





Author for correspondence: Tatiana Reichert Email: tatiana.reichert@tum.de

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# Tansley review

# Plant phosphorus-use and -acquisition strategies in Amazonia

Tatiana Reichert<sup>1</sup> (D), Anja Rammig<sup>1</sup> (D), Lucia Fuchslueger<sup>2</sup> (D),

Laynara F. Lugli<sup>3</sup> (b), Carlos A. Quesada<sup>3</sup> and Katrin Fleischer<sup>1,4</sup> (b)

<sup>1</sup>School of Life Sciences, Technical University of Munich, Freising 85354, Germany; <sup>2</sup>Centre of Microbiology and Environmental

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# Summary

In the tropical rainforest of Amazonia, phosphorus (P) is one of the main nutrients controlling forest dynamics, but its effects on the future of the forest biomass carbon (C) storage under elevated atmospheric CO<sub>2</sub> concentrations remain uncertain. Soils in vast areas of Amazonia are P-impoverished, and little is known about the variation or plasticity in plant P-use and - acquisition strategies across space and time, hampering the accuracy of projections in vegetation models. Here, we synthesize current knowledge of leaf P resorption, fine-root P foraging, arbuscular mycorrhizal symbioses, and root acid phosphatase and organic acid exudation and discuss how these strategies vary with soil P concentrations and in response to elevated atmospheric CO<sub>2</sub>. We identify knowledge gaps and suggest ways forward to fill those gaps. Additionally, we propose a conceptual framework for the variations in plant P-use and - acquisition strategies along soil P gradients of Amazonia. We suggest that in soils with intermediate to high P concentrations, at the plant community level, investments are primarily directed to P foraging strategies via roots and arbuscular mycorrhizas, whereas in soils with intermediate to low P concentrations, investments shift to prioritize leaf P resorption and mining strategies via phosphatases and organic acids.

# I. Introduction

The tropical rainforest of Amazonia provides significant global climate regulation services by sequestering carbon (C) in biomass, probably acting as a net C sink (Brienen *et al.*, 2015; Hubau *et al.*, 2020); this is possibly due to the  $CO_2$  fertilization effect, which implies increased C uptake due to elevated atmospheric  $CO_2$  concentrations (eCO<sub>2</sub>) from anthropogenic activities (Walker

*et al.*, 2021). However, the future of this biomass C sink remains unclear under  $eCO_2$  and climate change, namely warming, changes in precipitation and increasing drought events (Phillips *et al.*, 2009; Zemp *et al.*, 2017; Sampaio *et al.*, 2019). Forest inventory data point to a potential saturation of the biomass C sink of Amazonia (Hubau *et al.*, 2020), while most dynamic vegetation models predict an increase in this C sink with  $eCO_2$  (Fleischer *et al.*, 2019). Few of these vegetation models consider phosphorus (P) feedbacks,

though P is one of the crucial limiting nutrients for plant productivity (Vitousek *et al.*, 2010). The vegetation models that do consider P predict a significantly lower biomass C sink response to  $eCO_2$  than those that do not (Fleischer *et al.*, 2019). In Amazonia, where very weathered and low-P soils characterize large areas, it is highly uncertain whether plants can meet the increased P demand for the projected biomass growth under  $eCO_2$  conditions.

Plants only take up inorganic P (Pi) from the soil solution (available P<sub>i</sub>) (Lambers, 2022). In young soils, P<sub>i</sub> becomes available mainly through the weathering of parent material, and most of the total P (Pt) is lost or transformed to less available forms over geological timescales (Walker & Syers, 1976). Therefore, in highly weathered soils, only small amounts of P<sub>i</sub> are available to plants, and most P is in organic forms (Po) or bound to soil minerals or aluminum (Al) and iron (Fe) (hydr)oxides (Walker & Svers, 1976). In Amazonia, geological development has created soil P gradients (Quesada et al., 2010, 2011). In the Andean Cordillera, in the western region, soils form mainly on younger geological substrates with soil  $P_t$  concentrations of up to 1000 mg kg<sup>-1</sup> (Quesada *et al.*, 2010). By contrast, soils in central Amazonia vary in geological age but are highly weathered, with Pr concentrations ranging from 25 to 200 mg kg<sup>-1</sup> (Quesada *et al.*, 2010). In the northern, southern and eastern regions, soils are formed predominantly on old, highly weathered parent material, with Pr concentrations rarely exceeding 200 mg kg<sup>-1</sup> (Quesada et al., 2010, 2011). Although large forest areas occur on soils with low P concentrations, the forests are highly productive (Quesada et al., 2012).

Plants have adapted and evolved several strategies to cope with low P<sub>i</sub> supply in soils (Lloyd et al., 2001; Lambers et al., 2008; Sánchez-Calderón et al., 2010). They can increase their P-use efficiency by reducing their P requirement, optimizing P allocation and increasing the residence time of P, for instance by remobilizing P from senescing organs, including leaves, roots and the sapwood (Chapin III, 1980; Veneklaas et al., 2012; Heineman et al., 2016). They can enhance P<sub>i</sub> acquisition by investing in absorptive roots and acclimating their root architecture and morphology to improve their P<sub>i</sub>-foraging capacity and efficiency (Lynch & Brown, 2001; Lambers et al., 2006). They can release phosphatase enzymes that mineralize Po forms into available Pi (Nannipieri et al., 2011) and release low-molecular-weight organic acids that can mobilize Pi and Po from the soil matrix (Lambers et al., 2006). Moreover, roots can interact with microorganisms such as fungi and bacteria in the rhizosphere, which might increase plant P<sub>i</sub> availability and uptake (Box 1; Richardson & Simpson, 2011). Mycorrhizal fungi, in particular, can directly connect with plants by colonizing and acting as an extension of the roots, increasing soil surface area for P<sub>i</sub> acquisition (Smith et al., 2004).

The variation in plant P-use and -acquisition strategies among species and in communities depends on plant phylogenetic constraints, nutrient demand and supply, and environmental stimuli (Lambers *et al.*, 2008; Kramer-Walter *et al.*, 2016; Valverde-Barrantes *et al.*, 2016; Canarini *et al.*, 2019). Plant economic theory predicts that plants can adjust their internal resource allocation to acquire the most limiting resource (Bloom *et al.*, 1985). There is evidence that the energy costs of mechanisms and soil P concentrations can explain some of the variation in plant Box 1 Rhizosphere microorganisms and plant interactions.

Microorganisms (mycorrhizal and saprotrophic fungi, bacteria, archaea) constitute a significant pool of living biomass in soils. They may contain as much P as plant biomass (Turner et al., 2013) and are the dominant metabolic machines driving organic matter decomposition and turnover, linking C and P cycling (Falkowski et al., 2008). Specialized microorganisms can mobilize Pi through different pathways, such as exudation of phosphatase enzymes, organic acids and siderophores (Fe-chelating compounds: Khan et al., 2014). By immobilizing P in their living biomass, the soil microbial community can act as a sink for P, and by remineralization after cell death, they can be a source of available P<sub>i</sub> (Achat et al., 2010). As microbial biomass P turns over rapidly, the dynamic exchange between the microbial biomass and the soil solution contributes significantly to replenishing P in the soil solution contributing to forest P nutrition (Achat et al., 2010, 2012). Litter and soil organic matter availability, composition and stoichiometry can exert control over microbial activity (Mooshammer et al., 2014). Under eCO2, increased leaf Piresorption proficiency could decrease P returns for soil microbes and slow litter decomposition (Jin et al., 2015). Increased plant investments in P acquisition under eCO<sub>2</sub>, such as in longer and finer roots with faster turnover rates, increased root exudation of phosphatases, and labile C such as sugars and low-molecular-weight organic acids (LMWOAs), provide an easily available C source for microbial communities, stimulating their metabolic activity and biomass turnover (Spohn et al., 2013; Jin et al., 2014; but see Xu et al., 2017). This stimulation of microbial activity has been related to increased P<sub>o</sub> in the rhizosphere, which could contribute to enhancing plant P<sub>i</sub> uptake under eCO<sub>2</sub> (Jin et al., 2014). However, as microorganisms use plant-derived C sources, they may reduce the effectiveness of plant strategies to acquire P<sub>i</sub> (Deubel & Merbach, 2005; Fujii et al., 2012), which could be counteracted by increasing root surface area, for instance, increasing plant competitiveness for  $P_i$  in the solution. The methodological challenges associated with quantifying these interactions in the rhizosphere hinder our understanding and implementation of these dynamics in vegetation models (Deubel & Merbach, 2005).

P-use and -acquisition strategies (Box 2; Lynch & Ho, 2005; Lambers *et al.*, 2008; Raven *et al.*, 2018). Plant communities shift and utilize different P-use and -acquisition strategies along soil P gradients, and in highly infertile soils, communities rely on a range of strategies (Zemunik *et al.*, 2015). Similarly, in the low-P soils of central Amazonia, plants have adopted multiple  $P_i$ -acquisition strategies to meet their  $P_i$  needs (Lugli *et al.*, 2020). However, there is no synthesized knowledge of how plant P-use and -acquisition strategies vary across soil P gradients in Amazonia.

Understanding the underlying mechanisms and variation in plant P-use and -acquisition strategies along soil P gradients is highly relevant to reliably project the resilience of Amazonia under climate change and eCO<sub>2</sub>. Currently, projected P feedbacks to eCO<sub>2</sub> in Amazonia from process-based vegetation models diverge due to different representations of these plant strategies (Box 3; Fleischer *et al.*, 2019). Based on these model projections, the ability of plants to upregulate P-use and -acquisition strategies under eCO<sub>2</sub> is decisive for the future of Amazonia but remains unclear. Similarly, climatic changes have the potential to alter the structure

#### **Box 2** Costs of plant P<sub>i</sub> use and acquisition.

The costs of plant P<sub>i</sub> use and acquisition include resources allocated to the construction, function and maintenance of all the elements involved (Lynch & Ho, 2005). To improve tropical forest projections in vegetation models, a better understanding of these costs is essential, particularly the costs of C and N per P<sub>i</sub> acquired and how they vary. Few studies have focused on this subject. For instance, Raven et al. (2018) estimated and compared the minimum C cost per Pi acquired via different strategies and discussed how they might vary. They proposed that the costs of acquiring P<sub>i</sub> via foraging strategies increase with decreasing available soil P<sub>i</sub> concentrations in the soil. At high soil P<sub>i</sub> concentrations, the C cost per P<sub>i</sub> acquired via roots and root hairs may be initially lower than via arbuscular mycorrhizal fungi (AMF) due to the high cost of P transport through hyphae. As P<sub>i</sub> concentrations decline, the C costs of acquiring P<sub>i</sub> via AMF become more competitive than those of roots, as hyphae can more efficiently explore large volumes of soils per unit of C invested (Raven et al., 2018). Similarly, with increasing concentrations of sorbed P in the soil, the C cost per P<sub>i</sub> acquired via LMWOAs may decrease, as P<sub>i</sub> return may increase, with roots probably being a more effective pathway than ectomycorrhizas (Raven et al., 2018). This relationship among soil P pools and the C costs of P<sub>i</sub> acquisition can facilitate the implementation of these costs in vegetation models by making the C cost per Pi acquired a result of a function of soil P pools, for instance. A similar approach was taken by a nutrient uptake optimization model (FUN 3.0), which showed that C costs of acquiring P<sub>i</sub> in dry tropical forests were about three times greater than those of temperate forests (Allen et al., 2020). However, the lack of data on C and P fluxes and pool dynamics related to different P-use and -acquisition strategies are still challenging the parameterization and validation of such modeling approaches.

and functioning of Amazonia (Brando *et al.*, 2008; Olivares *et al.*, 2015); here, we focus on the effects of  $eCO_2$  on plant P-use and -acquisition strategies that are currently considered in vegetation models, synthesize observational knowledge for model parameterization and benchmarking, and additionally inform future research.

We consider five strategies: leaf P<sub>i</sub> resorption; root P<sub>i</sub> foraging; arbuscular mycorrhizal fungal symbioses; and exudation of acid phosphatase enzymes; and low-molecular-weight organic acids (LMWOAs) (Fig. 1). Focusing on Amazonia, we discuss how plant P-use and -acquisition strategies vary with soil P concentrations and how eCO2 may affect these strategies under low soil P availability in the short term (decades). We have categorized soil Pr concentrations and often refer to them as very high (>  $800 \text{ mg P kg}^{-1}$ ), intermediate-high (800- $501 \text{ mg kg}^{-1}$ ), intermediate-low ( $500-200 \text{ mg kg}^{-1}$ ) and very low ( $\leq 200 \text{ mg kg}^{-1}$ ). Although multiple P-use and -acquisition strategies can cooccur in individual plants and communities, we review them separately but discuss tradeoffs among root morphological traits and the strategies. Moreover, we suggest a conceptual framework for the relative distribution of plant P-use and -acquisition strategies along soil P gradients in Amazonia. Finally, we highlight critical knowledge gaps and measurements needed to deepen our understanding of variation

#### Box 3 Plant P-use and -acquisition strategies in vegetation models.

Vegetation models differ in their representation of plant P-use and acquisition strategies, while similarities can still be found. Leaf P<sub>i</sub> resorption, for instance, is a fixed coefficient based on resorption efficiency, ranging from 50% to 75% (Fleischer et al., 2019). Resorption proficiency is a modeled outcome and can be benchmarked with field observations. So far, models neither consider nonlinear relationships of resorption efficiency with plant P stress nor energy costs. Fine root production based on fixed C allocation fractions is increasingly being replaced by dynamic modeling schemes considering resource availability and allometric relationships between roots and leaves (e.g. Thum et al., 2019). Currently, root Pi uptake from the soil solution is represented by functions of root biomass and parameters describing P<sub>i</sub>-uptake efficiency or kinetics of uptake from the soil solution. Root exudation of phosphatases is represented implicitly as 'biochemical P mineralization', which can be upregulated under higher plant P demand and decouples P mineralization from organic matter decomposition (e.g. Goll et al., 2017). Similarly, root exudation of LMWOAs (or other exudates) is only implicitly represented in a few vegetation models, where P desorption is upregulated when plants experience P limitation (Fleischer et al., 2019). However, the benefits of P<sub>i</sub>-mining strategies come at no cost for plants. Currently, there are emerging approaches to representing mycorrhizal Pi uptake explicitly. For instance, in the Coup-CNP model, plants allocate C to fungi as long as the Pi return is advantageous (He et al., 2021). The fungi can explore both the available P<sub>i</sub> pool and P<sub>o</sub> pool, but ultimately P<sub>i</sub> uptake is limited by the fungal biomass (He et al., 2021). Moreover, vegetation models are designed for modeling large-scale vegetation dynamics; consequently, the challenge lies in finding generalities of microscale dynamics occurring at the root surface that allow upscaling along resource gradients and projecting impacts of global change at the ecosystem scale. Recently, more complex interactions between mineral surfaces, microorganisms and plants in their competition for P in the soil solution have been included (Zhu et al., 2016). However, such schemes require data for parameterization that may not be easily quantifiable in practice. Incomplete process understanding and a lack of data for parameterization contribute to uncertainties in P feedbacks to environmental change (Fleischer et al., 2019).

in plant P-use and -acquisition strategies over space and time in Amazonia and improve their mechanistic representations in vegetation models.

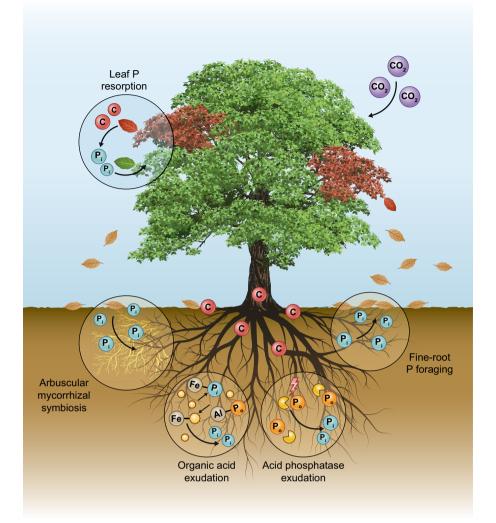
### II. Plant P-use and -acquisition strategies

*Leaf nutrient resorption* is an effective mechanism that increases plant nutrient-use efficiency, that is the biomass production per unit P taken up, by remobilizing nutrients from senescing leaves (Chapin III, 1980; Vitousek, 1982). Globally, it is estimated that leaf nutrient resorption supplies 31% of nitrogen (N) and 40% of plant P<sub>i</sub> needs for new productivity; in tropical forests, resorbed P may supply up to 48% of P<sub>i</sub> needs (Cleveland *et al.*, 2013). Leaf nutrient resorption directly affects biogeochemical cycles, as nutrient concentrations in leaf litter affect decomposition rates (Box 1; Vitousek, 1984). Killingbeck (2004) suggested that the maximum nutrient resorption from leaves is probably a result of evolutionary processes and adaptations to biochemical limitations.

Fig. 1 Illustration of the plant P-use and acquisition strategies reviewed in this study. Leaf P<sub>i</sub> resorption is an indicator for plant P use; P is remobilized from senescing leaves before they fall and is reused in sink organs. Root P<sub>i</sub> foraging can be characterized as the ability of the roots to find and explore soil Pi-rich patches. Arbuscular mycorrhizas act as an extension of absorptive roots and explore large volumes of soil in search of P<sub>i</sub> in exchange for plant C. Acid phosphatases (yellow) are enzymes that mobilize P<sub>i</sub> from organic compounds. Low-molecular-weight organic acids (yellow) exuded by roots can compete for and occupy sorption sites in the solid phase of the soil and form complexes with metals, increasing P<sub>i</sub> and P<sub>o</sub> dissolution, making P<sub>i</sub> available for uptake, or Po available for hydrolysis by phosphatases. Plants must allocate C to implement P-use and -acquisition strategies, which is commonly referred to as the 'C cost' or 'C investment'.

By contrast, the actual resorption of nutrients from leaves varies within the lifetime of a plant and depends on factors such as resource availability (water, nutrients, light), available energy and disturbances. Leaf nutrient resorption is typically discussed in terms of efficiency, the proportion of nutrients resorbed, or in terms of proficiency, the final concentration of nutrients that remains in senesced leaves as they are shed (Killingbeck, 1996). We focus on leaf P<sub>i</sub> resorption in terms of proficiency, which is less prone to methodological bias than efficiency (Supporting Information Notes S1).

*Fine roots* are commonly defined as below 2 mm in diameter (but see McCormack *et al.*, 2015). In highly weathered soils, where P input from parent material is negligible, P typically accumulates in the topsoil; therefore, root traits and adaptations that improve topsoil exploration are advantageous (Lynch & Brown, 2001, 2008). In Amazonia, fine roots in the upper 30 cm of soil can account for more than 50% of the total fine-root biomass in the soil profile (Nepstad *et al.*, 1994; Trumbore *et al.*, 2006; Cordeiro *et al.*, 2020). In some nutrient-impoverished soils of Amazonia, fine roots can be densely concentrated in the litter layer on top of the soil, forming superficial root mats up to 50 cm thick (Stark & Jordan, 1978; Jordan & Escalante, 1980; Medina *et al.*, 1980),



which is a highly efficient strategy to intercept nutrients and prevent leaching (Stark & Jordan, 1978; Martins *et al.*, 2021).

Fine roots can acquire  $P_i$  through strategies that can be classified as foraging and mining. Here, we define fine-root  $P_i$  foraging as related to the capacity of the roots to find and explore soil  $P_i$ -rich patches, which has been associated mainly to root architectural traits, such as branching intensity and angles, and morphological traits, such as root hairs, length, diameter, specific root length, specific root area and root tissue density (Lynch & Brown, 2001, 2008; Hodge, 2004). By contrast, fine-root  $P_i$  mining is related to the capacity of the roots to acquire  $P_i$  from less available forms, for instance through the release of phosphatases or organic acids. Here, we focus on fine-root morphological traits and consider tradeoffs among traits fostering different  $P_i$ -acquisition strategies.

*Mycorrhizal fungi* provide various benefits to plants in exchange for C, including nutrients, water and pathogen defense (Tedersoo *et al.*, 2020). Arbuscular mycorrhizal fungi (AMF) and ectomycorrhizal fungi are commonly associated with trees (Brundrett & Tedersoo, 2018). In tropical forests, AMF are the most abundant, widespread and critical fungi for plant P<sub>i</sub> uptake (Smith & Smith, 1990; Wang *et al.*, 2017). Although ectomycorrhizas can efficiently mine P (Plassard *et al.*, 2011), they are more often related to N-poor

#### Box 4 Soil organic P and mineralization.

Phosphomonoesters and phosphodiesters are the most common forms of P<sub>o</sub> in soils (Turner, 2008). Inositol phosphates, primarily phytate, are a more complex form of phosphomonoester, yet are an important fraction of soil Po. Simple phosphomonoesters are hydrolyzed by phosphomonoesterases, the dominant class of phosphatases expressed in roots (Jarosch et al., 2019), while phytate can be hydrolyzed only by phosphatases known as phytases. By contrast, phosphodiesters must first be hydrolyzed by phosphodiesterase and subsequently by phosphomonoesterases to make Pi available (Turner, 2008). We currently know little about the variation in the composition of Po compounds in Amazonian soils. In general, phytate seems to accumulate in some soils as it is strongly adsorbed to the soil matrix (Gerke, 2015b); consequently, phytate must be mobilized before hydrolysis. In a range of tropical soils, Po mineralized by phytases accounted for 51% of all hydrolyzable P (Darch et al., 2016). By contrast, more labile phosphomonoesters can be more rapidly utilized by phosphomonoesterases (Turner, 2008). Nevertheless, a study considering various soil types, including ferralsols, indicated that the hydrolysis of phytate and phosphodiesters in the soil might be limited by enzymes, while the hydrolysis of phosphomonoesters might be limited by substrate (Jarosch et al., 2019), although it is unclear if the same pattern applies to the rhizosphere. Root phytase activity is a research topic that has been little explored in tropical forests. In a low-P Bornean tropical rainforest, Pi fertilization reduced root phytase activity significantly, suggesting that P-limited trees might depend on phytate as a Pi source (Yokoyama et al., 2017). In another experiment with four tropical plant species with contrasting Pi-acquisition strategies, the nonmycorrhizal Proteaceae species showed significantly higher phytate usage (Steidinger et al., 2014). This could be related to the ability of Proteaceae species to exude large quantities of LMWOAs, thus mobilizing phytate in the rhizosphere. Previous studies suggest that phytate may be an important P<sub>i</sub> source, particularly in low-P soils where plants may exude LMWOAs at rapid rates, probably representing the Piacquisition strategy with the highest C cost (Turner, 2008).

soils, supporting plant N uptake (Stuart & Plett, 2020), and their role in tropical forests has received less attention compared with AMF. In the neotropics, nearly 80% of the tree species may form symbioses with AMF, while it is estimated that only 6% of the species form symbioses with ectomycorrhizas, and 19% in the paleotropics (Corrales *et al.*, 2018). In Amazonia, ectomycorrhizas are considered rare but may play a significant role in white-sand forests on arenosols (Roy *et al.*, 2016). Since little is known about the role of ectomycorrhizas in Amazonia, and arenosols may cover less than 3% of the area of the Amazon basin (Quesada *et al.*, 2011), we focus on AMF.

Plant P<sub>i</sub> uptake through the AMF pathway relative to the direct root pathway is highly variable (Smith *et al.*, 2004; Nagy *et al.*, 2009; Stonor *et al.*, 2014). The presence of AMF can suppress or deactivate the expression of P<sub>i</sub> transporters involved in direct root P<sub>i</sub> uptake (Smith *et al.*, 2004; Courty *et al.*, 2016), but the mechanisms controlling P<sub>i</sub> uptake through the AMF vs the root pathway are not completely understood (see Kobae, 2019). Like roots, AMF hyphae rely on specialized P<sub>i</sub> transporters and can only take up available P<sub>i</sub> (Saito & Ezawa, 2016). The capacity of AMF to support plant P<sub>i</sub> uptake has often been related to their P<sub>i</sub>-foraging efficiency. AMF hyphae have a potential soil exploration efficiency estimated to be six times greater than that of fine roots alone (McCormack & Iversen, 2019), and the P<sub>i</sub> uptake capacity of AMF is positively correlated with the extraradical hyphal network surface area (Jakobsen *et al.*, 1992). Moreover, AMF may also mobilize P<sub>i</sub> from various pools, for example organic and sorbed P, and possibly apatite (Bolan *et al.*, 1987; Yao *et al.*, 2001; Andrino *et al.*, 2019, 2021). Studies in tropical forests suggest that the litter layer may be an essential source of nutrients to AMF (Posada *et al.*, 2012; Sheldrake *et al.*, 2018). Possible mechanisms include the release of phosphatases (Sato *et al.*, 2015; Zeng *et al.*, 2018) or the stimulation of P-mobilizing bacteria (Box 1; Zhang *et al.*, 2016; Jiang *et al.*, 2021).

Phosphatase enzymes hydrolyze a wide range of substrates (i.e. phosphorylated organic compounds) (Box 4). In Amazonia, Pi availability decreases with the intensification of geological weathering, so that Po hydrolyzed by phosphatase enzymes becomes an increasingly important source of Pi for plants (Quesada et al., 2010). The expression and exudation of phosphatases by microorganisms and plant roots is regulated by P<sub>i</sub> demand and availability (McGill & Cole, 1981; Allison et al., 2010), but in some soils, enzyme expression may be constrained by N availability (Notes S2). Phosphatase activities depend highly on the concentration of the enzymes and the substrate and factors such as pH and temperature (Tabatabai, 1994). Rates are typically measured in laboratory assays, most determining the potential but not the actual field activity rates (Nannipieri et al., 2018). We focus on the root surface potential activity of extracellular acid phosphomonoesterase enzymes (PMEs), the class of phosphatases most widely studied in tropical forests (but see Box 4).

LMWOAs are C compounds containing one or more carboxyl groups (Ryan et al., 2001) and can originate from roots, microorganisms and organic matter in soils (Adeleke et al., 2017). In the soil, LMWOAs can occupy and compete for sorption sites and form complexes with Al and Fe (hydr)oxides at acidic pH or calcium at alkaline pH, preventing P sorption and accelerating desorption, making P<sub>i</sub> available for uptake or P<sub>o</sub> available for mineralization (Wang & Lambers, 2019). The rates of root LMWOA exudation are highly variable and depend mainly on the species (Jones, 1998), plant nutritional status (Lambers et al., 2006) and environmental stimuli (Canarini et al., 2019). The effectiveness of LMWOAs in mobilizing P in the soil increases with increasing LMWOA concentrations and differs among different LMWOAs (Jones, 1998; Adeleke et al., 2017). Dicarboxylates (malate, oxalate) and tricarboxylates (citrate) are often reported as most effective at mobilizing P, as they have a high affinity to sorption sites in the soil (Gerke, 2015; Darch et al., 2016), but in the soil solution, they may be rapidly biodegraded (Van Hees et al., 2002; Fujii et al., 2012), reducing their effectiveness (Box 1).

In extremely low-P soils, some species have developed specialized proteoid and dauciform roots, highly efficient in mobilizing P by accumulating and exuding large quantities of LMWOAs over a short period, resulting in very high LMWOA concentrations in the rhizosphere (Lambers *et al.*, 2006). However, it remains unknown how frequently species forming such specialized roots occur and under what circumstances they are formed in Amazonia (Notes S3).

# III. Soil P effects on plant P-use and -acquisition strategies

# 1. Leaf P<sub>i</sub> resorption proficiency

Globally, at the plant community level, leaf P concentrations in senesced leaves consistently decline with declining soil P concentrations; that is, leaf P<sub>i</sub>-resorption proficiency increases (Richardson et al., 2005; Tang et al., 2013; Hayes et al., 2014). This phenomenon has been attributed to the plasticity of frequent species and species turnover along P gradients (Richardson et al., 2005). The same pattern of leaf P<sub>i</sub>-resorption proficiency has been observed along soil P gradients in the French Guianese and Colombian Amazonia (Lips & Duivenvoorden, 1996; Soong et al., 2020), as well as other tropical forests (Fig. 2; Table S1; Vitousek, 1998; Hidaka & Kitayama, 2011; Tsujii et al., 2017). For instance, along a soil P gradient in French Guiana (7–600 mg  $P_t$  kg<sup>-1</sup>), P concentrations in senesced leaves varied approximately four-fold (Soong et al., 2020). Moreover, several studies across Amazonia suggest a high variation in leaf P<sub>i</sub>-resorption proficiency (Table S2). Measured P concentrations in senesced leaves can vary from  $0.09 \text{ mg g}^{-1}$  (Hättenschwiler *et al.*, 2008) to 0.75 mg g<sup>-1</sup> (Dantas & Phillipson, 1989), while the average in tropical forests, as shown in a meta-analysis, is  $0.4 \text{ mg g}^{-1}$  (Yuan & Chen, 2009).

Soil P<sub>i</sub> fertilization experiments additionally support the effect of soil P fertility on leaf Pi-resorption proficiency. A global meta-analysis showed that P; fertilization increased P concentrations in senesced leaves by 82% (Yuan & Chen, 2015). Responses to P<sub>i</sub> fertilization have not been reported in Amazonia; however, results from other tropical forests suggest that Pi-resorption proficiency may decrease in low-P soils with P; additions, but not in high-P soils (Fig. 2; Table S3). In tropical forests with low-P soils in Hawaii, Indonesia and Ecuador, Pi fertilization significantly increased P concentrations in senesced leaves (Vitousek, 1998; Mirmanto et al., 1999; Homeier et al., 2012). In a tropical forest in Panama, with intermediate soil Pt concentrations, Pi fertilization slightly increased P concentrations in senesced leaves, but the differences were significant only for one out of four species (Mayor et al., 2014). Moreover, in tropical forests with high-P soils in Hawaii and Costa Rica, Pi fertilization did not affect P concentrations in senesced leaves (Vitousek, 1998; Alvarez-Clare & Mack, 2015). Thus, plants may invest in leaf P<sub>i</sub> resorption to conserve P only when soil P is scarce (Fig. 3). Studies suggest that the C and N costs of P resorption from leaves may increase with decreasing soil P concentrations, as plants in low-P soils achieve high proficiency by increasing P<sub>i</sub> resorption from leaf fractions requiring hydrolysis, and thus investment in various enzymes (Hidaka & Kitayama, 2011; Veneklaas et al., 2012; Tsujii et al., 2017).

# 2. Root morphology and tradeoffs among $\mathsf{P}_{i}\text{-}acquisition}$ strategies

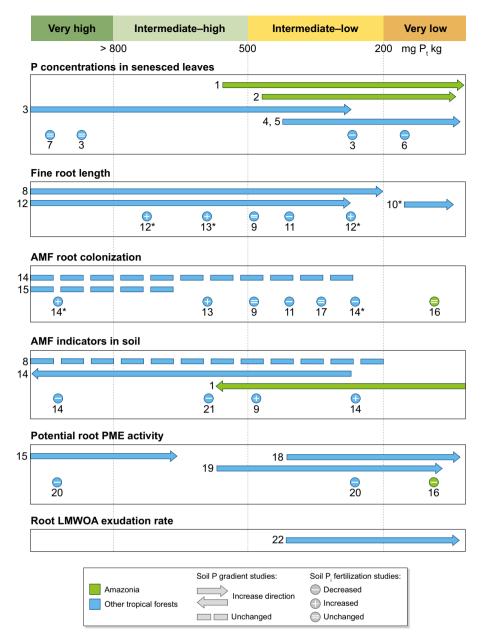
Across Amazonia, root morphological traits vary significantly (Table S4), but their relationship with soil P concentrations remains mostly unclear. Root morphological traits such as length, specific root length, specific root area and root tissue density

typically increase, while diameter decreases at the low end of a soil P gradient; thus, fine roots become thinner, longer and probably longer-lived (Lynch & Brown, 2001; Holdaway et al., 2011; Kramer-Walter et al., 2016). Nonetheless, high specific root length and specific root area were also observed in young soils (Holdaway et al., 2011; Girardin et al., 2013), which may be related to the acquisition of N (Freschet et al., 2021). In different tropical forests, including forest sites in northeastern Amazonia, fine-root length, diameter, specific root length and specific root area followed the expected patterns (Table S1; Ostertag, 2001; Powers et al., 2005; Metcalfe et al., 2008; Zangaro et al., 2008; Kochsiek et al., 2013; Ushio et al., 2015). Similarly, a meta-analysis of tropical forests worldwide showed that specific root length tended to be higher in sites with very low soil P<sub>i</sub> availability (Addo-Danso et al., 2020). By contrast, root tissue density did not change along a soil P gradient in Borneo (Ushio et al., 2015), whereas in central Amazonia, it was positively correlated with available P<sub>i</sub>, although the range of soil P variation was small (Lugli et al., 2020). Despite the low data availability, changes in root morphological traits along soil P gradients suggest increased fine-root nutrient uptake capacity and increased soil exploration efficiency (C per length unit) with declining P<sub>i</sub> availability (Fig. 2).

In tropical forests, soil P<sub>i</sub> fertilization rarely affected root morphological traits (Fig. 2; Table S3). It neither affected fine root length along a soil chronosequence in Hawaii (Ostertag, 2001) nor various fine root morphological traits in central Amazonia and Panama (Yavitt et al., 2011; Wurzburger & Wright, 2015), apart from increasing root diameter in central Amazonia, comparably the most P-impoverished forest (Lugli et al., 2021). By contrast, in a subtropical forest in China, Pi fertilization significantly decreased root length (Liu et al., 2015). This suggests that multiple nutrients may control fine root morphological traits (Wurzburger & Wright, 2015) or that these traits and their plasticity may be highly constrained by plant phylogeny (Holdaway et al., 2011; Kramer-Walter & Laughlin, 2017; Valverde-Barrantes et al., 2017). For instance, in a pot experiment with seedlings of 17 Amazonian species growing at different P levels, only a few species showed plastic responses in specific root length (Quintero-Vallejo et al., 2015).

The distinction among plants that meet their P<sub>i</sub> needs through fine root foraging, from those that outsource P<sub>i</sub> uptake to mycorrhizal fungi, and those that actively mine P<sub>i</sub>, is challenging based on root morphological traits alone. This distinction should be based on additional Pi-acquisition indicators, such as root elongation rate and hair density (Freschet et al., 2021), and the tradeoffs among traits fostering different Pi-acquisition strategies. For instance, absorptive roots in low-P soils of central Amazonia have high specific root length and area and small diameter (Lugli et al., 2020). Roots hosting AMF are often characterized by low specific root length and large diameter, as a large root cortical area might better accommodate fungal structures (McCormack & Iversen, 2019; Bergmann et al., 2020); this suggests that most plants in low-P<sub>i</sub> Amazonian soils might depend less on this strategy. However, it is unlikely that roots, such as those in central Amazonia, are primarily foraging for P<sub>i</sub>; such a strategy would be costly and yield low P; returns. Moreover, root phosphatase activity

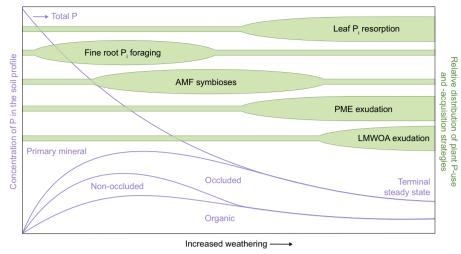
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**Fig. 2** A summary of some of the main soil P effects on different indicators of plant P-use and -acquisition strategies. AMF, arbuscular mycorrhizal fungi; PME, acid phosphomonoesterases; LMWOA, low-molecular-weight organic acids. AMF indicators in soil include measures of hyphal length, biomass or abundance. Arrows represent studies along soil P gradients, and the length of the arrow shows the extent of the gradient covered by the study, while the direction of the arrows determines whether a strategy increased or decreased with soil P. Symbols in the circles represent results from P<sub>i</sub> fertilization experiments, and their placement indicates soil P<sub>t</sub> concentrations in control plots. Note that the effects of soil P<sub>i</sub> fertilization on different indicators could be influenced by the initial P<sub>t</sub> concentrations and composition in soils, soil properties and the quantity and form of P added, which may vary among studies. We classified soil P<sub>t</sub> concentrations as high (> 800 mg kg<sup>-1</sup>), intermediate-high (800–501 mg kg<sup>-1</sup>), intermediate-low (500–200 mg kg<sup>-1</sup>) and very low (< 200 mg kg<sup>-1</sup>). Numbers point to references that are summarized in Supporting Information Tables S1 and S3. References marked with an asterisk refer to results that showed tendencies, but differences were not statistically significant. <sup>1</sup>Soong *et al.* (2020); <sup>2</sup>Lips & Duivenvoorden (1996); <sup>3</sup>Vitousek (1998); <sup>4</sup>Hidaka & Kitayama (2011); <sup>5</sup>Tsujii *et al.* (2017); <sup>6</sup>Mirmanto *et al.* (1999); <sup>7</sup>Alvarez-Clare & Mack (2015); <sup>8</sup>Powers *et al.* (2005); <sup>9</sup>Camenzind *et al.* (2016); <sup>10</sup>Kochsiek *et al.* (2013); <sup>11</sup>Liu *et al.* (2014); <sup>18</sup>Ushio *et al.* (2015); <sup>19</sup>Cabugao *et al.* (2017); Cabugao *et al.* (2021); <sup>20</sup>Treseder & Vitousek (2001); <sup>21</sup>Sheldrake *et al.* (2018); <sup>22</sup>Aoki *et al.* (2012).

was positively correlated with specific root length and area in central Amazonia and other tropical forests (Ushio *et al.*, 2015; Lugli *et al.*, 2020; Cabugao *et al.*, 2021). Similarly, root organic acid exudation rates were positively correlated with specific root area in tropical forests in Borneo (Aoki *et al.*, 2012; Fujii *et al.*, 2021). These results

suggest that long and fine roots in low-P Amazonian soils are primarily mining  $P_i$  to meet their needs. We speculate that along soil P gradients in Amazonia, independent fine-root  $P_i$  foraging would be a viable primary strategy for plants in soils with high  $P_i$ availability (Fig. 3). In such soils, fine-root  $P_i$  foraging is probably



**Fig. 3** Conceptual representation of the predicted distribution in plant phosphorus (P)-use and -acquisition strategies at the plant community level along soil P gradients in Amazonia. The elongated green bubbles represent where, along a soil P gradient, the respective strategies receive the most resource investments from plant communities and where they contribute the most to meet plant P<sub>i</sub> needs, as they offer advantages under those soil conditions. The conceptual framework was built upon our synthesis and extending previous work on how plant P<sub>i</sub> acquisition may vary along soil nutrient gradients (see Lambers *et al.*, 2008). We predict that P<sub>i</sub> foraging by independent fine roots or in symbioses with arbuscular mycorrhizal fungi (AMF) are primarily used in soils with intermediate to high soil P concentrations. By contrast, mining strategies, namely exudation of phosphomonoesterase enzymes (PMEs) and low-molecular-weight organic acids (LMWOAs), along with finer and longer roots, are prioritized in soils with low to intermediate P concentrations. Similarly, leaf P<sub>i</sub> resorption is increasingly relevant at the low end of the soil P gradient. The image was modified from Turner & Condron (2013) based on the Walker & Syers (1976) model of soil P changes with long-term soil development and supported by soil P data throughout Amazonia (Quesada *et al.*, 2010).

achieved by constructing short-lived roots that may cost more C per unit of soil explored, compared with low-P soils, but yield high  $P_i$  returns, as they can quickly move further from the root P-depletion zone (see Aragao *et al.*, 2009; Lugli *et al.*, 2021).

### 3. Arbuscular mycorrhizal fungi

The abundance of AMF in roots (intraradical colonization) is commonly indicated by the percentage root length colonized by fungal structures. Although this measure is often considered an indicator for plant benefits, it requires careful interpretation, as differences can arise from the level of AMF abundance and/or standing root length (Allen, 2001; Treseder, 2013). Moreover, fungal structures along the root length may not contribute equally to resource exchange, depending on the age of the structures (Tisserant et al., 1996). Nonetheless, a meta-analysis showed percentage root length colonized by AMF explained 23.5% of the changes in plant P content (Treseder, 2013). In contrast to the expectation that AMF root colonization increases with decreasing soil P concentrations, we found no consistent patterns in tropical forests (Fig. 2; Table S1). Along soil P gradients in Hawaii and Costa Rica, AMF root colonization was unchanged (Treseder & Allen, 2002; Nasto et al., 2014). On the other hand, in southwest Brazil, root AMF colonization increased with decreasing P<sub>i</sub> availability, although forest sites had different succession stages, and root AMF colonization also increased with decreasing succession stage (Zangaro et al., 2008).

AMF measures of soil colonization (i.e. extraradical hyphal length, biomass or abundance) could be an excellent complementary indicator of AMF P<sub>i</sub>-uptake capacity (see Jakobsen *et al.*, 1992;

Sheldrake et al., 2018). Along soil chronosequences in Hawaii and Australia, AMF extraradical hyphal length and biomass decreased with decreasing soil P concentrations (Table S1; Treseder & Allen, 2002; Balser et al., 2005; Teste et al., 2016). A similar pattern emerged along a soil P gradient in French Guiana, where AMF abundance in soil declined with declining soil Pt concentrations (Soong et al., 2020). These previous studies support the soil P control on AMF abundance and functioning and suggest that AMF may play a smaller role in soils with very low P concentrations (Treseder & Allen, 2002). By contrast, in a study including forest sites in Peru, Panama and Costa Rica, mostly in high-P soils, AMF extraradical hyphal length was not affected by soil P (Powers et al., 2005). Hyphal length in soils was much lower than in other tropical forests and ecosystems, and various sites did not contain hyphae in the soil, particularly in Panama, suggesting that plants may meet their nutrient needs exclusively through roots (Powers et al., 2005). However, that is contradictory to later studies conducted in the same area in Panama, suggesting AMF are abundant both in roots and in soil (Wurzburger & Wright, 2015; Sheldrake et al., 2018).

Responses of AMF to soil  $P_i$  fertilization varied in tropical forests (Fig. 2; Table S3). In soils with intermediate to very low  $P_t$  concentrations, the percentage root length colonized by AMF was unchanged or decreased in response to  $P_i$  fertilization. In central Amazonia and Ecuador,  $P_i$  fertilization did not change root colonization by AMF (Camenzind *et al.*, 2014, 2016; Lugli *et al.*, 2021), while it decreased in Hawaii and subtropical China (Treseder & Allen, 2002; Liu *et al.*, 2015). By contrast, in soils with intermediate to very high  $P_t$  concentrations in Hawaii and Panama, AMF root colonization increased (Treseder & Allen, 2002; Wurzburger & Wright, 2015). Moreover, in soils with

intermediate to very high  $P_t$  concentrations in Hawaii and Panama,  $P_i$  fertilization decreased AMF extraradical hyphal length and biomass, suggesting that plants shifted strategies to rely more on their roots to acquire  $P_i$ , probably through foraging (Fig. 2; Treseder & Allen, 2002; Sheldrake *et al.*, 2018). By contrast, AMF extraradical indicators increased in soils with low  $P_t$  concentrations in Hawaii and Ecuador (Fig. 2; Treseder & Allen, 2002; Camenzind *et al.*, 2016), suggesting plants increased investments in AMF symbioses when  $P_i$  availability increased.

Overall these studies corroborate that at the plant community level, AMF symbioses may be a more beneficial  $P_i$  acquisition strategy for plants in soils with intermediate  $P_i$  concentrations (Fig. 3), as suggested in previous studies (Treseder & Allen, 2002; Lambers *et al.*, 2008). This may be due to AMF and plants being limited by P in low-P soils, and, by contrast, AMF being limited by C in high-P soils due to decreased C investments by plants (Treseder & Allen, 2002). The decrease in C investments in high-P soils could be caused by interactions with low N, for example in young soils, as N can limit C uptake, increasing C trade costs for P (Johnson, 2010). This has been supported by evidence in a dune chronosequence in Australia and in rupestrian grasslands in Brazil, where the number of plants associated with AMF declines significantly with decreasing soil P concentrations (Oliveira *et al.*, 2015; Zemunik *et al.*, 2015, 2018).

### 4. Root acid phosphatase exudation and activity

Root PME exudation and activity, although variable among species (Ushio et al., 2015; Png et al., 2017), are highly responsive to changes in plant P demand and soil P<sub>i</sub> availability. These direct plant responses are well established; increasing soil Pi availability often represses PME expression and activity (Tian & Liao, 2018; Wang & Liu, 2018). Accordingly, root PME activity was negatively correlated with soil P<sub>i</sub> availability in tropical forests in Costa Rica, Borneo, Puerto Rico and Panama (Nasto et al., 2014; Ushio et al., 2015; Cabugao et al., 2017, 2021; Guilbeault-Mayers et al., 2020). However, for Amazonia, there are currently no published studies on how root PME activity varies across soil P gradients. In central Amazonia, root PME activity varied between 15 and 65 µmol 4methylumbelliferyl-P g<sup>-1</sup> fine-root h<sup>-1</sup> (Lugli *et al.*, 2020), which is similar to what was observed in tropical forests in Puerto Rico and Borneo with a similar range of soil Pr concentrations (Kitayama, 2013; Cabugao et al., 2017), but higher than in more fertile soils in Costa Rica and Puerto Rico (Table S5; Nasto et al., 2014; Cabugao et al., 2017).

In P<sub>i</sub> fertilization experiments, root PME activity consistently decreased, following the expected inverse relationship with increasing P<sub>i</sub> availability (Marklein & Houlton, 2012). In central Amazonia, P<sub>i</sub> fertilization significantly decreased root PME activity by 23% (Lugli *et al.*, 2021). The same trend was observed in other tropical forests; root PME activity in response to P<sub>i</sub> fertilization decreased by *c*. 37% and 46% in Hawaii and Borneo, respectively (Treseder & Vitousek, 2001; Yokoyama *et al.*, 2017). Consistent with observations along soil P gradients, the results from P<sub>i</sub> fertilization experiments additionally support the control of soil P on root PME activity.

Variation in root PME activity is also expected to be partially controlled by substrate concentrations. Along a soil P gradient in a Bornean tropical forest, root PME activity showed a strong negative relationship to the most labile form of Po, but not total Po in the soil (Kitayama, 2013). Similarly, there was no relationship between total soil P<sub>o</sub> and root PME activity along a soil P gradient in Puerto Rico (Cabugao et al., 2021). The lack of relationship between total soil Po and root PME activity might be explained by the composition of the soil Po pool. For instance, phytate is strongly adsorbed to the soil matrix and can only be mineralized by phytases (Box 4; Gerke, 2015a). It is therefore plausible that root PME activity was only correlated with the most labile form of  $P_0$  in the soil since simple forms of P monoesters hydrolyzed by PMEs are more labile than phytate, which is not hydrolyzable by PMEs (Gerke, 2015b). Moreover, substrate availability probably affects the plant C and N costs per P; acquired through PMEs; root PME activity increases with substrate availability, probably increasing the cost-benefit ratio of this strategy. Overall, we expect higher rates of PME activity in soils with low P<sub>i</sub> availability and a high proportion of substrate, as PME expression is driven by P<sub>i</sub> demand and supply, and the rate of activity may be determined by substrate concentrations (Fig. 3).

### 5. Root low-molecular-weight organic acid exudation

Rates of root LMWOA exudation in various plant species have mostly been measured in controlled environments and are commonly greater under low soil P<sub>i</sub> supply; these findings have been extensively reviewed in the literature (Jones, 1998; Gerke, 2015a; Adeleke et al., 2017; Wang & Lambers, 2019). Similarly, an increase in the abundance of species that appear to rely on LMWOA exudation for P<sub>i</sub> acquisition has been observed with declining soil P concentrations in different ecosystems (Oliveira et al., 2015; Zemunik et al., 2015, 2018). However, studies investigating rates of root exudation of LMWOAs in natural ecosystems remain rare due to the inherent challenges in collecting exudates in the field (Oburger & Jones, 2018). To date, only one study has investigated root LMWOA exudation rates along a soil P gradient in a tropical forest (Fig. 2; Aoki et al., 2012; see also Fujii et al., 2021). Total LMWOA exudation varied with species but increased with decreasing soil Pt concentrations, with rates of citrate exudation alone up to 1648 nmol C  $g^{-1}$  root  $h^{-1}$  from a Myrtaceae species in the poorest forest (Aoki et al., 2012), which may be similar to rates of citrate exudation in proteoid roots of Lupinus albus reported in different studies (Roelofs et al., 2001 and references therein).

Concentrations of LMWOAs are thought to be high in soils dominated by organic and adsorbed P, such as highly weathered tropical soils (Aoki *et al.*, 2012; Darch *et al.*, 2016). In tropical soils, the efficacy of LMWOAs seems to be site-dependent; low concentrations of citric acid (10  $\mu$ mol g<sup>-1</sup> soil) were able to mobilize up to 8.06 mg P<sub>t</sub> kg<sup>-1</sup> (Darch *et al.*, 2016). In low-P soils in subtropical China, citric, malic and oxalic acids significantly mobilized P<sub>i</sub> and P<sub>o</sub>, although P<sub>o</sub> more effectively (Hou *et al.*, 2018). In another study, low concentrations of citric acid significantly mobilized P<sub>o</sub> but not P<sub>i</sub> (Wei *et al.*, 2010). Hence, the exudation of LMWOAs in tropical soils might be an efficient strategy even without cluster-root formation, and may be particularly important to mobilize  $P_o$  (Andrade *et al.*, 2003; Gerke, 2015a; Darch *et al.*, 2016). Although no study has investigated the role of LMWOAs in Amazonia, given observations from other ecosystems and under controlled conditions, we expect faster rates of LMWOA exudation in highly weathered Amazonian soils, where a greater proportion of  $P_t$  is adsorbed to the soil matrix (Fig. 3).

# IV. Elevated CO<sub>2</sub> effects on plant P-use and - acquisition strategies

### 1. Leaf P<sub>i</sub> resorption proficiency

Only a few experiments have investigated leaf P<sub>i</sub> resorption under eCO2. Of these, most experiments have been performed in temperate regions, where P concentrations in fully senesced leaves have not changed or have declined only slightly and not significantly (Finzi et al., 2001; Frost & Tuchman, 2005; Zheng et al., 2010; Dray et al., 2014). The lack of response to eCO<sub>2</sub> in temperate regions may have occurred because the plants were not limited by P. However, in free-air CO<sub>2</sub> concentration enrichment (FACE) studies in a eucalypt forest in Australia (EucFACE, Crous et al., 2019) and in shrubs in the Mojave Desert (Housman et al., 2012), both ecosystems considered to be P-limited, no significant changes in leaf P resorption were observed under eCO<sub>2</sub>. In the EucFACE, very low P concentrations were found in senesced leaves  $(0.32 \pm 0.04 \text{ mg g}^{-1})$ ; this very high proficiency could mean that the trees had already reached maximum leaf Pi resorption levels (Crous et al., 2019). Similarly, plants in the most P-poor soils of Amazonia may be unable to upregulate leaf P<sub>i</sub> resorption further in response to eCO<sub>2</sub>, as the reported proficiency may be near their maximum (Table S2). Therefore, the potential to upregulate leaf  $P_i$  resorption in response to  $eCO_2$ might be limited to relatively more fertile regions of Amazonia, where plants have not yet reached their maximum resorption proficiency.

## 2. Root morphology

Most root morphological traits have received very little attention in eCO<sub>2</sub> studies, with fine root length being the most commonly measured root trait. Overall, fine root diameter and length responded positively to eCO2 across various plant types and ecosystems and increased by 8% and 26%, respectively (Nie et al., 2013). Similarly, a meta-analysis including mostly crop species grown under eCO<sub>2</sub> indicated that soil P did not affect root length responses to eCO2, and, overall, root length increased (Jiang et al., 2020a). It is worth noting that soil P concentrations classified as low in the previous analyses may vary (Jiang et al., 2020a). Consistent with the previous studies, in glasshouse experiments with tropical species in fertilized and unfertilized low-P soils (soil P concentrations not provided), community-level root length increased by c. 50% under eCO2 independent of the soil P level (Körner & Arnone, 1992; Arnone III, 1997). However, at low P, root proliferation towards artificially created nutrient-enriched soil

patches was unchanged, and it was suggested that root foraging efficiency did not increase under  $eCO_2$  (Arnone III, 1997). The interactive effects of soil P concentrations and  $eCO_2$  on root morphological traits in tropical species remain uncertain; however, species-specific responses are expected. Overall, root length shows potential for upregulation by plants under  $eCO_2$  in Amazonia. However, in very low-P Amazonian soils, roots may already be highly efficient at exploring the soil, and root productivity is limited by P (Lugli *et al.*, 2021); hence, responses at the community level might be less pronounced than in relatively more fertile soils.

## 3. Arbuscular mycorrhizal fungi

Globally, mycorrhizas have the potential to moderate the CO<sub>2</sub> fertilization effect on plant growth through their role in nutrient acquisition (Terrer et al., 2019). Although less representative of tropical ecosystems, several meta-analyses showed eCO2 promoted positive responses of various measures in host plants and AMF abundance in roots and soil (Table S6; Treseder, 2004; Alberton et al., 2005; Terrer et al., 2016; Dong et al., 2018). Nonetheless, responses were highly variable, depending on the combination of plant and mycorrhizal species and P<sub>i</sub> supply (Dong et al., 2018). Under eCO<sub>2</sub>, low P<sub>i</sub> availability constrained the extraradical hyphal length, while no significant differences were observed in soil fungal biomass or root colonization (Dong et al., 2018). Similarly, in a Plimited eucalypt forest (EucFACE), eCO2 appeared to have prompted only a slight increase in plant C investments in mycorrhizas, which resulted in a small nonsignificant increase in the mycorrhizal C pool (Jiang et al., 2020b).

Our understanding of the eCO<sub>2</sub> effects on AMF in tropical species is restricted to small-scale experiments with immature plants. In a seedling experiment with four tropical species, root length colonized by AMF increased in response to eCO<sub>2</sub> in intermediate- and highfertility treatments but not in low-fertility treatments (Nasto et al., 2019). Additionally, responses were species-specific, and mostly driven by one of the species, considered a low-P specialist in Panama (Nasto et al., 2019). Moreover, in an artificial glasshouse tropical system with seven species, no hyphal growth or root colonization was observed in response to eCO<sub>2</sub>, although these plants were regularly fertilized (Insam et al., 1999). Overall, AMF responses to eCO2 seem to be species-specific and may be constrained in soils with very low P concentrations. Therefore, at the community level, the positive effects of eCO<sub>2</sub> on AMF are expected to be more pronounced in Amazonian soils with intermediate to high P concentrations. This is in line with the observations that AMF functioning may already be constrained by low P or other nutrients in low-P soils at current CO<sub>2</sub> levels (Treseder & Allen, 2002; Wurzburger & Wright, 2015; Lugli et al., 2021).

### 4. Root acid phosphatase exudation and activity

Only a few studies have investigated root PME activity under  $eCO_2$ , and responses were weak. In the P-limited EucFACE, no changes were detected in root PME activity under  $eCO_2$  (Ochoa-Hueso *et al.*, 2017). Similarly, in *Pinus densiflora* seedlings, root PME activity was unaffected by  $eCO_2$  independent of P supply

treatment (Norisada et al., 2006), and in seedlings of Pinus ponderosa, root PME activity decreased (DeLucia et al., 1997). Moreover, among seedlings of four tropical species, eCO<sub>2</sub> induced an overall increase in root PME activity, which was mainly driven by two N2-fixing species, suggesting the strength of the eCO2 effect on this strategy in tropical forests might depend on community species composition (Nasto et al., 2019). However, root PME activity responses to eCO2 will additionally depend on changes in the quantity and quality of organic matter inputs. While increased plant C availability may increase root PME expression, the actual efficacy of these enzymes will depend on the substrate available. For instance, at EucFACE, leaf litter input and quality did not change significantly under eCO<sub>2</sub> (Crous et al., 2019; Jiang et al., 2020b). An increase in root PME activity in Amazonia in response to increased P demand under eCO2 is more likely to occur in soils where organic matter is the most dominant P source for plants. Upregulating PME expression may increase plant competitiveness for Po, while higher litter inputs may favor PME activity, but lower litter quality may constrain this response.

### 5. Root low-molecular-weight organic acid exudation

A recent meta-analysis including various model plants, crops and Pinus species showed that exudation of LMWOAs significantly increased in response to eCO<sub>2</sub> (Dong et al., 2021). However, few studies have focused on the interactive effects of eCO2 and soil P concentrations on LMWOA exudation. For instance, citrate exudation in Lupinus albus, a species that forms proteoid roots, tended to increase under eCO<sub>2</sub> in plants with a low P<sub>i</sub> supply compared with that in plants with a high P<sub>i</sub> supply, but differences were not significant (Campbell & Sage, 2002; Wasaki et al., 2005). There is currently a small amount of data to speculate on how eCO<sub>2</sub> may change root LMWOA exudation in Amazonia. However, considering the overall positive responses (Dong et al., 2021) and assuming plant C uptake will increase due to increased photosynthetic rates, increases in root LMWOA exudation are expected under eCO<sub>2</sub>, particularly in the most weathered Amazonian soils, where this strategy might offer more advantages. Since a high proportion of P<sub>t</sub> in low-P Amazonian soils is in less available forms, root exudation of LMWOAs may be one of the most critical strategies for plants under eCO2. Moreover, a possible increase in root length density and overlap under eCO2 might cause an increase in LMWOA concentration in the rhizosphere, further enhancing the positive effects of increased exudation by individual roots (Dong et al., 2021).

# V. Discussion and conclusions

Based on our synthesis, we provide a conceptual framework on the relative importance of five plant P-use and -acquisition strategies that work in concert at the plant community level to supply the plants' P<sub>i</sub> needs along soil P gradients in Amazonia (Fig. 3). Our conceptual framework builds upon previous work on how P<sub>i</sub>-acquisition strategies may vary with soil age (see Lambers *et al.*, 2008) and extends it to include other P<sub>i</sub>-use and -acquisition strategies, but focuses mainly on tropical forests across Amazonia.

We further speculate on the response of these strategies to  $eCO_2$ . In summary, we found that:

(1) Leaf  $P_i$ -resorption proficiency consistently increases with decreasing soil P concentrations.  $P_i$  fertilization causes a rapid negative response in leaf  $P_i$  resorption in low-P soils, suggesting this strategy may be energy costly and increasingly relevant in low-P soils. Under eCO<sub>2</sub>, leaf  $P_i$  resorption was unaffected in low- and high-P soils, possibly because in low-P soils, maximum resorption rates were reached. By contrast, in high-P soils, increasing resorption may be more energy costly than acquiring  $P_i$  from the soil. We expect the same pattern in Amazonian soils, so that in soils with intermediate P concentrations, changes in leaf  $P_i$  resorption may be more notable under eCO<sub>2</sub>.

(2) Most studies suggest root nutrient uptake capacity and exploration efficiency (i.e. root length and specific root length) increase as soil P concentrations decline across soil P gradients. However, most root morphological traits did not significantly respond to  $P_i$  fertilization, suggesting they may be limited by multiple nutrients or phylogeny. Trade-offs among morphological traits fostering other  $P_i$ -acquisition strategies suggest fine-root  $P_i$  foraging may be more relevant at higher  $P_i$  availability, where roots can rapidly move past the P-depletion zone. Under eCO<sub>2</sub>, root length showed high potential for upregulation, including under soil P conditions reported as low. Therefore, we expect that eCO<sub>2</sub> may increase root length in vast areas of Amazonia if roots have not yet reached their maximum length.

(3) Root length colonized by AMF does not show consistent patterns in response to soil P changes, while AMF abundance in soil mostly increased with increasing soil P concentrations. Results from  $P_i$  fertilization studies suggest AMF symbioses may be an essential strategy for plants in soils with intermediate P concentrations. Under eCO<sub>2</sub>, responses of extraradical hyphae in soil were constrained by low soil P concentrations; thus, we expect notable plant investments in this strategy in Amazonian soils with intermediate and towards high P concentrations, but not in soils with very low P concentrations.

(4) We found a consistent pattern that potential root PME activity increased with decreasing soil P concentrations, but actual activities and PME efficacy at field conditions remain unknown. We speculate that this strategy is highly relevant for plants growing in soils with low  $P_i$  availability and high substrate availability. Root PME activity in most studies did not respond strongly to eCO<sub>2</sub>, although some species showed increased potential for upregulation. Hence, we expect an increase in PME expression for some species in Amazonia under eCO<sub>2</sub>, but PME efficacy might depend on substrate availability.

(5) Root LMWOA exudation rates increased with decreasing soil P concentrations in a tropical forest, following a trend observed in various species and other ecosystems. Therefore, we predict that LMWOA exudation is essential in highly weathered Amazonian soils, where most soil P is in less available forms, giving plants a competitive advantage. Under eCO<sub>2</sub>, root LMWOA exudation often increased, but interactions between eCO<sub>2</sub> and soil P remain mostly uncertain. In Amazonia, this strategy may have the greatest potential to mobilize significant P amounts under eCO<sub>2</sub>, especially on the low side of the soil P gradient. However, the efficacy of this

strategy may depend on the interplay of roots and rhizosphere microorganisms.

Overall, our review provides evidence that community-level patterns in the investment and the relative importance in plant Puse and -acquisition strategies may exist across soil P gradients in Amazonia and possibly other tropical forests (Figs 2, 3). These patterns may result from species composition and functional trait turnover along soil gradients, as some species have adapted to different nutrient conditions (Condit et al., 2013; Umaña et al., 2021). Although highly variable among species, we also show that most plants have some degree of plasticity to upregulate P-use and acquisition strategies under eCO2. However, in Amazonia, upregulation under eCO<sub>2</sub> may be constrained under a very low Pi supply, as plants are already highly efficient at using and acquiring P<sub>i</sub>. In the short term, as P demand for biomass increases due to the CO2 fertilization effect, plants may also shift C investments to other Pi-use and -acquisition strategies that may have a better cost-to-benefit ratio (e.g. DeLucia et al., 1997). In the long term, eCO2 may induce gradual changes in species composition by favoring species that are better adapted in terms of P-use and -acquisition efficiency (see Turner, 2008; Turner et al., 2018; Esquivel-Muelbert et al., 2019).

The overall response of the forest to increased P demand for biomass under eCO<sub>2</sub> will also depend on other factors, such as the interplay with soil microorganisms (Box 1) and possible climatic changes, namely warming, increasing drought events and changes in precipitation patterns, which may affect how plants invest their resources and may affect the P-use and -acquisition strategies themselves (Ho et al., 2005). Moreover, increased resource investments by plants in the different strategies in response to eCO<sub>2</sub> are expected to have differential effects on C cycling. For instance, increased leaf Pi-resorption proficiency inevitably leads to lower quality leaf litter and may decrease decomposition rates (Jin et al., 2015; Sayer et al., 2020). Investments in belowground Pi acquisition will increase C fluxes to the soil, although the fate and longterm stability of newly input C are uncertain under eCO<sub>2</sub> and the accompanying climate changes (Cleveland et al., 2010; Terrer et al., 2021).

### VI. Ways forward

As a way forward, we suggest further utilizing and extending the large network of Amazonian forest inventory sites (Malhi *et al.*, 2002; Quesada *et al.*, 2011; Brienen *et al.*, 2015) by selecting a subset of sites along a representative soil P gradient that minimizes the confounding effects of climate, biogeographic regions or topography. Although measurements at the species level are highly desirable, they are notoriously challenging for belowground traits in diverse tropical forests. We argue that identifying patterns in traits and indicators of plant P-use and -acquisition strategies at the community level, along with the use of permanent measurement sites and standardized methodologies, may allow estimates of community investment in different strategies and evaluate and adapt our conceptual framework (Fig. 3). Repeated measurements over the year are additionally encouraged to identify temporal patterns in plant P-use and -acquisition strategies. Following our

conceptual framework (Fig. 3), we propose the measurement of different traits and indicators to experimentally test the predicted patterns in plant P-use and -acquisition strategies, which we considered feasible under coordinated efforts among the scientific community over time:

(1) Leaf  $P_i$  resorption is relatively more important for plants in low-P soils – we recommend measuring P resorption from different chemical fractions in senesced leaves to identify the underlying mechanisms allowing high P resorption proficiency in Amazonia and the role of leaf  $P_i$  resorption efficiency (e.g. Hidaka & Kitayama, 2011; Tsujii *et al.*, 2017). This may allow an estimation of where plants may invest proportionally more resources in this strategy. Additionally, the measurement of  $P_i$  resorption from other organs would significantly improve our understanding of plant P use.

(2) Fine-root  $P_i$  foraging is relatively more important in soils with high  $P_i$  availability – classifying roots according to their functions (McCormack *et al.*, 2015) and measuring root morphological and architectural traits and dynamics (e.g. root elongation rate and turnover), as well as accounting for the effects of other edaphic factors, can significantly improve our understanding of fine-root  $P_i$ uptake and functioning (Freschet *et al.*, 2021). These measurements, in addition to the measurements suggested for the other strategies, may allow us to identify possible areas in Amazonia where roots primarily forage for  $P_i$ .

(3) AMF symbioses are relatively more important in soils with intermediate soil  $P_i$  availability – measurements of intraradical colonization are often preferred in most studies, but the extent of the AMF hyphal network in the soil may be an essential complementary measure of  $P_i$  acquisition capacity and plant C investments. We recommend measuring AMF intraradical and extraradical indicators (hyphal length and biomass) while also analyzing root traits promoting AMF colonization, as well as their dynamics in space and time (i.e. production and turnover rates).

(4) Root phosphatase exudation is relatively more important in soils with low  $P_i$  availability and high substrate availability – we recommend measuring and extending our knowledge on potential activity rates of root-associated PMEs and phytases complementary to root traits fostering these strategies, as well as a better understanding of enzyme production and turnover rates and their efficacy *in situ* is needed for model development and benchmarking (see Wallenstein & Weintraub, 2008; Razavi *et al.*, 2019).

(5) Fast rates of root LMWOA exudation are relatively more important where most P is in less available forms – we recommend quantifying root exudation rates while also analyzing associated root traits (e.g. Phillips *et al.*, 2008; Aoki *et al.*, 2012). We additionally recommend testing the use of leaf manganese (Mn) concentrations to serve as a proxy for LMWOA exudation in Amazonia, as they are easier to measure and have the potential to uncover large patterns in plant investment in this strategy (see Lambers *et al.*, 2015; 2021). Additionally, investigating LMWOA effectiveness in mobilizing P and residence time in Amazonian soils would significantly improve our understanding of their role in the rhizosphere.

By measuring the same traits and indicators previously suggested, nutrient manipulation and  $eCO_2$  experiments (FACE or open-top chambers) provide the opportunity to advance our

understanding by revealing species plasticity to adjust Pi-use and -acquisition strategies. Likewise, process-based model efforts may find value in testing and challenging our current understanding of the role of the strategies discussed (Box 3). The variation in plant investments in different strategies and their relative importance in supplying plant P<sub>i</sub> needs can be investigated using different modeling approaches, for instance by incorporating different mechanistic possibilities into a modular framework and constraining the consequences of uncertain mechanisms on plant function and C storage. Moreover, the current challenge lies in overcoming the low agreement between actually measurable entities and model parameters. Coordinated efforts among field studies, experiments and modeling need to be used to address the challenge of mechanistically representing P and eCO<sub>2</sub> interactions (Reed et al., 2015; Hofhansl et al., 2016). Likewise, the impacts of climatic changes and plant-microbial interactions (not focused on here) need to be assessed to reliably project the response of Amazonia to global change so that suitable management actions can be taken.

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## ORCID

Katrin Fleischer https://orcid.org/0000-0002-9093-9526 Lucia Fuchslueger https://orcid.org/0000-0002-9615-4439 Laynara F. Lugli https://orcid.org/0000-0001-8404-4841 Anja Rammig https://orcid.org/0000-0001-5425-8718 Tatiana Reichert https://orcid.org/0000-0002-6300-2084

## References

- Achat DL, Augusto L, Bakker MR, Gallet-Budynek A, Morel C. 2012. Microbial processes controlling P availability in forest spodosols as affected by soil depth and soil properties. *Soil Biology and Biochemistry* 44: 39–48.
- Achat DL, Morel C, Bakker MR, Augusto L, Pellerin S, Gallet-Budynek A, Gonzalez M. 2010. Assessing turnover of microbial biomass phosphorus: combination of an isotopic dilution method with a mass balance model. *Soil Biology and Biochemistry* 42: 2231–2240.

- Addo-Danso SD, Defrenne CE, McCormack ML, Ostonen I, Addo-Danso A, Foli EG, Borden KA, Isaac ME, Prescott CE. 2020. Fine-root morphological trait variation in tropical forest ecosystems: an evidence synthesis. *Plant Ecology* 221: 1– 13.
- Adeleke R, Nwangburuka C, Oboirien B. 2017. Origins, roles and fate of organic acids in soils: a review. *South African Journal of Botany* 108: 393–406.
- Alberton O, Kuyper TW, Gorissen A. 2005. Taking mycocentrism seriously: mycorrhizal fungal and plant responses to elevated CO<sub>2</sub>. *New Phytologist* 167: 859–868.
- Allen MF. 2001. Modeling arbuscular mycorrhizal infection: is % infection an appropriate variable? *Mycorrhiza* 10: 255–258.
- Allen K, Fisher JB, Phillips RP, Powers JS, Brzostek ER. 2020. Modeling the carbon cost of plant nitrogen and phosphorus uptake across temperate and tropical forests. *Frontiers in Forests and Global Change* **3**. doi: 10.3389/ffgc.2020. 00043.
- Allison SD, Weintraub MN, Gartner TB, Waldrop MP. 2010. Evolutionaryeconomic principles as regulators of soil enzyme production and ecosystem function. In: Shukla G, Varma A, eds. *Soil enzymology*. Berlin/Heidelberg, Germany: Springer, 229–243.
- Alvarez-Clare S, Mack MC. 2015. Do foliar, litter, and root nitrogen and phosphorus concentrations reflect nutrient limitation in a lowland tropical wet forest? *PLoS ONE* 10: e0123796.
- Andrade F, Mendonça E, Alvarez V, Novais R. 2003. Adição de ácidos orgânicos e húmicos em Latossolos e adsorção de fosfato. *Revista Brasileira de Ciência do Solo* 27: 1003–1011.
- Andrino A, Boy J, Mikutta R, Sauheitl L, Guggenberger G. 2019. Carbon investment required for the mobilization of inorganic and organic phosphorus bound to goethite by an arbuscular mycorrhiza (*Solanum lycopersicum* x *Rhizophagus irregularis*). *Frontiers in Environmental Science* 7: 26.
- Andrino A, Guggenberger G, Sauheitl L, Burkart S, Boy J. 2021. Carbon investment into mobilization of mineral and organic phosphorus by arbuscular mycorrhiza. *Biology and Fertility of Soils* 57: 47–64.
- Aoki M, Fujii K, Kitayama K. 2012. Environmental control of root exudation of low-molecular weight organic acids in tropical rainforests. *Ecosystems* 15: 1194– 1203.
- Aragão LEOC, Malhi Y, Metcalfe DB, Silva-Espejo JE, Jiménez E, Navarrete D, Almeida S, Costa ACL, Salinas N, Phillips OL *et al.* 2009. Above-and belowground net primary productivity across ten Amazonian forests on contrasting soils. *Biogeosciences Discuss* 6: 2759–2778.
- Arnone JA III. 1997. Temporal responses of community fine root populations to long-term elevated atmospheric CO<sub>2</sub> and soil nutrient patches in model tropical ecosystems. *Acta Oecologica* 18: 367–376.
- **Balser T, Treseder K, Ekenler M. 2005**. Using lipid analysis and hyphal length to quantify AM and saprotrophic fungal abundance along a soil chronosequence. *Soil Biology and Biochemistry* **37**: 601–604.
- Bergmann J, Weigelt A, van der Plas F, Laughlin DC, Kuyper TW, Guerrero-Ramirez N, Valverde-Barrantes OJ, Bruelheide H, Freschet GT, Iversen CM *et al.* 2020. The fungal collaboration gradient dominates the root economics space in plants. *Science Advances* 6: eaba3756. doi: 10.1126/sciadv.aba3756.
- Bloom AJ, Chapin FS III, Mooney HA. 1985. Resource limitation in plants-an economic analogy. *Annual Review of Ecology and Systematics* 16: 363–392.
- Bolan N, Robson A, Barrow N. 1987. Effects of vesicular-arbuscular mycorrhiza on the availability of iron phosphates to plants. *Plant and Soil* 99: 401–410.
- Brando PM, Nepstad DC, Davidson EA, Trumbore SE, Ray D, Camargo P. 2008. Drought effects on litterfall, wood production and belowground carbon cycling in an Amazon forest: results of a throughfall reduction experiment. *Philosophical Transactions of the Royal Society B: Biological Sciences* 363: 1839–1848.
- Brienen RJW, Phillips OL, Feldpausch TR, Gloor E, Baker TR, Lloyd J, Lopez-Gonzalez G, Monteagudo-Mendoza A, Malhi Y, Lewis SL *et al.* 2015. Long-term decline of the Amazon carbon sink. *Nature* 519: 344–348.
- Brundrett MC, Tedersoo L. 2018. Evolutionary history of mycorrhizal symbioses and global host plant diversity. *New Phytologist* 220: 1108–1115.
- Cabugao KG, Timm CM, Carrell AA, Childs J, Lu T-YS, Pelletier DA, Weston DJ, Norby RJ. 2017. Root and rhizosphere bacterial phosphatase activity varies with tree species and soil phosphorus availability in Puerto Rico tropical forest. *Frontiers in Plant Science* **8**: 1834.

Cabugao KG, Yaffar D, Stenson N, Childs J, Phillips J, Mayes MA, Yang X, Weston DJ, Norby RJ. 2021. Bringing function to structure: root–soil interactions shaping phosphatase activity throughout a soil profile in Puerto Rico. *Ecology and Evolution* 11: 1150–1164.

Camenzind T, Hempel S, Homeier J, Horn S, Velescu A, Wilcke W, Rillig MC. 2014. Nitrogen and phosphorus additions impact arbuscular mycorrhizal abundance and molecular diversity in a tropical montane forest. *Global Change Biology* 20: 3646–3659.

Camenzind T, Homeier J, Dietrich K, Hempel S, Hertel D, Krohn A, Leuschner C, Oelmann Y, Olsson PA, Suárez JP *et al.* 2016. Opposing effects of nitrogen versus phosphorus additions on mycorrhizal fungal abundance along an elevational gradient in tropical montane forests. *Soil Biology and Biochemistry* 94: 37–47.

Campbell C, Sage R. 2002. Interactions between atmospheric CO<sub>2</sub> concentration and phosphorus nutrition on the formation of proteoid roots in white lupin (*Lupinus albus* L.). *Plant, Cell & Environment* 25: 1051–1059.

Canarini A, Kaiser C, Merchant A, Richter A, Wanek W. 2019. Root exudation of primary metabolites: mechanisms and their roles in plant responses to environmental stimuli. *Frontiers in Plant Science* 10: 157.

Chapin FS III. 1980. The mineral nutrition of wild plants. *Annual Review of Ecology* and Systematics 11: 233–260.

Cleveland CC, Houlton BZ, Smith WK, Marklein AR, Reed SC, Parton W, Del Grosso SJ, Running SW. 2013. Patterns of new versus recycled primary production in the terrestrial biosphere. *Proceedings of the National Academy of Sciences, USA* 110: 12733–12737.

Cleveland CC, Wieder WR, Reed SC, Townsend AR. 2010. Experimental drought in a tropical rain forest increases soil carbon dioxide losses to the atmosphere. *Ecology* 91: 2313–2323.

Condit R, Engelbrecht BM, Pino D, Pérez R, Turner BL. 2013. Species distributions in response to individual soil nutrients and seasonal drought across a community of tropical trees. *Proceedings of the National Academy of Sciences, USA* 110: 5064–5068.

Cordeiro AL, Norby RJ, Andersen KM, Valverde-Barrantes O, Fuchslueger L, Oblitas E, Hartley IP, Iversen CM, Gonçalves NB, Takeshi B *et al.* 2020. Fineroot dynamics vary with soil depth and precipitation in a low-nutrient tropical forest in the Central Amazonia. *Plant-Environment Interactions* 1: 3–16.

Corrales A, Henkel TW, Smith ME. 2018. Ectomycorrhizal associations in the tropics–biogeography, diversity patterns and ecosystem roles. *New Phytologist* 220: 1076–1091.

Courty PE, Doidy J, Garcia K, Wipf D, Zimmermann SD. 2016. The transportome of mycorrhizal systems. In: Martin F, ed. *Molecular mycorrhizal symbiosis*. Hoboken, NJ, USA: John Wiley & Sons, 239–256.

Crous KY, Wujeska-Klause A, Jiang M, Medlyn BE, Ellsworth DS. 2019. Nitrogen and phosphorus retranslocation of leaves and stemwood in a mature eucalyptus forest exposed to 5 years of elevated CO<sub>2</sub>. *Frontiers in Plant Science* **10**: 664.

Dantas M, Phillipson J. 1989. Litterfall and litter nutrient content in primary and secondary Amazonian 'terra firme' rain forest. *Journal of Tropical Ecology* 5: 27–36.

Darch T, Blackwell MS, Chadwick D, Haygarth PM, Hawkins JM, Turner BL. 2016. Assessment of bioavailable organic phosphorus in tropical forest soils by organic acid extraction and phosphatase hydrolysis. *Geoderma* 284: 93–102.

**DeLucia E, Callaway R, Thomas E, Schlesinger W. 1997**. Mechanisms of phosphorus acquisition for ponderosa pine seedlings under high CO<sub>2</sub> and temperature. *Annals of Botany* **79**: 111–120.

Deubel A, Merbach W. 2005. Influence of microorganisms on phosphorus bioavailability in soils. In: Varma A, Buscot F, eds. *Microorganisms in soils: roles in* genesis and functions. Berlin, Heidelberg: Springer, 177–191.

**Dong J, Hunt J, Delhaize E, Zheng SJ, Jin CW, Tang C. 2021**. Impacts of elevated CO<sub>2</sub> on plant resistance to nutrient deficiency and toxic ions via root exudates: a review. *Science of the Total Environment* **754**: 142434.

Dong Y, Wang Z, Sun H, Yang W, Xu H. 2018. The response patterns of arbuscular mycorrhizal and ectomycorrhizal symbionts under elevated CO<sub>2</sub>: a meta-analysis. *Frontiers in Microbiology* 9: 1248.

Dray MW, Crowther TW, Thomas SM, A'Bear AD, Godbold DL, Ormerod SJ, Hartley SE, Jones TH. 2014. Effects of elevated CO<sub>2</sub> on litter chemistry and subsequent invertebrate detritivore feeding responses. *PLoS ONE* 9: e86246.

- Esquivel-Muelbert A, Baker TR, Dexter KG, Lewis SL, Brienen RJW, Feldpausch TR, Lloyd J, Monteagudo-Mendoza A, Arroyo L, Álvarez-Dávila E *et al.* 2019. Compositional response of Amazon forests to climate change. *Global Change Biology* 25: 39–56.
- Falkowski PG, Fenchel T, Delong EF. 2008. The microbial engines that drive Earth's biogeochemical cycles. *Science* **320**: 1034–1039.

Finzi AC, Allen AS, DeLucia EH, Ellsworth DS, Schlesinger WH. 2001. Forest litter production, chemistry, and decomposition following two years of free-air CO<sub>2</sub> enrichment. *Ecology* 82: 470–484.

Fleischer K, Rammig A, De Kauwe MG, Walker AP, Domingues TF, Fuchslueger L, Garcia S, Goll DS, Grandis A, Jiang M *et al.* 2019. Amazon forest response to CO<sub>2</sub> fertilization dependent on plant phosphorus acquisition. *Nature Geoscience* 12: 736–741.

Freschet GT, Roumet C, Comas LH, Weemstra M, Bengough AG, Rewald B, Bardgett RD, De Deyn GB, Johnson D, Klimešová J *et al.* 2021. Root traits as drivers of plant and ecosystem functioning: current understanding, pitfalls and future research needs. *New Phytologist* 232: 1123–1158.

Frost PC, Tuchman NC. 2005. Nutrient release rates and ratios by two stream detritivores fed leaf litter grown under elevated atmospheric CO<sub>2</sub>. Archiv für Hydrobiologie 163: 463–477.

Fujii K, Aoki M, Kitayama K. 2012. Biodegradation of low molecular weight organic acids in rhizosphere soils from a tropical montane rain forest. *Soil Biology* and Biochemistry 47: 142–148.

Fujii K, Hayakawa C, Sukartiningsih. 2021. Root exudation and biodegradation of organic acids in a tropical forest soil under dipterocarp and pioneer trees. *Plant and Soil* 469: 213–226.

Gerke J. 2015a. The acquisition of phosphate by higher plants: effect of carboxylate release by the roots. A critical review. *Journal of Plant Nutrition and Soil Science* 178: 351–364.

Gerke J. 2015b. Phytate (inositol hexakisphosphate) in soil and phosphate acquisition from inositol phosphates by higher plants. A review. *Plants* 4: 253–266.

Girardin C, Aragão L, Malhi Y, Huaraca Huasco W, Metcalfe D, Durand L, Mamani M, Silva-Espejo J, Whittaker R. 2013. Fine root dynamics along an elevational gradient in tropical Amazonian and Andean forests. *Global Biogeochemical Cycles* 27: 252–264.

Goll DS, Vuichard N, Maignan F, Jornet-Puig A, Sardans J, Violette A, Peng S, Sun Y, Kvakic M, Guimberteau M et al. 2017. A representation of the phosphorus cycle for ORCHIDEE (revision 4520). Geoscientific Model Development 10: 3745–3770.

Guilbeault-Mayers X, Turner BL, Laliberté E. 2020. Greater root phosphatase activity of tropical trees at low phosphorus despite strong variation among species. *Ecology* **101**: e03090.

Hättenschwiler S, Aeschlimann B, Coûteaux MM, Roy J, Bonal D. 2008. High variation in foliage and leaf litter chemistry among 45 tree species of a neotropical rainforest community. *New Phytologist* **179**: 165–175.

Hayes P, Turner BL, Lambers H, Laliberté E. 2014. Foliar nutrient concentrations and resorption efficiency in plants of contrasting nutrientacquisition strategies along a 2-million-year dune chronosequence. *Journal of Ecology* 102: 396–410.

He H, Jansson P-E, Gärdenäs AI. 2021. COUPMODEL (v.6.0): an ecosystem model for coupled phosphorus, nitrogen, and carbon dynamics–evaluated against empirical data from a climatic and fertility gradient in Sweden. *Geoscientific Model Development* 14: 735–761.

Heineman KD, Turner BL, Dalling JW. 2016. Variation in wood nutrients along a tropical soil fertility gradient. *New Phytologist* 211: 440–454.

Hidaka A, Kitayama K. 2011. Allocation of foliar phosphorus fractions and leaf traits of tropical tree species in response to decreased soil phosphorus availability on Mount Kinabalu, Borneo. *Journal of Ecology* 99: 849–857.

Ho MD, Rosas JC, Brown KM, Lynch JP. 2005. Root architectural tradeoffs for water and phosphorus acquisition. *Functional Plant Biology* 32: 737–748.

Hodge A. 2004. The plastic plant: root responses to heterogeneous supplies of nutrients. *New Phytologist* 162: 9–24.

Hofhansl F, Andersen KM, Fleischer K, Fuchslueger L, Rammig A, Schaap KJ, Valverde-Barrantes OJ, Lapola DM. 2016. Amazon forest ecosystem responses to

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elevated atmospheric  $CO_2$  and alterations in nutrient availability: filling the gaps with model-experiment integration. *Frontiers in Earth Science* 4: 19.

- Holdaway RJ, Richardson SJ, Dickie IA, Peltzer DA, Coomes DA. 2011. Speciesand community-level patterns in fine root traits along a 120 000-year soil chronosequence in temperate rain forest. *Journal of Ecology* **99**: 954–963.
- Homeier J, Hertel D, Camenzind T, Cumbicus NL, Maraun M, Martinson GO, Poma LN, Rillig MC, Sandmann D, Scheu S *et al.* 2012. Tropical Andean forests are highly susceptible to nutrient inputs—rapid effects of experimental N and P addition to an Ecuadorian montane forest. *PLoS ONE7*: e47128.
- Hou E, Tang S, Chen C, Kuang Y, Lu X, Heenan M, Wen D. 2018. Solubility of phosphorus in subtropical forest soils as influenced by low-molecular organic acids and key soil properties. *Geoderma* **313**: 172–180.
- Housman D, Killingbeck K, Evans RD, Charlet T, Smith S. 2012. Foliar nutrient resorption in two Mojave Desert shrubs exposed to free-air CO<sub>2</sub> enrichment (FACE). *Journal of Arid Environments* 78: 26–32.
- Hubau W, Lewis SL, Phillips OL, Affum-Baffoe K, Beeckman H, Cuní-Sanchez A, Daniels AK, Ewango CE, Fauset S, Mukinzi JM. 2020. Asynchronous carbon sink saturation in African and Amazonian tropical forests. *Nature* 579: 80–87.
- Insam H, Bååth E, Berreck M, Frostegård Å, Gerzabek M, Kraft A, Schinner F, Schweiger P, Tschuggnall G. 1999. Responses of the soil microbiota to elevated CO<sub>2</sub> in an artificial tropical ecosystem. *Journal of Microbiological Methods* 36: 45– 54.
- Jakobsen I, Abbott L, Robson A. 1992. External hyphae of vesicular-arbuscular mycorrhizal fungi associated with *Trifolium subterraneum* L. 1. Spread of hyphae and phosphorus inflow into roots. *New Phytologist* **120**: 371–380.
- Jarosch KA, Kandeler E, Frossard E, Bünemann EK. 2019. Is the enzymatic hydrolysis of soil organic phosphorus compounds limited by enzyme or substrate availability? *Soil Biology and Biochemistry* 139. doi: 10.1016/j.soilbio.2019. 107628.
- Jiang F, Zhang L, Zhou J, George TS, Feng G. 2021. Arbuscular mycorrhizal fungi enhance mineralization of organic phosphorus (P) by carrying bacteria along their extraradical hyphae. *New Phytologist* 230: 304–315.
- Jiang M, Caldararu S, Zhang H, Fleischer K, Crous KY, Yang J, De Kauwe MG, Ellsworth DS, Reich PB, Tissue DT. 2020a. Low phosphorus supply constrains plant responses to elevated CO<sub>2</sub>: a meta-analysis. *Global Change Biology* 26: 5856–5873.
- Jiang M, Medlyn BE, Drake JE, Duursma RA, Anderson IC, Barton CV, Boer MM, Carrillo Y, Castañeda-Gómez L, Collins L. 2020b. The fate of carbon in a mature forest under carbon dioxide enrichment. *Nature* 580: 227–231.
- Jin J, Tang C, Robertson A, Franks AE, Armstrong R, Sale P. 2014. Increased microbial activity contributes to phosphorus immobilization in the rhizosphere of wheat under elevated CO<sub>2</sub>. *Soil Biology and Biochemistry* 75: 292–299.
- Jin J, Tang C, Sale P. 2015. The impact of elevated carbon dioxide on the phosphorus nutrition of plants: a review. *Annals of Botany* 116: 987–999.
- Johnson NC. 2010. Resource stoichiometry elucidates the structure and function of arbuscular mycorrhizas across scales. *New Phytologist* 185: 631–647.
- Jones DL. 1998. Organic acids in the rhizosphere–a critical review. *Plant and Soil* 205: 25–44.
- Jordan CF, Escalante G. 1980. Root productivity in an Amazonian rain forest. *Ecology* 61: 14–18.
- Khan MS, Zaidi A, Ahmad E. 2014. Mechanism of phosphate solubilization and physiological functions of phosphate-solubilizing microorganisms. In: Khan MS, Zaidi A, Musarrat J, eds. *Phosphate solubilizing microorganisms: principles and application of microphos technology.* Cham, CH: Springer International, 31–62.
- Killingbeck KT. 1996. Nutrients in senesced leaves: keys to the search for potential resorption and resorption proficiency. *Ecology* 77: 1716–1727.
- Killingbeck KT. 2004. Nutrient resorption. In: Noodén LD, ed. *Plant cell death processes*. San Diego, CA, USA: Academic Press, 215–226.
- Kitayama K. 2013. The activities of soil and root acid phosphatase in the nine tropical rain forests that differ in phosphorus availability on Mount Kinabalu, Borneo. *Plant and Soil* 367: 215–224.
- Kobae Y. 2019. Dynamic phosphate uptake in arbuscular mycorrhizal roots under field conditions. *Frontiers in Environmental Science* 6: 159.
- Kochsiek A, Tan S, Russo SE. 2013. Fine root dynamics in relation to nutrients in oligotrophic Bornean rain forest soils. *Plant Ecology* 214: 869–882.

- Körner C, Arnone JA. 1992. Responses to elevated carbon dioxide in artificial tropical ecosystems. *Science* 257: 1672–1675.
- Kramer-Walter KR, Bellingham PJ, Millar TR, Smissen RD, Richardson SJ, Laughlin DC. 2016. Root traits are multidimensional: specific root length is independent from root tissue density and the plant economic spectrum. *Journal of Ecology* 104: 1299–1310.
- Kramer-Walter KR, Laughlin DC. 2017. Root nutrient concentration and biomass allocation are more plastic than morphological traits in response to nutrient limitation. *Plant and Soil* 416: 539–550.
- Lambers H. 2022. Phosphorus acquisition and utilization in plants. Annual Review of Plant Biology 73. doi: 10.1146/annurev-arplant-102720-125738.
- Lambers H, Hayes PE, Laliberté E, Oliveira RS, Turner BL. 2015. Leaf manganese accumulation and phosphorus-acquisition efficiency. *Trends in Plant Science* 20: 83–90.
- Lambers H, Raven JA, Shaver GR, Smith SE. 2008. Plant nutrientacquisition strategies change with soil age. *Trends in Ecology & Evolution* 23: 95–103.
- Lambers H, Shane MW, Cramer MD, Pearse SJ, Veneklaas EJ. 2006. Root structure and functioning for efficient acquisition of phosphorus: matching morphological and physiological traits. *Annals of Botany* **98**: 693–713.
- Lambers H, Wright IJ, Guilherme Pereira C, Bellingham PJ, Bentley LP, Boonman A, Cernusak LA, Foulds W, Gleason SM, Gray EF et al. 2021. Leaf manganese concentrations as a tool to assess belowground plant functioning in phosphorus-impoverished environments. *Plant and Soil* 461: 43–61.
- Lips JM, Duivenvoorden JF. 1996. Fine litter input to terrestrial humus forms in Colombian Amazonia. *Oecologia* 108: 138–150.
- Liu B, Li H, Zhu B, Koide RT, Eissenstat DM, Guo D. 2015. Complementarity in nutrient foraging strategies of absorptive fine roots and arbuscular mycorrhizal fungi across 14 coexisting subtropical tree species. *New Phytologist* 208: 125–136.
- Lloyd J, Bird M, Veenendaal E, Kruijt B. 2001. Should phosphorus availability be constraining moist tropical forest responses to increasing CO<sub>2</sub> concentrations? In: Schulze M, Heimann S, Holland E, Lloyd J, Prentice IC, Schimel D, eds. *Global biogeochemical cycles in the climate system*. San Diego, CA, USA: Elsevier, 95–114.
- Lugli LF, Andersen KM, Aragão LEOC, Cordeiro AL, Cunha HFV, Fuchslueger L, Meir P, Mercado LM, Oblitas E, Quesada CA et al. 2020. Multiple phosphorus acquisition strategies adopted by fine roots in low-fertility soils in Central Amazonia. *Plant and Soil* 450: 49–63.
- Lugli LF, Rosa JS, Andersen KM, Di Ponzio R, Almeida RV, Pires M, Cordeiro AL, Cunha HFV, Martins NP, Assis RL et al. 2021. Rapid responses of root traits and productivity to phosphorus and cation additions in a tropical lowland forest in Amazonia. New Phytologist 230: 116–128.
- Lynch JP, Brown KM. 2001. Topsoil foraging an architectural adaptation of plants to low phosphorus availability. *Plant and Soil* 237: 225–237.
- Lynch JP, Brown KM. 2008. Root strategies for phosphorus acquisition. In: White PJ, Hammond JP, eds. *The ecophysiology of plant-phosphorus interactions*. Dordrecht, the Netherlands: Springer, 83–116.
- Lynch JP, Ho MD, phosphorus L. 2005. Rhizoeconomics: carbon costs of phosphorus acquisition. *Plant and Soil* 269: 45–56.
- Malhi Y, Phillips OL, Lloyd J, Baker T, Wright J, Almeida S, Arroyo L, Frederiksen T, Grace J, Higuchi N *et al.* 2002. An international network to monitor the structure, composition and dynamics of Amazonian forests (RAINFOR). *Journal* of Vegetation Science 13: 439–450.
- Marklein AR, Houlton BZ. 2012. Nitrogen inputs accelerate phosphorus cycling rates across a wide variety of terrestrial ecosystems. *New Phytologist* 193: 696–704.
- Martins NP, Fuchslueger L, Fleischer K, Andersen KM, Assis RL, Baccaro FB, Camargo PB, Cordeiro AL, Grandis A, Hartley IP *et al.* 2021. Fine roots stimulate nutrient release during early stages of leaf litter decomposition in a Central Amazon rainforest. *Plant and Soil* 469: 287–303.
- Mayor JR, Wright SJ, Turner BL. 2014. Species-specific responses of foliar nutrients to long-term nitrogen and phosphorus additions in a lowland tropical forest. *Journal of Ecology* 102: 36–44.
- McCormack ML, Dickie IA, Eissenstat DM, Fahey TJ, Fernandez CW, Guo D, Helmisaari H-S, Hobbie EA, Iversen CM, Jackson RB *et al.* 2015. Redefining fine roots improves understanding of below-ground contributions to terrestrial biosphere processes. *New Phytologist* 207: 505–518.

McCormack ML, Iversen CM. 2019. Physical and functional constraints on viable belowground acquisition strategies. *Frontiers in Plant Science* 10: 1215.

McGill W, Cole C. 1981. Comparative aspects of cycling of organic C, N, S and P through soil organic matter. *Geoderma* 26: 267–286.

Medina E, Klinge H, Jordan C, Herrera R. 1980. Soil respiration in Amazonian rain forests in the Rio Negro Basin. *Flora* 170: 240–250.

Metcalfe DB, Meir P, Aragão LEOC, da Costa ACL, Braga AP, Gonçalves PHL, de Athaydes Silva Junior J, de Almeida SS, Dawson LA, Malhi Y *et al.* 2008. The effects of water availability on root growth and morphology in an Amazon rainforest. *Plant and Soil* 311: 189–199.

Mirmanto E, Proctor J, Green J, Nagy L, Suriantata. 1999. Effects of nitrogen and phosphorus fertilization in a lowland evergreen rainforest. *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences* 354: 1825–1829.

Mooshammer M, Wanek W, Zechmeister-Boltenstern S, Richter AA. 2014. Stoichiometric imbalances between terrestrial decomposer communities and their resources: mechanisms and implications of microbial adaptations to their resources. *Frontiers in Microbiology* 5: 22.

Nagy R, Drissner D, Amrhein N, Jakobsen I, Bucher M. 2009. Mycorrhizal phosphate uptake pathway in tomato is phosphorus-repressible and transcriptionally regulated. *New Phytologist* 181: 950–959.

Nannipieri P, Giagnoni L, Landi L, Renella G. 2011. Role of phosphatase enzymes in soil. In: Bünemann EK, Oberson A, Frossard E, eds. *Phosphorus in action*. Berlin, Heidelberg, Germany: Springer, 215–243.

Nannipieri P, Trasar-Cepeda C, Dick RP. 2018. Soil enzyme activity: a brief history and biochemistry as a basis for appropriate interpretations and meta-analysis. *Biology and Fertility of Soils* 54: 11–19.

Nasto MK, Alvarez-Clare S, Lekberg Y, Sullivan BW, Townsend AR, Cleveland CC. 2014. Interactions among nitrogen fixation and soil phosphorus acquisition strategies in lowland tropical rain forests. *Ecology Letters* 17: 1282–1289.

Nasto MK, Winter K, Turner BL, Cleveland CC. 2019. Nutrient acquisition strategies augment growth in tropical N<sub>2</sub>-fixing trees in nutrient-poor soil and under elevated CO<sub>2</sub>. *Ecology* 100: e02646.

Nepstad DC, de Carvalho CR, Davidson EA, Jipp PH, Lefebvre PA, Negreiros GH, da Silva ED, Stone TA, Trumbore SE, Vieira S. 1994. The role of deep roots in the hydrological and carbon cycles of Amazonian forests and pastures. *Nature* 372: 666–669.

Nie M, Lu M, Bell J, Raut S, Pendall E. 2013. Altered root traits due to elevated CO<sub>2</sub>: a meta-analysis. *Global Ecology and Biogeography* 22: 1095–1105.

Norisada M, Motoshige T, Kojima K, Tange T. 2006. Effects of phosphate supply and elevated CO<sub>2</sub> on root acid phosphatase activity in *Pinus densiflora* seedlings. *Journal of Plant Nutrition and Soil Science* 169: 274–279.

Oburger E, Jones DL. 2018. Sampling root exudates-mission impossible? *Rhizosphere* 6: 116–133.

Ochoa-Hueso R, Hughes J, Delgado-Baquerizo M, Drake JE, Tjoelker MG, Piñeiro J, Power SA. 2017. Rhizosphere-driven increase in nitrogen and phosphorus availability under elevated atmospheric CO<sub>2</sub> in a mature *Eucalyptus* woodland. *Plant and Soil* 416: 283–295.

Olivares I, Svenning J-C, van Bodegom PM, Balslev H. 2015. Effects of warming and drought on the vegetation and plant diversity in the Amazon basin. *Botanical Review* 81: 42–69.

Oliveira RS, Galvão HC, de Campos MC, Eller CB, Pearse SJ, Lambers H. 2015. Mineral nutrition of campos rupestres plant species on contrasting nutrientimpoverished soil types. *New Phytologist* 205: 1183–1194.

Ostertag R. 2001. Effects of nitrogen and phosphorus availability on fine-root dynamics in Hawaiian montane forests. *Ecology* 82: 485–499.

Phillips OL, Aragão LEOC, Lewis SL, Fisher JB, Lloyd J, López-González G, Malhi Y, Monteagudo A, Peacock J, Quesada CA et al. 2009. Drought sensitivity of the Amazon rainforest. Science 323: 1344–1347.

Phillips RP, Erlitz Y, Bier R, Bernhardt ES. 2008. New approach for capturing soluble root exudates in forest soils. *Functional Ecology* 22: 990–999.

Plassard C, Louche J, Ali MA, Duchemin M, Legname E, Cloutier-Hurteau B. 2011. Diversity in phosphorus mobilisation and uptake in ectomycorrhizal fungi. *Annals of Forest Science* 68: 33–43.

Png GK, Turner BL, Albornoz FE, Hayes PE, Lambers H, Laliberté E. 2017. Greater root phosphatase activity in nitrogen-fixing rhizobial but not actinorhizal plants with declining phosphorus availability. *Journal of Ecology* **105**: 1246–1255.

Posada RH, Madriñan S, Rivera E-L. 2012. Relationships between the litter colonization by saprotrophic and arbuscular mycorrhizal fungi with depth in a tropical forest. *Fungal Biology* 116: 747–755.

Powers JS, Treseder KK, Lerdau MT. 2005. Fine roots, arbuscular mycorrhizal hyphae and soil nutrients in four neotropical rain forests: patterns across large geographic distances. *New Phytologist* 165: 913–921.

Quesada CA, Lloyd J, Schwarz M, Patiño S, Baker TR, Czimczik C, Fyllas NM, Martinelli L, Nardoto GB, Schmerler J et al. 2010. Variations in chemical and physical properties of Amazon forest soils in relation to their genesis. *Biogeosciences* 7: 1515–1541.

Quesada CA, Lloyd J, Anderson LO, Fyllas NM, Schwarz M, Czimczik CI. 2011. Soils of Amazonia with particular reference to the RAINFOR sites. *Biogeosciences* 8: 1415–1440.

Quesada CA, Phillips OL, Schwarz M, Czimczik CI, Baker TR, Patiño S, Fyllas NM, Hodnett MG, Herrera R, Almeida S *et al.* 2012. Basin-wide variations in Amazon forest structure and function are mediated by both soils and climate. *Biogeosciences* 9: 2203–2246.

Quintero-Vallejo E, Pena-Claros M, Bongers F, Toledo M, Poorter L. 2015. Effects of Amazonian Dark Earths on growth and leaf nutrient balance of tropical tree seedlings. *Plant and Soil* **396**: 241–255.

Raven JA, Lambers H, Smith SE, Westoby M. 2018. Costs of acquiring phosphorus by vascular land plants: patterns and implications for plant coexistence. *New Phytologist* 217: 1420–1427.

Razavi BS, Zhang X, Bilyera N, Guber A, Zarebanadkouki M. 2019. Soil zymography: Simple and reliable? Review of current knowledge and optimization of the method. *Rhizosphere* 11: 100161.

Reed SC, Yang X, Thornton PE. 2015. Incorporating phosphorus cycling into global modeling efforts: a worthwhile, tractable endeavor. *New Phytologist* 208: 324–329.

Richardson AE, Simpson RJ. 2011. Soil microorganisms mediating phosphorus availability update on microbial phosphorus. *Plant Physiology* 156: 989–996.

Richardson SJ, Peltzer DA, Allen RB, McGlone MS. 2005. Resorption proficiency along a chronosequence: responses among communities and within species. *Ecology* 86: 20–25.

Roelofs R, Rengel Z, Cawthray G, Dixon K, Lambers H. 2001. Exudation of carboxylates in Australian Proteaceae: chemical composition. *Plant, Cell & Environment* 24: 891–904.

Roy M, Schimann H, Braga-Neto R, Da Silva RA, Duque J, Frame D, Wartchow F, Neves MA. 2016. Diversity and distribution of ectomycorrhizal fungi from Amazonian lowland white-sand forests in Brazil and French Guiana. *Biotropica* 48: 90–100.

Ryan PR, Delhaize E, Jones DL. 2001. Function and mechanism of organic anion exudation from plant roots. *Annual Review of Plant Physiology and Plant Molecular Biology* 52: 527–560.

Saito K, Ezawa T. 2016. Phosphorus metabolism and transport in arbuscular mycorrhizal symbiosis. In: Martin F, ed. *Molecular mycorrhizal symbiosis*. Hoboken, NJ, USA: John Wiley & Sons, 197–216.

Sampaio G, Borma LS, Cardoso M, Alves LM, von Randow C, Rodriguez DA, Nobre CA, Alexandre FF. 2019. Assessing the possible impacts of a 4°C or higher warming in Amazonia. In: Nobre CA, Marengo JA, Soares RS, eds. *Climate change risks in Brazil.* Cham, Switzerland: Springer, 201–218.

Sánchez-Calderón L, Chacon-López A, Pérez-Torres C-A, Herrera-Estrella L. 2010. Phosphorus: plant strategies to cope with its scarcity. In: Hell R, Mendel R-R, eds. *Cell biology of metals and nutrients*. Berlin/Heidelberg, Germany: Springer, 173–198.

Sato T, Ezawa T, Cheng W, Tawaraya K. 2015. Release of acid phosphatase from extraradical hyphae of arbuscular mycorrhizal fungus Rhizophagus clarus. *Soil Science and Plant Nutrition* 61: 269–274.

Sayer EJ, Rodtassana C, Sheldrake M, Bréchet LM, Ashford OS, Lopez-Sangil L, Kerdraon-Byrne D, Castro B, Turner BL, Wright SJ. 2020. Revisiting nutrient cycling by litterfall—Insights from 15 years of litter manipulation in old-growth lowland tropical forest. In: Dumbrell AJ, Turner EC, Fayle TM, eds. Advances in Ecological Research, vol. 62. Oxford, UK: Academic Press, 173–223.

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Sheldrake M, Rosenstock NP, Mangan S, Revillini D, Sayer EJ, Olsson PA, Verbruggen E, Tanner EV, Turner BL, Wright SJ. 2018. Responses of arbuscular mycorrhizal fungi to long-term inorganic and organic nutrient addition in a lowland tropical forest. *ISME Journal* 12: 2433–2445.

Smith S, Smith F. 1990. Structure and function of the interfaces in biotrophic symbioses as they relate to nutrient transport. *New Phytologist* 114: 1–38.

Smith SE, Smith FA, Jakobsen I. 2004. Functional diversity in arbuscular mycorrhizal (AM) symbioses: the contribution of the mycorrhizal P uptake pathway is not correlated with mycorrhizal responses in growth or total P uptake. *New Phytologist* 162: 511–524.

Soong JL, Janssens IA, Grau O, Margalef O, Stahl C, Van Langenhove L, Urbina I, Chave J, Dourdain A, Ferry B et al. 2020. Soil properties explain tree growth and mortality, but not biomass, across phosphorus-depleted tropical forests. *Scientific Reports* 10: 1–13.

Spohn M, Ermak A, Kuzyakov Y. 2013. Microbial gross organic phosphorus mineralization can be stimulated by root exudates – a <sup>33</sup>P isotopic dilution study. *Soil Biology and Biochemistry* 65: 254–263.

Stark NM, Jordan CF. 1978. Nutrient retention by the root mat of an Amazonian rain forest. *Ecology* 59: 434–437.

Steidinger BS, Turner BL, Corrales A, Dalling JW. 2014. Variability in potential to exploit different soil organic phosphorus compounds among tropical montane tree species. *Functional Ecology* 29: 121–130.

Stonor RN, Smith SE, Manjarrez M, Facelli E, Smith FA. 2014. Mycorrhizal responses in wheat: shading decreases growth but does not lower the contribution of the fungal phosphate uptake pathway. *Mycorrhiza* 24: 465–472.

Stuart EK, Plett KL. 2020. Digging deeper: in search of the mechanisms of carbon and nitrogen exchange in ectomycorrhizal symbioses. *Frontiers in Plant Science* 10: 1658.

Tabatabai MA. 1994. Soil enzymes. *Methods of Soil Analysis: Part 2 Microbiological* and Biochemical Properties 5: 775–833.

Tang L, Han W, Chen Y, Fang J. 2013. Resorption proficiency and efficiency of leaf nutrients in woody plants in eastern China. *Journal of Plant Ecology* 6: 408–417.

Tedersoo L, Bahram M, Zobel M. 2020. How mycorrhizal associations drive plant population and community biology. *Science* 367: eaba1223.

Terrer C, Jackson RB, Prentice IC, Keenan TF, Kaiser C, Vicca S, Fisher JB, Reich PB, Stocker BD, Hungate BA *et al.* 2019. Nitrogen and phosphorus constrain the CO<sub>2</sub> fertilization of global plant biomass. *Nature Climate Change* 9: 684–689.

Terrer C, Phillips RP, Hungate BA, Rosende J, Pett-Ridge J, Craig ME, van Groenigen KJ, Keenan TF, Sulman BN, Stocker BD *et al.* 2021. A trade-off between plant and soil carbon storage under elevated CO<sub>2</sub>. *Nature* 591: 599–603.

Terrer C, Vicca S, Hungate BA, Phillips RP, Prentice IC. 2016. Mycorrhizal association as a primary control of the CO<sub>2</sub> fertilization effect. *Science* **353**: 72–74.

Teste FP, Laliberté E, Lambers H, Auer Y, Kramer S, Kandeler E. 2016. Mycorrhizal fungal biomass and scavenging declines in phosphorus-impoverished soils during ecosystem retrogression. *Soil Biology and Biochemistry* 92: 119–132.

Thum T, Caldararu S, Engel J, Kern M, Pallandt M, Schnur R, Yu L, Zaehle S. 2019. A new model of the coupled carbon, nitrogen, and phosphorus cycles in the terrestrial biosphere (QUINCY v.1.0; revision 1996). *Geoscientific Model Development* 12: 4781–4802.

Tian J, Liao H. 2018. The role of intracellular and secreted purple acid phosphatases in plant phosphorus scavenging and recycling. *Annual Plant Reviews* 265–287. doi: 10.1002/9781119312994.apr0525.

Tisserant B, Gianinazzi S, Gianinazzi-Pearson V. 1996. Relationships between lateral root order, arbuscular mycorrhiza development, and the physiological state of the symbiotic fungus in Platanus acerifolia. *Canadian Journal of Botany* 74: 1947–1955.

Treseder KK. 2004. A meta-analysis of mycorrhizal responses to nitrogen, phosphorus, and atmospheric CO<sub>2</sub> in field studies. *New Phytologist* 164: 347–355.

Treseder KK. 2013. The extent of mycorrhizal colonization of roots and its influence on plant growth and phosphorus content. *Plant and Soil* 371: 1–13.

Treseder KK, Allen MF. 2002. Direct nitrogen and phosphorus limitation of arbuscular mycorrhizal fungi: a model and field test. *New Phytologist* 155: 507–515.

**Treseder KK, Vitousek PM. 2001**. Effects of soil nutrient availability on investment in acquisition of N and P in Hawaiian rain forests. *Ecology* **82**: 946–954.

Trumbore S, Da Costa ES, Nepstad DC, Barbosa De Camargo P, Martinelli LA, Ray D, Restom T, Silver W. 2006. Dynamics of fine root carbon in Amazonian tropical ecosystems and the contribution of roots to soil respiration. *Global Change Biology* **12**: 217–229.

- Tsujii Y, Onoda Y, Kitayama K. 2017. Phosphorus and nitrogen resorption from different chemical fractions in senescing leaves of tropical tree species on Mount Kinabalu, Borneo. *Oecologia* 185: 171–180.
- Turner BL. 2008. Resource partitioning for soil phosphorus: a hypothesis. *Journal of Ecology* 96: 698–702.

Turner BL, Brenes-Arguedas T, Condit R. 2018. Pervasive phosphorus limitation of tree species but not communities in tropical forests. *Nature* 555: 367–370.

Turner BL, Condron LM. 2013. Pedogenesis, nutrient dynamics, and ecosystem development: the legacy of T.W. Walker and J.K. Syers. *Plant and Soil* 367: 1–10.

Turner BL, Lambers H, Condron LM, Cramer MD, Leake JR, Richardson AE, Smith SE. 2013. Soil microbial biomass and the fate of phosphorus during longterm ecosystem development. *Plant and Soil* 367: 225–234.

Umaña MN, Condit R, Pérez R, Turner BL, Wright SJ, Comita LS. 2021. Shifts in taxonomic and functional composition of trees along rainfall and phosphorus gradients in central Panama. *Journal of Ecology* **109**: 51–61.

Ushio M, Fujiki Y, Hidaka A, Kitayama K. 2015. Linkage of root physiology and morphology as an adaptation to soil phosphorus impoverishment in tropical montane forests. *Functional Ecology* 29: 1235–1245.

Valverde-Barrantes OJ, Freschet GT, Roumet C, Blackwood CB. 2017. A worldview of root traits: the influence of ancestry, growth form, climate and mycorrhizal association on the functional trait variation of fine-root tissues in seed plants. *New Phytologist* 215: 1562–1573.

Valverde-Barrantes OJ, Horning AL, Smemo KA, Blackwood CB. 2016. Phylogenetically structured traits in root systems influence arbuscular mycorrhizal colonization in woody angiosperms. *Plant and Soil* 404: 1–12.

Van Hees PA, Jones DL, Godbold DL. 2002. Biodegradation of low molecular weight organic acids in coniferous forest podzolic soils. *Soil Biology and Biochemistry* 34: 1261–1272.

Veneklaas EJ, Lambers H, Bragg J, Finnegan PM, Lovelock CE, Plaxton WC, Price CA, Scheible WR, Shane MW, White PJ. 2012. Opportunities for improving phosphorus-use efficiency in crop plants. *New Phytologist* 195: 306–320.

Vitousek PM. 1982. Nutrient cycling and nutrient use efficiency. American Naturalist 119: 553–572.

Vitousek PM. 1984. Litterfall, nutrient cycling, and nutrient limitation in tropical forests. *Ecology* 65: 285–298.

Vitousek PM. 1998. Foliar and litter nutrients, nutrient resorption, and decomposition in Hawaiian Metrosideros polymorpha. Ecosystems 1: 401–407.

Vitousek PM, Porder S, Houlton BZ, Chadwick OA. 2010. Terrestrial phosphorus limitation: mechanisms, implications, and nitrogen–phosphorus interactions. *Ecological Applications* 20: 5–15.

Walker AP, De Kauwe MG, Bastos A, Belmecheri S, Georgiou K, Keeling R, McMahon SM, Medlyn BE, Moore DJ, Norby RJ. 2021. Integrating the evidence for a terrestrial carbon sink caused by increasing atmospheric CO<sub>2</sub>. New Phytologist 229: 2413–2445.

Walker T, Syers JK. 1976. The fate of phosphorus during pedogenesis. *Geoderma* 15: 1–19.

Wallenstein MD, Weintraub MN. 2008. Emerging tools for measuring and modeling the in situ activity of soil extracellular enzymes. *Soil Biology and Biochemistry* 40: 2098–2106.

Wang L, Liu D. 2018. Functions and regulation of phosphate starvation-induced secreted acid phosphatases in higher plants. *Plant Science* 271: 108–116.

Wang W, Shi J, Xie Q, Jiang Y, Yu N, Wang E. 2017. Nutrient exchange and regulation in arbuscular mycorrhizal symbiosis. *Molecular Plant* 10: 1147–1158.

Wang Y, Lambers H. 2019. Root-released organic anions in response to low phosphorus availability: recent progress, challenges and future perspectives. *Plant* and Soil 447: 135–156.

Wasaki J, Rothe A, Kania A, Neumann G, Römheld V, Shinano T, Osaki M, Kandeler E. 2005. Root exudation, phosphorus acquisition, and microbial diversity in the rhizosphere of white lupine as affected by phosphorus supply and atmospheric carbon dioxide concentration. *Journal of Environmental Quality* 34: 2157–2166.

Wei L, Chen C, Xu Z. 2010. Citric acid enhances the mobilization of organic phosphorus in subtropical and tropical forest soils. *Biology and Fertility of Soils* 46: 765–769. Wurzburger N, Wright SJ. 2015. Fine-root responses to fertilization reveal multiple nutrient limitation in a lowland tropical forest. *Ecology* 96: 2137– 2146.

Xu Q, Wang X, Tang C. 2017. Wheat and white lupin differ in rhizosphere priming of soil organic carbon under elevated CO<sub>2</sub>. *Plant and Soil* 421: 43–55.

Yao Q, Li X, Feng G, Christie P. 2001. Mobilization of sparingly soluble inorganic phosphates by the external mycelium of an abuscular mycorrhizal fungus. *Plant* and Soil 230: 279–285.

Yavitt JB, Harms KE, Garcia MN, Mirabello MJ, Wright SJ. 2011. Soil fertility and fine root dynamics in response to 4 years of nutrient (N, P, K) fertilization in a lowland tropical moist forest, Panama. *Austral Ecology* 36: 433–445.

Yokoyama D, Imai N, Kitayama K. 2017. Effects of nitrogen and phosphorus fertilization on the activities of four different classes of fineroot and soil phosphatases in Bornean tropical rain forests. *Plant and Soil* 416: 463–476.

Yuan Z, Chen HY. 2009. Global trends in senesced-leaf nitrogen and phosphorus. *Global Ecology and Biogeography* 18: 532–542.

Yuan Z, Chen HY. 2015. Negative effects of fertilization on plant nutrient resorption. *Ecology* 96: 373–380.

Zangaro W, de Assis RL, Rostirola LV, de Souza PB, Gonçalves MC, Andrade G, Nogueira MA. 2008. Changes in arbuscular mycorrhizal associations and fine root traits in sites under different plant successional phases in southern Brazil. *Mycorrhiza* 19: 37–45.

Zemp DC, Schleussner C-F, Barbosa HM, Hirota M, Montade V, Sampaio G, Staal A, Wang-Erlandsson L, Rammig A. 2017. Self-amplified Amazon forest loss due to vegetation-atmosphere feedbacks. *Nature Communications* 8: 1–10.

Zemunik G, Lambers H, Turner BL, Laliberté E, Oliveira RS. 2018. High abundance of non-mycorrhizal plant species in severely phosphorusimpoverished Brazilian campos rupestres. *Plant and Soil* 424: 255–271.

Zemunik G, Turner BL, Lambers H, Laliberté E. 2015. Diversity of plant nutrientacquisition strategies increases during long-term ecosystem development. *Nature Plants* 1: 15050.

Zeng T, Holmer R, Hontelez J, te Lintel-Hekkert B, Marufu L, de Zeeuw T, Wu F, Schijlen E, Bisseling T, Limpens E. 2018. Host-and stage-dependent secretome of the arbuscular mycorrhizal fungus *Rhizophagus irregularis*. *The Plant Journal* 94: 411–425.

Zhang L, Xu M, Liu Y, Zhang F, Hodge A, Feng G. 2016. Carbon and phosphorus exchange may enable cooperation between an arbuscular mycorrhizal fungus and a phosphate-solubilizing bacterium. *New Phytologist* 210: 1022–1032.

Zheng J, Han S, Wang Y, Zhang C, Li M. 2010. Composition and function of microbial communities during the early decomposition stages of foliar litter exposed to elevated CO<sub>2</sub> concentrations. *European Journal of Soil Science* 61: 914–925. Zhu Q, Riley W, Tang J, Koven C. 2016. Multiple soil nutrient competition between plants, microbes, and mineral surfaces: model development, parameterization, and example applications in several tropical forests. *Biogeosciences* 13: 341–363.

# **Supporting Information**

Additional Supporting Information may be found online in the Supporting Information section at the end of the article.

**Notes S1** Leaf P<sub>i</sub>-resorption efficiency.

Notes S2 Effects of N on acid phosphatase exudation and activity.

Notes S3 Cluster roots in Amazonia.

Table S1 Summary of soil P gradient studies.

**Table S2** Phosphorus concentrations in senesced leaves in different regions of Amazonia.

Table S3 Summary of P<sub>i</sub> fertilization studies.

**Table S4** Root morphological traits in Amazonia, other tropicalforests and meta-analyses.

**Table S5** Potential root acid phosphatase activity in Amazonia andother tropical forests.

Table S6 Plant and mycorrhizal responses to  $eCO_2$  in metaanalyses.

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