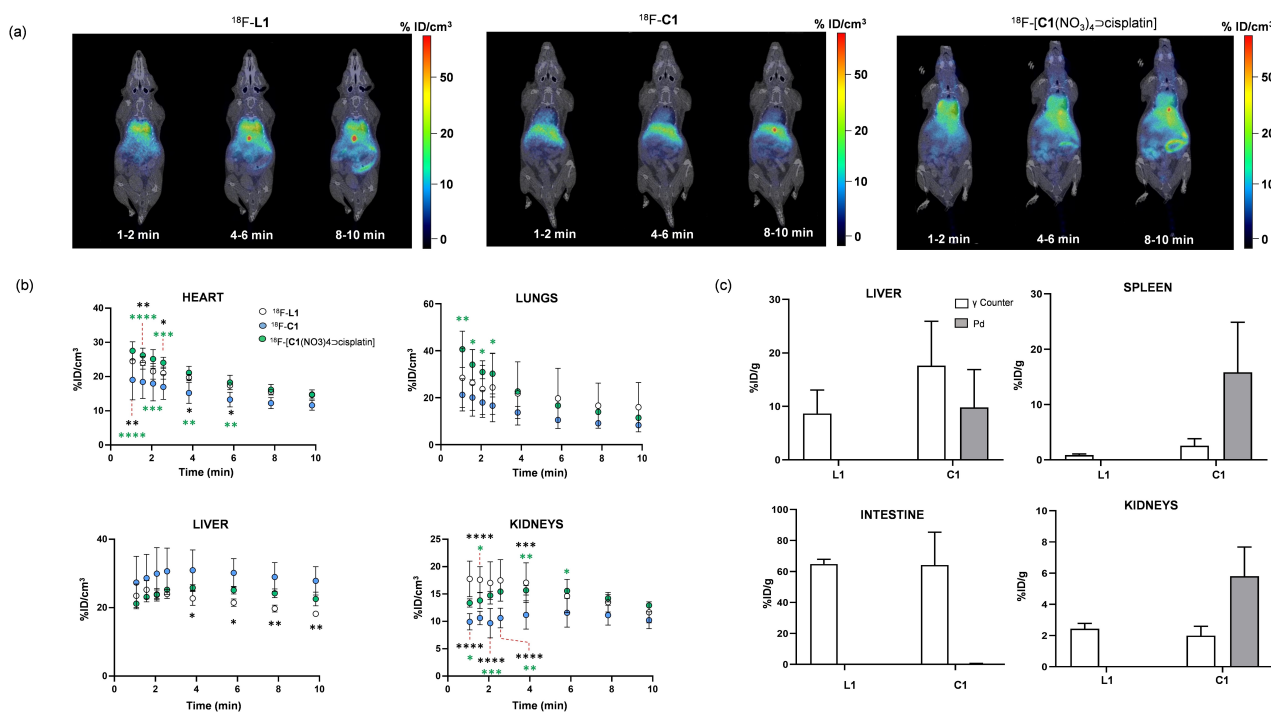


Figure 2. (a) Representative PET-CT images (coronal projections of PET images co-registered with representative CT slices) obtained at different time points after administration of ^{18}F -L1, ^{18}F -C1 and ^{18}F -[C1(NO₃)₄]cisplatin; (b) Time activity curves obtained from quantification of PET images. Values are expressed as percentage of injected dose per cubic centimetre of tissue (%ID/cm³). Values are expressed as mean \pm standard deviation ($n=4$); probability values are depicted in black for ^{18}F -L1 vs. ^{18}F -C1 and in green for ^{18}F -C1 vs. ^{18}F -[C1(NO₃)₄]cisplatin as $P < 0.05$, *, $P < 0.01$, **, $P < 0.001$, ***, and $P < 0.0001$, ****; (c) Concentration of injected compound per gram of tissue as determined by dissection/gamma counting (white bars) and ICP-MS (Pd; grey bars) after animal sacrifice; intestine represent only small intestine.

organs reached similar values at longer time points, irrespectively of the administered compound (see Figure S23 for representative PET-CT images). The lack of uptake in the bone confirms the absence of ^{18}F -defluorination of our ^{19}F -to- ^{18}F -trifluoroborate labelling strategy. After 60 min, ex vivo analysis based on dissection and gamma counting was carried out for ^{18}F -L1 and ^{18}F -C1 (Figure 2c). In this case, both compounds presented similar distribution pattern with major excretion through intestines (ca. 60% of injected dose per gram of tissue - %ID/g - in the small intestine), further suggesting a possible non-negligible disassembly of ^{18}F -C1 to ^{18}F -L1. Analysis of the palladium levels in selected organs was performed by ICP-MS after 60 min upon injection of ^{18}F -C1 (Figure 2c) as another way of differentiating between the fates of the cage and the disassembled ligand. Noteworthy, Pd accumulation appears to be mostly uncoupled with respect to the ligand biodistribution. In fact, Pd was detected in spleen, kidney and liver, while being virtually absent in the intestine, where the ligand ^{18}F -L1 prominently accumulates (Figure 2c). This result is in line with the observed different biodistribution among the ^{18}F -L1 and ^{18}F -C1.

Conclusion

In conclusion, we have reported on the straightforward synthesis and purification of metallacages as possible drug delivery

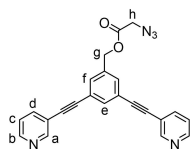
systems and studied their biodistribution in vivo by PET imaging. The obtained results show that the species arising upon cages injection accumulates in different organs with respect to their ligands in the early time points. Moreover, cisplatin encapsulation seems to favour cage disassembly in vivo, as suggested by PET. Certainly, further optimization of the ligand system, for example using more electron-donating tripyridyl ligands,^[18] is necessary to enable kinetically robust cage complexes. Alternatively, the use of Pt(II) ions instead of Pd(II) could also increase the kinetic stability, although the self-assembly may require different reaction conditions not necessarily compatible with the radiolabelling procedure.¹⁹ The cisplatin encapsulation process also requires in-depth investigation and should be performed on targeted and more hydrophilic cage systems. Overall, owing to their unique physicochemical properties, metal-coordinated supramolecular self-assemblies, including the selected metallacages, can bridge the boundary between traditional inorganic and organic materials, and our work further progresses their design for biomedical applications.

Experimental Section

Materials and methods: All commercially acquired reagents were used as received unless indicated otherwise. 2-azidoacetic acid,^[20] 3,3'-((5-azido-1,3-phenylene)bis(ethyne-2,1-diyl))dipyridine (1),^[9a] (3,5-bis(pyridin-3-ylethynyl)phenyl)methanol (2)^[6b] and ((dimeth-

yl(prop-2-yn-1-yl)ammonio)methyl)trifluoroborate (PPG-AMBF₃)^[14c] were prepared according to literature procedures or with slight modifications. HPLC grade ethanol, methanol and acetonitrile were purchased from Scharlab (Sentmenat, Barcelona, Spain). Reactions requiring inert atmosphere were conducted under argon atmosphere using standard Schlenk line techniques. Thin layer chromatography (TLC) was performed using Merck plastic-backed plates of TLC Silica gel 60 F254; the plates were revealed using UV light at 254 nm or by staining using potassium permanganate. Standard Flash Column chromatography was accomplished using Merck silica gel (60 Å pore size, 70–230 μm mesh size). Automated Flash Column chromatography was performed by a Teledyne ISCO CombiFlash Rf200 system through pre-packed RediSep Rf silica gel columns. HRMS data were acquired on a X500B SCIEX QTOF high-resolution mass spectrometer (ESI mode). Spectroscopic experiments for the characterization of compounds and encapsulation studies were carried out at the Structural Determination facility of IQS on a Varian 400 NMR spectrometer (400 MHz for ¹H, 100.5 MHz for ¹³C, 376 MHz for ¹⁹F and 128 MHz for ¹¹B). ¹⁹⁵Pt and ¹H DOSY experiments were performed at the NMR unit of Universitat de Barcelona on a Bruker Avance III 400 MHz spectrometer and at TUM on a Bruker Avance III 500 MHz spectrometer. Chemical shifts (δ_{HT}) are quoted in parts per million (ppm) and referenced to the appropriate NMR resonance, which for ¹H measurements would correspond to the residual portion component of the deuterated solvent. The ¹⁹F and ¹¹B chemical shift are referenced relative to CFCl₃ and BF₃·Et₂O resonance at 0.00 ppm, respectively. The ¹⁹⁵Pt chemical shift was referenced using an external reference of K₂PtCl₄ in D₂O (−1610 ppm). Spin-spin coupling constants (J) are reported in Hertz (Hz). Infrared spectra were recorded on a Thermo Scientific Nicolet iS10 FTIR spectrophotometer equipped with Smart iTR window and are reported in cm^{−1}. Mediterranean C18 column (4.6 × 150 mm, 5 μm) as stationary phase and 0.1 % TFA water/acetonitrile (0 min 10 % acetonitrile; 0–2 min 20 % acetonitrile; 2–10 min 70 % acetonitrile; 10–14 min 70 % acetonitrile; 14–16 min 10 % acetonitrile; 16–20 min 10 % acetonitrile) as mobile phase at a flow rate of 1 mL/min and wavelength of 254 nm. ICP-MS measurements were performed on a Thermo iCAP Q ICP-MS instrument.

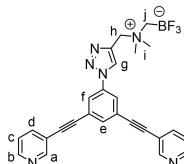
Synthesis of azide precursor (2): (3,5-bis(pyridin-3-ylethynyl)phenyl)methanol (2') (400 mg, 1.3 mmol, 1 equiv.), 2-azidoacetic acid (0.15 mL, 2.0 mmol, 1.5 equiv.) and 4-dimethylaminopyridine (DMAP) (31 mg, 0.26 mmol, 0.2 equiv.) were charged into an oven-dried 25 mL Schlenk tube and dissolved in anhydrous CH₂Cl₂ (15 mL). Then, *N,N'*-dicyclohexylcarbodiimide (DCC) (425 mg, 2.0 mmol, 1.5 equiv.) was added to the tube and the mixture was allowed to stir at room temperature for 2 h. At this point, the reaction mixture was filtered and evaporated to dryness. The product was isolated using automated flash-chromatography eluting with 1:0 to 0:1 cyclohexane:AcOEt gradient mixture (R_f = 0.3 in AcOEt). Pale orange powder, 429 mg, 85 %.



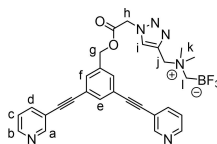
¹H NMR (DMSO-*d*₆, 400 MHz) δ 8.79 (dd, *J* = 2.2, 0.9 Hz, 2H, H_a), 8.62 (dd, *J* = 4.9, 1.7 Hz, 2H, H_b), 8.01 (ddd, *J* = 7.9, 2.2, 1.7 Hz, 2H, H_d), 7.80 (t, *J* = 1.6 Hz, 1H, H_e), 7.70 (dd, *J* = 1.5, 0.7 Hz, 2H, H_f), 7.49 (ddd, *J* = 7.9, 4.9, 0.9 Hz, 2H, H_c), 5.26 (s, 2H, H_g), 4.25 (s, 2H, H_h). ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 168.7, 151.7, 149.4, 138.7, 137.3, 133.8, 131.4, 123.7, 122.6, 118.9, 90.8, 87.3, 65.3, 49.5. FTIR (KBr) cm^{−1}: 3032 (ar C–H st), 2930 (C–H st), 2108 (N₃ st), 1749 (C=O st), 1595, 1478, 1407, 1286, 1186, 1023, 805, 704. HRMS-ESI: calc. for C₂₃H₁₆N₃O₂ [M + H]⁺: *m/z* = 394.1299; found 394.1286.

Synthesis of ligands L1 and L2

General procedure A: The corresponding azide-functionalized bis(pyridyl)ethynyl substrate (0.62 mmol, 1.5 equiv.) and ((dimethyl(prop-2-yn-1-yl)ammonio)methyl)trifluoroborate (PPG-AMBF₃) (68 mg, 0.41 mmol, 1 equiv.) were charged into an oven-dried 25 mL Schlenk tube and dissolved in anhydrous CH₃CN (19 mL). Then, a solution of CuBr (12 mg, 0.083 mmol, 0.2 equiv.) and *N,N,N',N',N''*-pentamethyldiethylenetriamine (PMDETA) (17 μL, 0.083 mmol, 0.2 equiv.) in anhydrous CH₃CN (1 mL), previously bubbled with argon for 15 min, was added. The mixture was allowed to stir at 70 °C for 4 h. After cooling to room temperature, the mixture was evaporated to dryness and the product was isolated using automatic flash-chromatography with CH₂Cl₂:MeOH mixtures as indicated.



(L1) Prepared following the *General procedure A* from 3,3'-(5-azido-1,3-phenylene)bis(ethyne-2,1-diyl)dipyridine (1) and purified using automatic flash-chromatography eluting with 1:0 to 95:5 CH₂Cl₂:MeOH gradient mixture (R_f = 0.4 in CH₂Cl₂:MeOH 9:1). Pale orange powder, 178 mg, 93 % yield. ¹H NMR (DMSO-*d*₆, 400 MHz) δ 9.21 (s, 1H, H_g), 8.83 (dd, *J* = 2.2, 0.9 Hz, 2H, H_a), 8.65 (dd, *J* = 4.8, 1.7 Hz, 2H, H_b), 8.28 (d, *J* = 1.4 Hz, 2H, H_f), 8.05 (ddd, *J* = 7.9, 2.2, 1.7 Hz, 2H, H_d), 7.96 (t, *J* = 1.4 Hz, 1H, H_e), 7.52 (ddd, *J* = 7.9, 4.9, 0.9 Hz, 2H, H_c), 4.65 (s, 2H, H_h), 3.01 (s, 6H, H_i), 2.37 (q, *J*_{H,F} = 4.8 Hz, 2H, H_j). ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 151.8, 149.6, 138.8, 137.6, 137.0, 134.1, 126.6, 124.0, 123.8, 123.3, 118.7, 90.0, 88.4, 60.2, 51.7. ¹⁹F NMR (DMSO-*d*₆, 376 MHz) δ −135.1. ¹¹B NMR (DMSO-*d*₆, 128 MHz) δ 1.93. ¹H DOSY NMR (400 MHz, DMF-*d*₇): *D* = 4.75 × 10^{−6} cm²/seg. FTIR (ATR) cm^{−1}: 3118, 3026, 1590, 1475, 1413, 1036, 987, 967, 899, 889, 805, 700. HRMS-ESI: calc. for C₂₆H₂₃BF₃N₆ [M + H]⁺: *m/z* = 487.2024; found 487.2015.



(L2) Prepared following the *General procedure A* from 3,5-bis(pyridin-3-ylethynyl)benzyl 2-azidoacetate (2) and purified using automatic flash-chromatography eluting with 1:0 to 95:5 CH₂Cl₂:MeOH gradient mixture (R_f = 0.2 in CH₂Cl₂:MeOH 9:1). Pale orange powder, 187 mg, 81 % yield.

¹H NMR (DMSO-*d*₆, 400 MHz) δ 8.80 (dd, *J* = 2.2, 0.9 Hz, 2H, H_a), 8.63 (dd, *J* = 4.9, 1.7 Hz, 2H, H_b), 8.42 (s, 1H, H_e), 8.02 (ddd, *J* = 7.9, 2.2, 1.7 Hz, 2H, H_d), 7.81 (t, *J* = 1.6 Hz, 1H, H_e), 7.70 (dd, *J* = 1.4, 0.7 Hz, 2H, H_f), 7.50 (ddd, *J* = 7.9, 4.9, 0.9 Hz, 2H, H_c), 5.63 (s, 2H, H_h), 5.28 (s, 2H, H_g), 4.56 (s, 2H, H_i), 2.93 (s, 6H, H_j), 2.22 (q, *J*_{H,F} = 4.7 Hz, 2H, H_k). ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 167.1, 151.7, 149.3, 138.7, 137.11, 136.5, 133.9, 131.4, 129.6, 123.9, 122.6, 119.2, 90.8, 87.4, 65.7, 59.9, 51.8, 50.6. ¹⁹F NMR (DMSO-*d*₆, 376 MHz) δ −135.3. ¹¹B NMR (DMSO-*d*₆, 128 MHz) δ 1.87. FTIR (ATR) cm^{−1}: 3148, 3030, 2956 (C–H st), 1752 (C=O st), 1596, 1477, 1407, 1199, 1022, 992, 969, 897, 804, 702. HRMS-ESI: calc. for C₂₉H₂₇BF₃N₆O₂ [M + H]⁺: *m/z* = 559.2235; found 559.2210.

Synthesis of metallacages C1 and C2

General procedure B: The corresponding AMBF₃-containing ligand (0.1 mmol, 2 equiv.) and Pd(NO₃)₂·2H₂O (12 mg, 0.05 mmol, 1 equiv.) were charged into a 50 mL conical centrifuge tube and dissolved in DMSO (4 mL). The mixture was allowed to stir at room temperature for 1 h. At this point, the product was precipitated by addition of acetone (4 mL) and diethyl ether (40 mL). The mixture was centrifugated, decanted and the obtained solid was washed with diethyl ether (3 × 5 mL). The product was dried under high vacuum.

(C1) Prepared following the *General procedure B* from **L1**. Yellow powder, 40 mg, 64%. ¹H NMR (DMSO-*d*₆, 400 MHz) δ 9.81 (d, *J* = 2.0 Hz, 2H, H₃), 9.46 (dd, *J* = 5.9, 1.4 Hz, 2H, H_b), 9.17 (s, 1H, H_g), 8.37 (d, *J* = 1.4 Hz, 2H, H_i), 8.31 (dt, *J* = 8.1, 1.5 Hz, 2H, H_d), 8.02 (t, *J* = 1.4 Hz, 1H, H₂), 7.87 (dd, *J* = 8.0, 5.8 Hz, 2H, H_c), 4.62 (s, 2H, H_h), 2.97 (s, 6H, H₁), 2.32 (q, *J*_{H-F} = 4.7 Hz, 2H, H_f). ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 153.1, 151.0, 143.0, 137.7, 137.3, 133.9, 127.5, 126.6, 124.5, 123.3, 121.9, 92.5, 86.4, 60.0, 51.7. ¹⁹F NMR (DMSO-*d*₆, 376 MHz) δ -135.1. ¹¹B NMR (DMSO-*d*₆, 128 MHz) δ 6.94. ¹H DOSY NMR (400 MHz, DMF-*d*₇): *D* = 2.1 × 10⁻⁶ cm²/seg. FTIR (ATR) cm⁻¹: 3070, 1589, 1477, 1325, 1195, 1012, 882, 815, 695. HRMS-ESI: calc. for [Pd₂L₄(NO₃)₁]³⁺: *m/z* = 739.8558; found 739.8604.

(C2) Prepared following the *General procedure B* from **L2**. Orange powder, 33 mg, 49%. ¹H NMR (DMSO-*d*₆, 400 MHz) δ 9.74 (d, *J* = 2.1 Hz, 2H, H₃), 9.42 (dd, *J* = 5.9, 1.4 Hz, 2H, H_b), 8.38 (s, 1H, H_g), 8.27 (dt, *J* = 8.0, 1.5 Hz, 2H, H_i), 7.90 (t, *J* = 1.5 Hz, 1H, H₂), 7.84 (dd, *J* = 8.0, 5.8 Hz, 2H, H_c), 7.76 (d, *J* = 1.5 Hz, 2H, H_d), 5.57 (s, 2H, H_h), 5.26 (s, 2H, H_e), 4.53 (s, 2H, H_f), 2.91 (s, 6H, H₁), 2.18 (q, *J*_{H-F} = 4.8 Hz, 2H, H_f). ¹³C NMR (DMSO-*d*₆, 100 MHz) δ 167.0, 152.9, 150.7, 142.9, 137.6, 136.5, 133.8, 132.6, 129.5, 127.4, 122.1, 122.0, 93.3, 85.4, 65.5, 59.8, 51.9, 50.5. ¹⁹F NMR (DMSO-*d*₆, 376 MHz) δ -135.3. ¹¹B NMR (DMSO-*d*₆, 128 MHz) δ 7.01. FTIR (ATR) cm⁻¹: 3067, 1753, 1479, 1332, 1196, 1023, 1007, 970, 897, 818, 695. HRMS-ESI: calc. for [Pd₂L₄(NO₃)₁]³⁺: *m/z* = 835.8887; found 835.8833.

Cisplatin encapsulation and stability experiments: Experimental procedures for the encapsulation studies of **C1** and **C2** by ¹H NMR, ¹⁹⁵Pt NMR, DOSY NMR and HR-ESI-MS, along with **C1** stability experiments by ¹H NMR and HR-ESI-MS are provided in the Supporting Information.

Radiolabelling

Radiolabelling of ¹⁸F-L1: **L1** was labelled with fluorine-18 by isotopic exchange reaction. Briefly, no-carrier-added [¹⁸F]F⁻ was obtained in 500 μL of ¹⁸O-enriched water by proton irradiation. **L1** (200 μg) was suspended in 20 μL of DMF-HCl-pyridazine buffer (pH 2.0) and mixed with 20 μL of [¹⁸F]F⁻ (ca. 145 ± 40 MBq). The reaction mixture was heated at 85 °C for 45 min. Afterwards, the crude was quenched with 2 mL of 5% NH₄OH solution and purified by preconditioned C18 cartridge (Sep-Pak® Light, Waters) to selectively retain ¹⁸F-L1. The desired product was eluted with 1 mL of EtOH, evaporated at 80 °C under N₂ flow and re-suspended in 1 mL of EtOH/saline (1:9) for subsequent in vivo injections. Quality control of ¹⁸F-L1 was performed via analytical radio-HPLC (tr = 7.5 min, Figure S21, chromatogram a). Total synthesis time: 1 h and 15 min.

Formation of ¹⁸F-C1: Prepared following the procedure described above for non-labelled **C1**, with minor modifications. To a vial containing evaporated ¹⁸F-L1, 40 μL of Pd(NO₃)₂·2H₂O (1 mg/mL, 0.15 μmol, 1 equiv.) were added and the mixture was stirred for 30 min at room temperature. Quality control was performed via analytical radio-HPLC (tr = 8.6 min, Figure S21, chromatogram b). Then, the resulting solution was diluted with 1 mL of saline for in vivo injections.

Encapsulation of cisplatin in ¹⁸F-C1: Prepared following the procedure described above. To a vial containing evaporated ¹⁸F-L1, 40 μL of Pd(NO₃)₂·2H₂O in DMSO (1 mg/mL, 0.15 μmol, 1 equiv.) were added and the mixture was stirred for 30 min at room temperature. The resulting ¹⁸F-C1 was added to a pre-loaded Eppendorf containing 800 μL of cisplatin in ultrapure water (62.5 μg/mL, 2 equiv.). The reaction mixture was incubated during 5 min. Quality control was performed via analytical radio-HPLC (tr = 8.6 min, Figure S21, chromatogram c). Afterwards, 160 μL of saline were added to reach a final volume of 1 mL for in vivo injections.

In vivo and ex vivo biodistribution studies

Animals: Female mice (BALB/cJrj, 8 weeks, Janvier; 9 animals) weighing 22 ± 2 g were used to conduct the biodistribution studies. The animals were maintained and handled in accordance with the Guidelines for Accommodation and Care of Animals (European Convention for the Protection of Vertebrate Animals Used for Experimental and Other Scientific Purposes) and internal guidelines. All experimental procedures were approved by the internal committee and the local authorities.

Biodistribution studies: Mice were anesthetized by inhalation of 3% isoflurane in pure O₂ and maintained by 1.5–2% isoflurane in 100% O₂. With the animal under anesthesia, ¹⁸F-L1, ¹⁸F-C1 or cisplatin loaded ¹⁸F-C1 were injected intravenously via one of the lateral tail's veins (110 μL, 1.48 ± 0.74 MBq, n = 3 per compound). Dynamic whole body 60-min PET scans were started immediately after administration using MOLECUBES β-CUBE (PET) scanner. After each PET scan, whole-body high-resolution CT acquisitions were performed on the MOLECUBES X-CUBE (CT) scanner to provide anatomical information of each animal as well as the attenuation map for later image reconstruction. Random and scatter corrections were automatically applied during image reconstruction (3D OSEM reconstruction algorithm). PET-CT images of the same mouse were co-registered and analyzed using the PMOD image processing tool. Volumes of interest (VOIs) were manually delineated on selected organs (brain, heart, lungs, liver, kidneys, and bladder). Time-activity curves (decay corrected) were obtained as cps/cm³ in each organ. Curves were transformed into real activity (Bq/cm³), and finally injected dose normalization was applied to express the results as percentage of injected dose per cm³ of tissue (% ID/cm³).

Ex vivo studies: After the imaging session, animals were sacrificed, organs of interest were collected and weighed, and the radioactivity was measured in a gamma-counter (Wallach Wizard, PerkinElmer, Waltham, MA, USA). The uptake was calculated as a percentage of the injected dose per gram of tissue (% ID/g). Then, the weighted organs were immersed in digest solution of HNO₃/HCl (4:1, 5 mL) and heated to boiling until complete dissolution. The solution was subsequently analyzed by ICP-MS to determine the concentration of Pd in each sample.

Statistical analysis: Differences in concentration of radioactivity in each organ and time points were analyzed using 2-way ANOVA with Tukey's multiple comparison test. Differences were concluded significant for P values < 0.05; P < 0.05, *; P < 0.01, **; P < 0.001, ***; and P < 0.0001, ****. Statistical tests were performed in GraphPad Prism 7.03 (GraphPad Software, CA, USA).

Acknowledgements

This work was funded by MICINN (PID2020-113661GB-I00) and AGAUR (2017 SGR 01051 and 2017 SGR 00294). Mr A. Lliberia and Mss M. Artigues are acknowledged for recording the HR-

ESI-MS data for the cage-cisplatin complexes. Support from TUM Innovation Network “Artificial Intelligence Powered Multifunctional Materials Design” (ARTEMIS) is gratefully acknowledged. Part of the work was supported by MCIN/AEI/10.13039/501100011033 (PID2020-117656RB-I00). Open Access funding enabled and organized by Projekt DEAL.

Conflict of Interest

The authors declare no conflict of interest.

Data Availability Statement

The data that support the findings of this study are available from the corresponding author upon reasonable request.

Keywords: ammonium trifluoroborates · cisplatin · drug delivery · metallacages · PET imaging · self-assembly

- [1] a) K. Lu, T. Aung, N. Guo, R. Weichselbaum, W. Lin, *Adv. Mater.* **2018**, *30*, 1707634–1707654; b) S. F. M. Van Dongen, S. Cantekin, J. A. A. W. Elemans, A. E. Rowan, R. J. M. Nolte, *Chem. Soc. Rev.* **2014**, *43*, 99–122; c) T. R. Cook, Y.-R. Zheng, P. J. Stang, *Chem. Rev.* **2013**, *113*, 734–777; d) S. V. Dummert, K. Yadava, H. Saini, M. Z. Hussain, K. Jayaramulu, A. Casini, R. A. Fischer, *Chem. Soc. Rev.* **2022**, *51*, 5175–5213; e) F. d’Orchymont, J. P. Holland, *Angew. Chem. Int. Ed.* **2022**, DOI: 10.1002/anie.202204072.
- [2] a) H. Amouri, C. Desmarests, J. Moussa, *Chem. Rev.* **2012**, *112*, 2015–2041; b) M. Han, D. M. Engelhard, G. H. Clever, *Chem. Soc. Rev.* **2014**, *43*, 1848–1860; c) M. D. Ward, C. A. Hunter, N. H. Williams, *Acc. Chem. Res.* **2018**, *51*, 2073–2082; d) C. J. Brown, F. D. Toste, R. G. Bergman, K. N. Raymond, *Chem. Rev.* **2015**, *115*(9), 3012–3035; e) E. Benchimol, B.-N. T. Nguyen, T. K. Ronson, J. R. Nitschke, *Chem. Soc. Rev.* **2022**, *51*, 5101–5135; f) Y. Fang, J. A. Powell, E. Li, Q. Wang, Z. Perry, A. Kirchon, X. Yang, Z. Xiao, C. Zhu, L. Zhang, F. Huang, H.-C. Zhou, *Chem. Soc. Rev.* **2019**, *48*, 4707–4730; g) Y. Sun, C. Chen, J. Liu, P. J. Stang, *Chem. Soc. Rev.* **2020**, *49*, 3889–3919; h) T. Keijer, T. Bouwens, J. Hessels, J. N. H. Reek, *Chem. Sci.* **2021**, *12*, 50–70; i) J. Zhao, Z. Zhou, G. Li, P. J. Stang, X. Yan, *Natl. Sci. Rev.* **2021**, *8*, nwab045; j) N. Dey, C. J. E. Haynes, *ChemPlusChem* **2021**, *86*, 418; k) S. Pullen, G. H. Clever, *Acc. Chem. Res.* **2018**, *51*, 3052–3064; l) X. Jing, C. He, L. Zhao, C. Duan, *Acc. Chem. Res.* **2019**, *52*(1), 100–109.
- [3] a) H. Seppehrpour, W. Fu, Y. Sun, P. J. Stang, *J. Am. Chem. Soc.* **2019**, *141*, 14005–14020; b) B. Therrien, G. Suess-Fink, P. Govindaswamy, A. K. Renfrew, P. J. Dyson, *Angew. Chem. Int. Ed.* **2008**, *47*, 3773–3776; *Angew. Chem.* **2008**, *120*, 3833–3836; c) J. E. M. Lewis, E. L. Gavey, S. A. Cameron, J. D. Crowley, *Chem. Sci.* **2012**, *3*, 778–784; d) Y.-R. Zheng, K. Suntharalingam, T. C. Johnstone, S. J. Lippard, *Chem. Sci.* **2015**, *6*, 1189–1193; e) W.-Q. Xu, Y.-Z. Fan, H.-P. Wang, J. Teng, Y.-H. Li, C.-X. Chen, D. Fenske, J.-J. Jiang, C.-Y. Su, *Chem. Eur. J.* **2017**, *23*, 3542–3547; f) F. Schmitt, J. Freudenreich, N. P. E. Barry, L. Juillerat-Jeanneret, G. Süß-Fink, B. Therrien, *J. Am. Chem. Soc.* **2012**, *134*, 754–757; g) G. Yu, B. Zhu, L. Shao, J. Zhou, M. L. Saha, B. Shi, Z. Zhang, T. Hong, S. Li, X. Chen, P. J. Stang, *Proc. Natl. Acad. Sci. USA* **2019**, *116*, 6618–6623; h) Z. Zhou, J. Liu, J. Huang, T. W. Rees, Y. Wang, H. Wang, X. Li, H. Chao, P. J. Stang, *Proc. Natl. Acad. Sci. USA* **2019**, *116*, 20296–20302.
- [4] a) B. P. Burke, W. Grantham, M. J. Burke, G. S. Nichol, D. Roberts, I. Renard, R. Hargreaves, C. Cawthorne, S. J. Archibald, P. J. Lusby, *J. Am. Chem. Soc.* **2018**, *140*, 16877–16881; b) H. Zhu, Q. Li, B. Shi, F. Ge, Y. Liu, Z. Mao, H. Zhu, S. Wang, G. Yu, F. Huang, P. J. Stang, *Angew. Chem. Int. Ed.* **2020**, *59*, 20208–20214; *Angew. Chem.* **2020**, *132*, 20383–20389.
- [5] a) J. Xu, J. Wang, J. Ye, J. Jiao, Z. Liu, C. Zhao, B. Li, Y. Fu, *Adv. Sci.* **2021**, *8*, 2101101; b) G. Yua, T. R. Cook, Y. Lic, X. Yand, D. Wuc, L. Shaoa, J. Shenc, G. Tangc, F. Huang, X. Chene, P. J. Stang, *Proc. Natl. Acad. Sci.* **2016**, *113*, 13720–13725; c) G. Yu, S. Yu, M. L. Saha, J. Zhou, T. R. Cook, B. C. Yung, J. Chen, Z. Mao, F. Zhang, Z. Zhou, Y. Liu, L. Shao, S. Wang, C. Gao, F. Huang, P. J. Stang, X. Chen, *Nat. Commun.* **2018**, *9*, 1–18.
- [6] a) A. Schmidt, V. Molano, M. Hollering, A. Pöthig, A. Casini, F. E. Kühn, *Chem. A Eur. J.* **2016**, *22*, 2253–2256; b) B. Woods, M. N. Wenzel, T. Williams, S. R. Thomas, R. L. Jenkins, A. Casini, *Front. Chem.* **2019**, *7*, 68.
- [7] A. Schmidt, M. Hollering, M. Drees, A. Casini, F. E. Kühn, *Dalton Trans.* **2016**, *45*, 8556.
- [8] A. Schmidt, M. Hollering, J. Han, A. Casini, F. E. Kühn, *Dalton Trans.* **2016**, *45*, 12297.
- [9] a) B. Woods, D. Döllerer, B. Aikman, M. N. Wenzel, E. Sayers, F. E. Kühn, A. Jones, A. Casini, *J. Inorg. Biochem.* **2019**, *199*, 110781; b) B. Aikman, R. Bonsignore, B. Woods, D. Doellerer, R. Scotti, C. Schmidt, A. A. Heidecker, A. Pöthig, E. J. Sayers, A. T. Jones, A. Casini, *Dalton Trans.* **2022**, DOI: 10.1039/d2dt00337f.
- [10] J. Han, A. Schmidt, T. Zhang, H. Permentier, G. M. M. Groothuis, R. Bischoff, F. E. Kühn, P. Horvatovich, A. Casini, *Chem. Commun.* **2017**, *53*, 1405–1408.
- [11] J. Han, A. F. B. Räder, F. Reichart, B. Aikman, M. N. Wenzel, B. Woods, M. Weinmüller, B. S. Ludwig, S. Stürup, G. M. M. Groothuis, H. P. Permentier, R. Bischoff, H. Kessler, P. Horvatovich, A. Casini, *Bioconjugate Chem.* **2018**, *29*, 3856–3865.
- [12] B. Woods, R. D. M. Silva, C. Schmidt, D. Wragg, M. Cavaco, V. Neves, V. F. C. Ferreira, L. Gano, T. S. Morais, F. Mendes, J. D. G. Correia, A. Casini, *Bioconjugate Chem.* **2021**, *32*, 1399–1408.
- [13] a) A. Casini, B. Woods, M. Wenzel, *Inorg. Chem.* **2017**, *56*, 14715–14729; b) A. Pöthig, A. Casini, *Theranostics* **2019**, *9*, 3150–3169.
- [14] a) Z. Liu, M. Pourghiasian, M. A. Radtke, J. Lau, J. Pan, G. M. Dias, D. Yapp, K.-S. Lin, F. Bénard, D. M. Perrin, *Angew. Chem. Int. Ed.* **2014**, *53*, 11876–11880; *Angew. Chem.* **2014**, *126*, 12070–12074; b) D. M. Perrin, *Curr. Opin. Chem. Biol.* **2018**, *45*, 86–94; c) A. Roxin, C. Zhang, S. Huh, M. Lepage, Z. Zhang, K.-S. Lin, F. Bénard, D. M. Perrin, *Bioconjugate Chem.* **2019**, *30*, 1210–1219.
- [15] M. E. Bakkari, J.-M. Vincent, *Org. Lett.* **2004**, *6*, 2765–2767.
- [16] H. Pohlit, M. Worm, J. Langhanki, E. Berger-Nicoletti, T. Opatz, H. Frey, *Macromolecules* **2017**, *50*, 9196–9206.
- [17] D. H. Leung, R. G. Bergman, K. N. Raymond, *J. Am. Chem. Soc.* **2008**, *130*, 2798–2805.
- [18] D. Preston, S. M. McNeill, J. E. M. Lewis, G. I. Giles, J. D. Crowley, *Dalton Trans.* **2016**, *45*, 8050–8060.
- [19] a) E. Puig, C. Desmarests, G. Gontard, M. N. Rager, A. L. Cooksy, H. Amouri, *Inorg. Chem.* **2019**, *58*, 3189–3195; b) F. Kaiser, A. Schmidt, W. Heydenreuter, P. J. Altmann, A. Casini, S. A. Sieber, F. E. Kühn, *Eur. J. Inorg. Chem.* **2016**, *33*, 5189–5196.
- [20] J. B. Williamson, E. R. Smith, J. R. Scheerer, *Synlett.* **2017**, *28*, 1170–1172.

Manuscript received: August 20, 2022

Accepted manuscript online: October 14, 2022

Version of record online: November 23, 2022