



# Impact of processing on important cocoa off-flavour compounds

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## Abstract

The compounds responsible for smoky and mouldy–musty off-flavours in fermented cocoa have recently been elucidated; however, their behaviour during further processing into chocolate was still unclear. The compounds 2-methoxyphenol, 3-methylphenol, 4-methylphenol, 3-ethylphenol, 4-ethylphenol, and 3-propylphenol known to contribute to smoky off-flavours showed a tendency towards a minor increase during roasting and processing into cocoa liquor. This increase amounted to 1.4-fold at the most, however, was clearly compensated by losses of 30–63% during further processing into chocolate mass and conching. Among the off-flavour compounds identified in mouldy–musty smelling cocoa, faecal, mothball-like 3-methyl-1*H*-indole showed a clear decrease during roasting and processing into cocoa liquor, at least at rather high roasting temperatures, and a further decrease during processing into chocolate mass and conching. In contrast, faecal, mothball-like 1*H*-indole substantially increased during roasting and processing into cocoa liquor, namely from concentrations below its odour threshold value to concentrations up to 8 times beyond its odour threshold value. During processing into chocolate mass and conching, 1*H*-indole remained virtually unchanged. The data suggested that the monitoring of off-flavour compounds at the incoming goods inspection in the chocolate industry should not be limited to the fermented beans as such but additionally include the analysis of a bean sample after test roasting to correctly assess the off-flavour potential of 3-methyl-1*H*-indole and 1*H*-indole.

**Keywords** Cocoa · *Theobroma cacao* · Smoky off-flavour · Mouldy–musty off-flavour · Roasting · Conching

## Abbreviations

GC–MS	Gas chromatography–mass spectrometry
Geosmin	(4 <i>S</i> ,4 <i>A</i> <i>S</i> ,8 <i>a</i> <i>R</i> )-4,8 <i>a</i> -Dimethyloctahydronaphthalen-4 <i>a</i> (2 <i>H</i> )-ol
MDMF	4-Methoxy-2,5-dimethylfuran-3(2 <i>H</i> )-one
OAV	Odour activity value
SAFE	Solvent-assisted flavour evaporation

## Introduction

To make chocolate, the fermented seeds of the cocoa tree (*Theobroma cacao* L.) are roasted and then ground into cocoa liquor. After addition of further ingredients such as sugar, cocoa butter, and milk powder, the mixture is ground to yield chocolate mass. Further homogenisation

and refining is achieved by conching, a process that lasts between hours and days, before the mass is finally moulded into chocolate [1, 2].

Fermented cocoa is occasionally tainted with off-flavours. Such batches must be sorted out during incoming goods inspection in the chocolate industry to avoid quality issues with the final product. In two recent studies, we identified the crucial odorants responsible for the smoky off-flavour and the crucial odorants responsible for the mouldy–musty off-flavour in fermented cocoa. We showed that the smoky off-note is caused by 2-methoxyphenol, 3-methylphenol, 4-methylphenol, 3-ethylphenol, 4-ethylphenol, and 3-propylphenol [3], whereas (–)-geosmin, 3-methyl-1*H*-indole, 1*H*-indole, and 4-methoxy-2,5-dimethylfuran-3(2*H*)-one (MDMF) are potential off-flavour compounds in mouldy–musty smelling cocoa samples [4]. Predominantly based on the odour threshold values in deodorized cocoa butter, maximum tolerable concentrations in fermented cocoa were suggested. These limits amounted to 1.1 µg/kg for 3-methyl-1*H*-indole, 1.6 µg/kg for (–)-geosmin, 2 µg/kg for 3-ethylphenol and 3-propylphenol, 20 µg/kg for 3-methylphenol, 4-methylphenol, and 4-ethylphenol,

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and 70 µg/kg for 2-methoxyphenol [3, 4]. The decision on acceptance or rejection can thus be made on a more objective basis than with sensory testing only. However, it was so far largely unclear, how further processing influences the concentrations of the off-flavour compounds. Given the elevated temperatures, a substantial impact could be expected from roasting and conching. Cocoa bean roasting is typically done at 120–140 °C for 20–30 min [2, 5, 6], whereas during conching, temperatures of ~50–80 °C are applied for up to 3 days [1, 2, 7].

Several studies have addressed the changes in odorant concentrations during cocoa processing [6, 8–15]. It was shown that the high temperatures during roasting converted thermolabile precursors formed during fermentation [16, 17] into important odour-active compounds such as Strecker aldehydes, pyrazines, and furanones [8–10, 18]. At the same time, the concentrations of some undesired odorants such as acetic acid were reduced [6, 11]. Conching not only improved the rheological properties of the chocolate but also affected chocolate flavour [14, 19]. Specifically, the concentrations of some undesired volatile acids decreased [14], whereas for example caramel-like smelling 4-hydroxy-2,5-dimethylfuran-3(2*H*)-one (HDMF; Furaneol®) increased during conching [14, 15]. So far, no study on the effect of cocoa processing focused on off-flavour compounds. Nonetheless, Frauendorfer et al. reported higher concentrations of 2-methoxyphenol and 4-methylphenol after roasting of Criollo and Forastero cocoa beans [9, 10]. Moreover, Counet et al. who quantitated 44 volatiles in dark chocolate before and after conching found that 2-methoxyphenol and 1*H*-indole increased [14]. In contrast, Beckett reported a reduction in the amount of phenols during conching; however, no data on individual compounds was provided [20].

To obtain a better view on the behaviour of the compounds contributing to smoky and mouldy–musty off-flavours during chocolate manufacturing, the aim of the current study was to determine their concentration changes induced by 1) cocoa roasting and processing into cocoa liquor and by 2) further processing into chocolate mass and conching. If substantial differences were to occur, the previously suggested maximum tolerable concentrations in fermented cocoa might need to be adjusted.

## Materials and methods

### Cocoa

The samples of fermented cocoa beans were obtained from German chocolate manufacturers. The samples SOF1 and SOF2 showed smoky off-flavours, the samples MOF1 and MOF2 were tainted with mouldy–musty off-flavours, and

the reference sample REF exhibited a characteristic aroma without off-notes. All samples were stored at +4 °C.

### Chemicals

The reference odorants 2-methoxyphenol, 4-methylphenol, 3-methylphenol, 4-ethylphenol, 3-ethylphenol, 3-propylphenol, MDMP, 3-methyl-1*H*-indole, 1*H*-indole, and geosmin as well as 4-(<sup>2</sup>H<sub>3</sub>)methyl(<sup>2</sup>H<sub>4</sub>)phenol were purchased from Merck (Darmstadt, Germany). The following isotopically substituted odorants were synthesised as detailed in the literature: 2-[(<sup>2</sup>H<sub>3</sub>)methoxy]phenol [21], 4-(1,1-<sup>2</sup>H<sub>2</sub>)ethylphenol [3], 4-(1,1-<sup>2</sup>H<sub>2</sub>)propylphenol [3], 2,5-dimethyl-4-[(<sup>2</sup>H<sub>3</sub>)methoxy]furan-3(2*H*)-one [22], 3-(<sup>2</sup>H<sub>3</sub>)methyl(2,4,5,6,7-<sup>2</sup>H<sub>5</sub>)-1*H*-indole [23], and ((4*S*,4*aS*,8*aR*)-4,8*a*-dimethyl(3,3,4-<sup>2</sup>H<sub>3</sub>)octahydronaphthalen-4*a*(2*H*)-ol) [24]. (2,3,4,5,6,7-<sup>2</sup>H<sub>6</sub>)-1*H*-Indole was synthesised from the isotopically unmodified compound 1*H*-indole according to the approach published for the synthesis of 3-(<sup>2</sup>H<sub>3</sub>)methyl(2,4,5,6,7-<sup>2</sup>H<sub>5</sub>)-1*H*-indole [23]. Dichloromethane was freshly distilled through a column (120 cm × 5 cm) packed with Raschig rings before use. Ethanol (99.9%) was purchased from Honeywell (Seelze, Germany).

### Roasting and processing into cocoa liquor

The fermented cocoa beans were roasted according to the protocol “Elements of harmonized international standards for cocoa quality and flavour assessment” of the Cocoa of Excellence (CoEx) Programme [25]. In brief, the fermented cocoa beans were placed in a single layer on a stainless steel mesh tray and put in a pre-heated convection oven FP53 (Binder, Tuttlingen, Germany) for 25 min. The roasting temperatures were 110, 125, and 140 °C. After roasting, the cocoa beans were allowed to cool down to room temperature before the shells were removed manually. The cocoa was processed into cocoa liquor using a preheated (45 °C) RM 200 mortar grinder (Retsch, Haan, Germany) equipped with a porcelain pestle and a porcelain mortar. The cocoa liquors were stored at +4 °C until analysis.

### Processing into chocolate mass and conching

Solutions of the off-flavour compounds (0.2–9 mg in 200 µl ethanol) were premixed with molten cocoa liquor (500 g). In the pilot plant of a German chocolate manufacturer, the spiked cocoa liquors were included into the recipes used for making a dark chocolate mass (10 kg; >50% cocoa liquor; further ingredients: cocoa butter, sugar) and a milk chocolate mass (10 kg; <10% cocoa liquor; further ingredients: cocoa butter, milk powder, sugar) and conched at a temperature of 80 °C. After 12 h, the mass was moulded into chocolate.

## Odorant quantitation

Samples of the cocoa liquors and chocolates were immersed in liquid nitrogen, crushed using a GM 200 laboratory mill (Retsch) and further ground into a fine powder using a 6875 Freezer Mill (Spex SamplePrep, Stanmore, UK). To portions (1–50 g) of the powder, dichloromethane (25–100 mL) was added and the mixture was spiked with the isotopically substituted odorants (0.04–8 µg) in dichloromethane (40–400 µL) as internal standards. After magnetic stirring at room temperature for ~ 15 h, the mixture was filtered through a folded paper filter. The filtrate was dried over anhydrous sodium sulphate. Non-volatiles were removed by SAFE at 40 °C [26]. The distillate was concentrated using a Vigreux column (50 × 1 cm) and a Bemelmans microdistillation device [27] to a final volume of 200 µL. Portions of the concentrates (1–2 µL) were analysed by GC–MS. Quantitation of 2-methoxyphenol, 3- and 4-methylphenol, 3- and 4-ethylphenol, 3-methyl-1*H*-indole, 1*H*-indole, and MDMF was accomplished using a heart-cut GC–GC–MS system. For the quantitation of 3-propylphenol and (–)-geosmin, a heart-cut GC–GC–HRMS system was employed. Odorant concentrations were finally calculated from the peak area counts of the analyte peak and the internal standard peak in the extracted ion chromatograms of characteristic quantifier ions, the amount of sample used, and the amount of standard added, by employing a calibration line equation obtained by linear regression after the analysis of analyte/standard mixtures in at least five different concentration ratios, covering a range of at least 1:5 to 5:1. Details on the GC–MS systems, the quantifier ions, and the calibrations are provided in the Supplementary Information file.

## Results and discussion

### Impact of roasting and processing into cocoa liquor on cocoa off-flavour compounds

Two samples of fermented cocoa beans with a smoky off-flavour, two samples of fermented cocoa beans with a mouldy–musty off-flavour, and a reference sample with a characteristic aroma and no off-flavour were roasted and then processed into cocoa liquor. Each of the five cocoa samples was roasted at three different temperatures, namely 110, 125, and 140 °C. Although roasting temperatures as low as 90 °C and as high as 170 °C have also been reported in the literature [1, 13, 28, 29], the range of 110–140 °C is the one most relevant in the chocolate industry [2, 6].

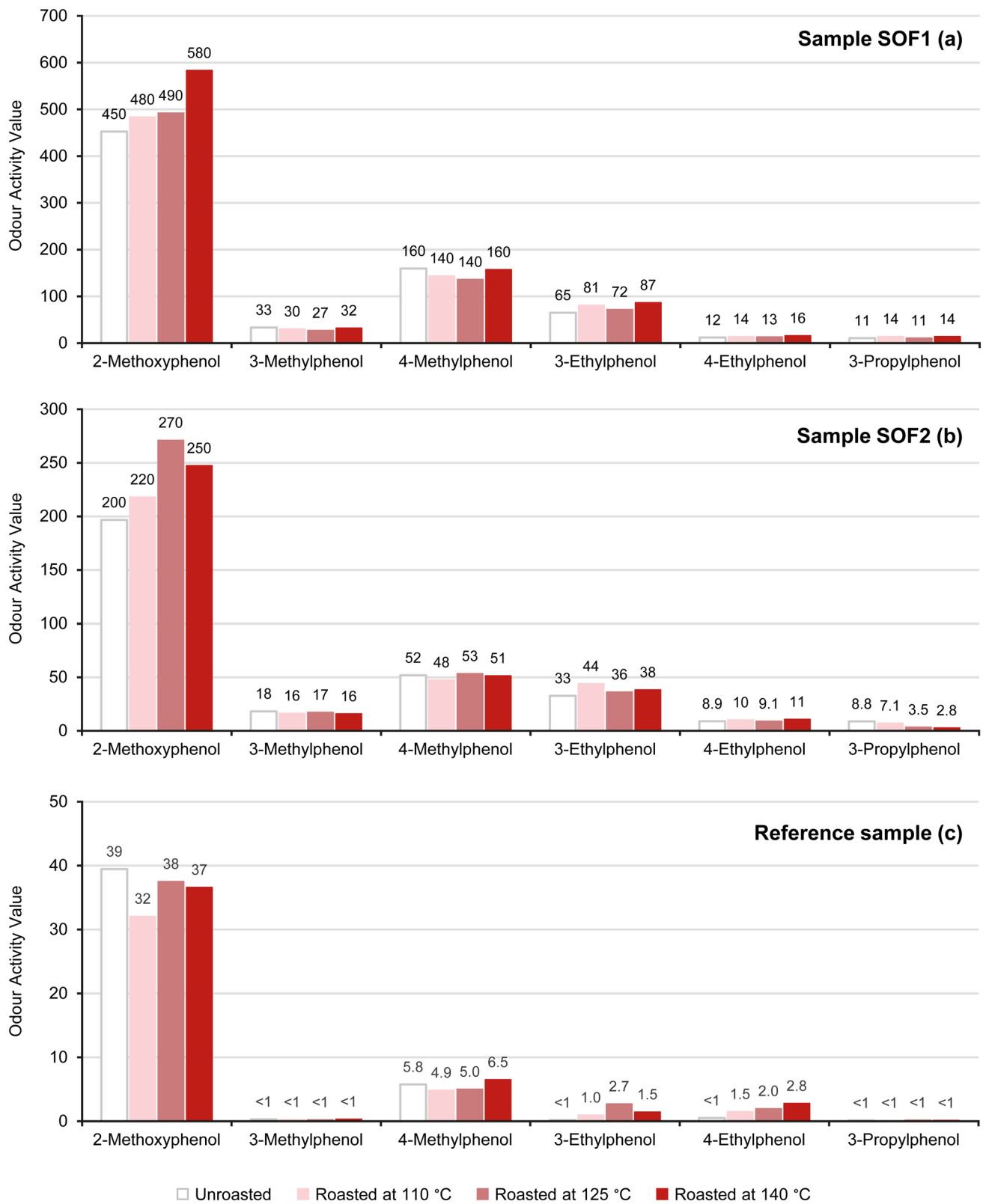
In the cocoa liquors obtained from the cocoa samples with a smoky off-flavour (SOF1, SOF2), the six phenols previously identified as causative for the off-note [3] were quantitated. In detail, the compounds and their odour qualities were 2-methoxyphenol (smoky, hammy), 3-methylphenol (smoky, phenolic), 4-methylphenol (horse stable-like, phenolic), 3-ethylphenol (smoky), 4-ethylphenol (smoky), and 3-propylphenol (smoky, phenolic). Quantitation was accomplished by GC–MS after solvent extraction and SAFE using stable isotopically substituted odorants as internal standards. The reference sample without off-flavour (REF) was analysed in parallel. The data obtained were compared to the concentrations before processing (Table 1). Moreover, the concentrations were divided by the odour threshold values of the compounds in deodorized cocoa butter to obtain odour activity values (OAVs) and thus to simultaneously assess the impact of processing and the odour relevance of the compounds (Fig. 1). The odour threshold values applied were 1.8 µg/kg for 2-methoxyphenol, 19 µg/kg for 3-methylphenol, 3.3 µg/kg for 4-methylphenol, 2.2 µg/kg for

**Table 1** Changes in the concentrations (µg/kg) of crucial off-flavour compounds in two cocoa samples with a smoky off-flavour (SOF1, SOF2) and a reference sample without off-flavour caused by roasting at 110, 125, and 140 °C and processing into cocoa liquor

Odorant	SOF1				SOF2				Reference sample			
	Before <sup>a</sup>	110 °C <sup>b</sup>	125 °C <sup>b</sup>	140 °C <sup>b</sup>	Before <sup>a</sup>	110 °C <sup>b</sup>	125 °C <sup>b</sup>	140 °C <sup>b</sup>	Before <sup>a</sup>	110 °C <sup>b</sup>	125 °C <sup>b</sup>	140 °C <sup>b</sup>
2-Methoxyphenol	815	871	886	1050	354	393	488	445	71.0	57.7	67.5	65.9
3-Methylphenol	635	575	517	611	346	312	329	302	5.82	4.97	4.39	6.45
4-Methylphenol	527	476	450	519	171	157	176	170	19.1	16.0	16.6	21.5
3-Ethylphenol	143	179	158	191	71.9	96.7	79.8	84.2	0.379	2.17	5.98	3.21
4-Ethylphenol	275	320	308	368	205	236	209	249	11.9	35.1	45.7	64.5
3-Propylphenol	21.3	28.4	21.9	28.1	17.5	14.3	6.93	5.56	0.255	0.264	0.347	0.292

<sup>a</sup>Data taken from [3]

<sup>b</sup>Mean of duplicate or triplicate workups; individual concentrations and standard deviations are available in the Supplementary Information file, Tables S2–S4



**Fig. 1** Changes in the odour activity values of crucial off-flavour compounds in two cocoa samples with a smoky off-flavour (SOF1, SOF2) and a reference sample without off-flavour caused by roasting at 110, 125, and 140 °C and processing into cocoa liquor

3-ethylphenol, 23 µg/kg for 4-ethylphenol, and 2.0 µg/kg for 3-propylphenol [3].

Results indicated only minor changes of the compounds during roasting and processing into cocoa liquor. In particular, there was no general decrease of off-flavour compounds that could have been expected if—despite the rather high boiling points of the compounds of ~202–230 °C—evaporation during roasting had played a substantial role. In this case, an adjustment of the maximum tolerable concentrations in fermented cocoa during incoming goods inspection in the chocolate industry could have been considered. A clear decrease that reflected the roasting temperatures was only observed for a single compound in a single sample, namely 3-propylphenol in sample SOF2. The compound decreased during roasting and processing into cocoa liquor from 17.5 to 14.3 µg/kg at 110 °C, 6.93 µg/kg at 125 °C, and 5.56 µg/kg at 140 °C (Table 1) which corresponded to OAVs of 8.8 before roasting and 7.1, 3.5, and 2.8 after roasting and processing into cocoa liquor (Fig. 1). However, no such behaviour was observed for 3-propylphenol in sample SOF1. By contrast, some of the off-flavour compounds in the off-flavour samples rather showed a tendency towards a slight increase. This increase was clearly beyond the concentration effect associated with the loss of water during roasting. Given a typical water content of 4–8% in the unroasted cocoa and a final water content of ~2% in the roasted cocoa [1, 30, 31], such an increase would be clearly below the 1.1-fold. On the other hand, the increase was also never greater than 1.4-fold and, in particular, did not change the order of the off-flavour compounds in terms of OAVs: before and after roasting and processing into cocoa liquor, 2-methoxyphenol showed the highest OAVs among the six compounds in the two off-flavour samples followed by 4-methylphenol and 3-ethylphenol. In the reference sample without off-flavour, the levels of all off-flavour compounds were clearly lower, before roasting as well as after roasting and processing into cocoa liquor. 2-Methoxyphenol and 4-methylphenol were the only compounds exceeding their odour threshold levels before roasting. During roasting and

processing into cocoa liquor, both did not show substantial changes; however, the concentrations of 3-ethylphenol and 4-ethylphenol increased up to 15-fold for 3-ethylphenol and up to 5.4-fold for 4-ethylphenol, resulting in concentrations slightly beyond their threshold values, whereas 3-methylphenol concentrations and 3-propylphenol concentrations stayed below the threshold value.

Different from our results, Frauendorfer and Schieberle reported a clear increase of 2-methoxyphenol and 4-methylphenol during roasting of cocoa beans without an off-flavour [9, 10], although conditions with 95 °C roasting temperature and 14 min roasting time were rather mild. For 2-methoxyphenol, they found an increase from 110 µg/kg (Criollo beans) and 61 µg/kg (Forastero beans) to 230 and 100 µg/kg, respectively, whereas in our reference sample, the concentration even slightly decreased from 71 to 57.7–67.5 µg/kg, depending on the roasting temperature. For 4-methylphenol, Frauendorfer and Schieberle observed an increase from 5.3 (Criollo) and 4.6 (Forastero) to 9.9 and 7.6 µg/kg, respectively. In our reference sample, the concentration before roasting was 19.1 µg/kg and after roasting and processing into cocoa liquor it ranged from 16.0 to 21.5 µg/kg.

In the cocoa liquors obtained from the cocoa samples with a mouldy–musty off-flavour (MOF1, MOF2), (–)-geosmin, 3-methyl-1*H*-indole, 1*H*-indole, and 4-methoxy-2,5-dimethylfuran-3(2*H*)-one (MDMF) were quantitated. This selection was based on our previous study [4]. Again, the reference sample without off-flavour (REF) was analysed in parallel. Results are depicted in Table 2 together with the concentrations before processing. Figure 2 shows the OAVs calculated from the concentrations and the odour threshold values of the compounds in deodorized cocoa butter. The odour threshold values used for these calculations were 1.6 µg/kg for (–)-geosmin, 1.1 µg/kg for 3-methyl-1*H*-indole, 51 µg/kg for 1*H*-indole, and 350 µg/kg for MDMF [4].

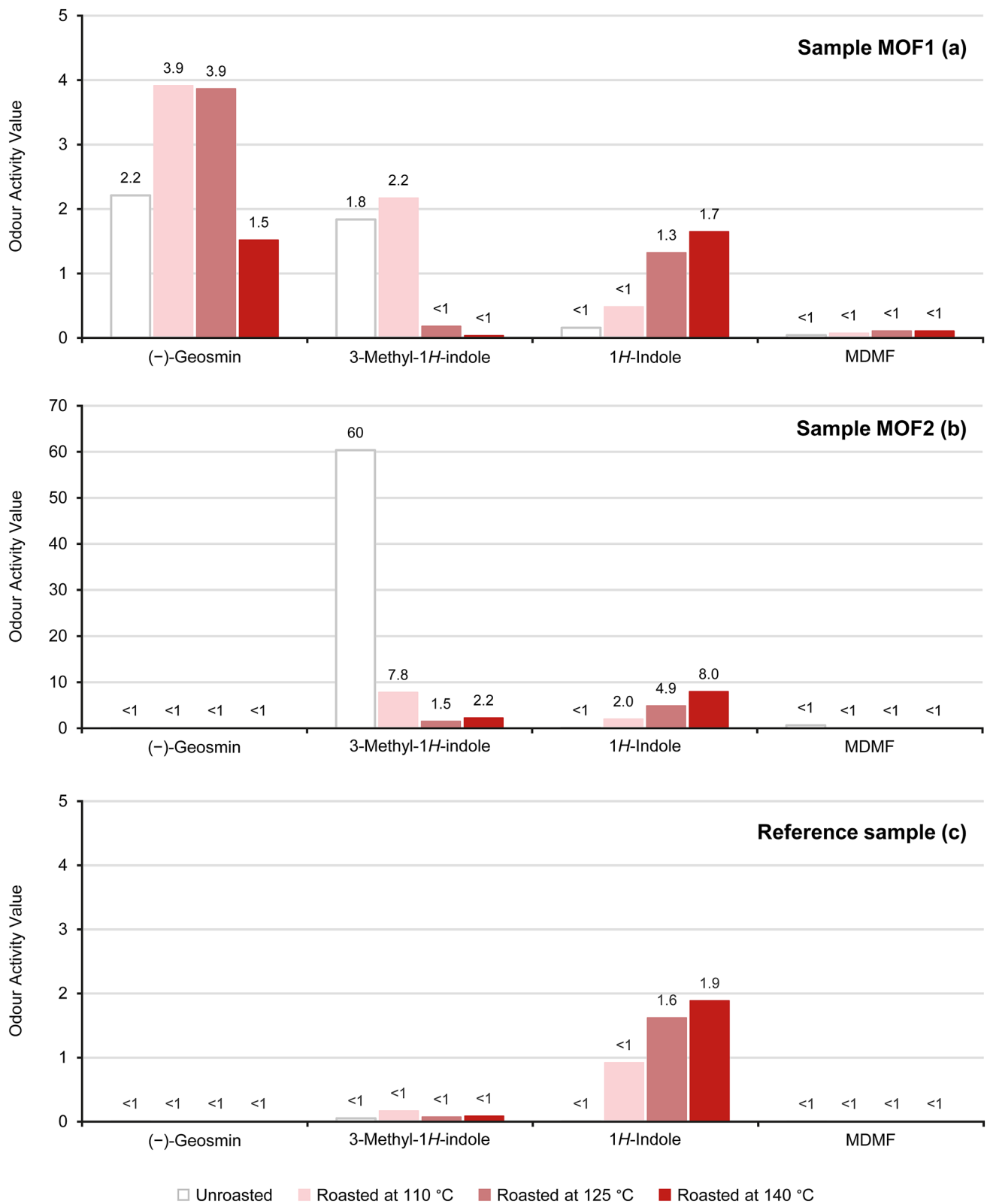
(–)-Geosmin, the compound with the most pronounced mouldy smell, was present in odour-active amounts only in sample MOF1. Its concentration in the samples roasted at 110 and 125 °C was almost twice the concentration in

**Table 2** Changes in the concentrations (µg/kg) of crucial off-flavour compounds in two cocoa samples with a mouldy–musty off-flavour (MOF1, MOF2) and a reference sample without off-flavour caused by roasting at 110, 125, and 140 °C and processing into cocoa liquor

Odorant	MOF1				MOF2				Reference sample			
	Before <sup>a</sup>	110 °C <sup>b</sup>	125 °C <sup>b</sup>	140 °C <sup>b</sup>	Before <sup>a</sup>	110 °C <sup>b</sup>	125 °C <sup>b</sup>	140 °C <sup>b</sup>	Before <sup>a</sup>	110 °C <sup>b</sup>	125 °C <sup>b</sup>	140 °C <sup>b</sup>
(–)-Geosmin	3.54	6.26	6.19	2.43	<0.2	<0.2	<0.2	<0.2	<0.2	<0.2	<0.2	<0.2
3-Methyl-1 <i>H</i> -indole	2.07	2.39	<1.0	<1.0	66.4	8.59	1.65	2.47	<1.0	<1.0	<1.0	<1.0
1 <i>H</i> -Indole	8.10	24.7	67.5	84.2	5.46	102	248	406	<1.0	47.0	82.7	96.2
MDMF	14.9	26.4	38.2	38.4	226	3.64	3.68	2.52	<1.0	1.32	1.45	1.62

<sup>a</sup>Data taken from [4]

<sup>b</sup>Mean of duplicate or triplicate workups; individual concentrations and standard deviations are available in the Supplementary Information file, Tables S5–S7



**Fig. 2** Changes in the odour activity values of crucial off-flavour compounds in two cocoa samples with a mouldy–musty off-flavour (MOF1, MOF2) and a reference sample without off-flavour caused by roasting at 110, 125, and 140 °C and processing into cocoa liquor



the unprocessed sample. This is hard to explain, given that the compound is a well-known product of microbiological metabolism but not known to be formed thermally [32–34]. Furthermore, (–)-geosmin has been found enriched in the cocoa seed shells [4]. The shells, however, had been removed during processing of the roasted cocoa samples to cocoa liquor. Thus, rather a lower concentration was to be expected in the roasted samples. This, however, was only the case for the sample roasted at 140 °C, in which not more than 2/3 of the initial (–)-geosmin was recovered.

Faecal, mothball-like smelling 3-methyl-1*H*-indole was the most odour-active compound in the unprocessed off-flavour sample MOF2 and the second most odour-active compound in the unprocessed off-flavour sample MOF1. In both off-flavour samples, its concentration clearly decreased during roasting and processing into cocoa liquor, at least when higher roasting temperatures were applied. After roasting at 125 and 140 °C and processing into cocoa liquor, its OAV in sample MOF1 had decreased from 1.8 to clearly below 1 and in sample MOF2, the decrease was from 60 to 1.5 and 2.2, respectively.

The most conclusive change during roasting and processing into cocoa liquor was observed for faecal, mothball-like smelling 1*H*-indole. In all three cocoa samples, the concentration in the unprocessed cocoa was below the odour threshold value of 51 µg/kg. In the processed samples, the amounts increased continuously with increasing roasting temperature leading to OAVs between 1.7 and 8.0 in the samples roasted at 140 °C. In the three 140 °C samples, 1*H*-indole was the most odour-active among the four off-flavour compounds. Moreover, in the reference sample 1*H*-indole was the only

compound of the four that exceeded the odour threshold, namely in the samples roasted at 125 and 140 °C.

Caramel-like and musty smelling MDMF showed a heterogeneous behaviour during roasting. Whereas in sample MOF1 and in the reference sample its concentration increased with the roasting temperature, a decrease was observed in sample MOF2. However, in none of the samples any concentration ever exceeded the odour threshold value of the compound.

### Impact of processing into chocolate mass and conching on cocoa off-flavour compounds

A cocoa liquor with a characteristic aroma and no off-flavour was spiked with the compounds previously identified as crucial for smoky and mouldy–musty off-flavours. The final concentrations resulting from the amounts naturally present in the cocoa liquor plus the spiked amounts were in the ranges previously determined in cocoa samples with the respective off-flavours [3, 4]. Portions of the spiked cocoa liquor were processed into a dark chocolate mass and into a milk chocolate mass, respectively. After conching at 80 °C for 12 h, the off-flavour compounds were quantitated and the concentrations obtained were compared to the concentrations in the cocoa liquor before processing.

The results (Table 3) revealed recoveries between 9 and 101%. For none of the compounds, a substantial difference between the dark chocolate mass and the milk chocolate mass was observed. The six phenolic compounds crucial for the smoky off-flavours showed recoveries of 37–70% in the dark chocolate mass and recoveries of 41–74% in the milk chocolate mass. The lowest recoveries were

**Table 3** Changes in the concentrations of crucial off-flavour compounds caused by processing spiked cocoa liquor into chocolate mass and conching

Odorant	Dark chocolate mass			Milk chocolate mass		
	Before (µg/kg) <sup>a</sup>	After (µg/kg) <sup>b</sup>	Recovery (%)	Before (µg/kg) <sup>a</sup>	After (µg/kg) <sup>b</sup>	Recovery (%)
2-Methoxyphenol	926	346	37	897	369	41
3-Methylphenol	362	202	56	360	226	63
4-Methylphenol	576	381	66	570	406	71
3-Ethylphenol	174	121	70	174	128	74
4-Ethylphenol	339	235	69	325	239	74
3-Propylphenol	21.1	10.7	51	21.0	11.5	55
(–)-Geosmin	5.10	4.02	79	5.10	4.16	82
3-Methyl-1 <i>H</i> -indole	106	83.9	79	106	84.4	80
1 <i>H</i> -Indole	83.5	84.3	101	28.4	26.4	93
MDMF	519	47.8	9	519	57.5	11

<sup>a</sup>Sum of the amount naturally present in the cocoa liquor and the spiked amount; details are available in the Supplementary Information file, Table S8

<sup>b</sup>Mean of triplicate workups; individual concentrations and standard deviations are available in the Supplementary Information file, Tables S9 and S10

obtained for 2-methoxyphenol. Interestingly, Counet et al. [14] reported higher concentrations for 2-methoxyphenol after conching. In two different chocolate samples, the 2-methoxyphenol concentrations increased by 50 and 13%, respectively. However, these experiments had been performed at much lower concentration levels (66 and 75 µg/kg) than our experiments (926 and 897 µg/kg), which probably shifted the balance between losses and formation from precursors.

Among the four compounds potentially contributing to mouldy–musty off-flavours, (–)-geosmin and 3-methyl-1*H*-indole showed recoveries around 80%. The recoveries for 1*H*-indole were higher, namely 101% in the dark chocolate mass and 93% in the milk chocolate mass. Possibly, losses of 1*H*-indole during conching were in parts compensated by its formation from thermolabile precursors. The rather high recoveries of 1*H*-indole during conching were thus in line with its clear increase during roasting. MDMF behaved differently from all other compounds as its recovery with 9% and 11% was quite low.

## Conclusions

Our data suggests that for the off-flavour compounds causing smoky off-flavours, namely 2-methoxyphenol, 3-methylphenol, 4-methylphenol, 3-ethylphenol, 4-ethylphenol, and 3-propylphenol, minor increases during roasting and processing into cocoa liquor are compensated by losses during processing into chocolate mass and conching. For these compounds, no adjustment of the previously suggested maximum tolerable concentrations on the level of the fermented cocoa is, therefore, necessary. The compounds potentially contributing to mouldy–musty off-flavours showed a less homogeneous behaviour. For (–)-geosmin, a clear statement is not possible, because only a single dataset was available to assess the changes during roasting and processing into cocoa liquor. Given the decomposition of 3-methyl-1*H*-indole during roasting and processing into cocoa liquor, its limit might be raised, at least when higher roasting temperatures are applied. By contrast, 1*H*-indole showed a clear increase during roasting and processing into cocoa liquor that was not compensated by processing into chocolate mass and conching. In the incoming goods inspection in the chocolate industry, monitoring of 1*H*-indole should therefore include roasting of the beans before analysis. MDMF needs not to be included in the monitoring. Its concentration in the fermented beans is typically below its relatively high odour threshold value and is further reduced during processing.

**Supplementary Information** The online version contains supplementary material available at <https://doi.org/10.1007/s00217-021-03873-0>.

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## Declarations

**Conflict of interest** The authors declare that they have no conflict of interest.

**Compliance with ethics requirements** This article does not contain any studies with human or animal subjects.

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