

Mitochondrial DNA Part B Resources

ISSN: (Print) 2380-2359 (Online) Journal homepage: <http://www.tandfonline.com/loi/tmdn20>

Four new mitochondrial genomes of the genus zosterops (aves: passeriformes: zosteropidae) from East Africa with a phylogenetic evaluation of the group

Martin Husemann, Sarah Sturm, Manuel Curto, Harald Meimberg & Jan Christian Habel

To cite this article: Martin Husemann, Sarah Sturm, Manuel Curto, Harald Meimberg & Jan Christian Habel (2016) Four new mitochondrial genomes of the genus zosterops (aves: passeriformes: zosteropidae) from East Africa with a phylogenetic evaluation of the group, Mitochondrial DNA Part B, 1:1, 544-548, DOI: [10.1080/23802359.2016.1198937](https://doi.org/10.1080/23802359.2016.1198937)

To link to this article: <http://dx.doi.org/10.1080/23802359.2016.1198937>



© 2016 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.



Published online: 21 Aug 2016.



Submit your article to this journal [↗](#)



Article views: 180



View related articles [↗](#)



View Crossmark data [↗](#)

Full Terms & Conditions of access and use can be found at
<http://www.tandfonline.com/action/journalInformation?journalCode=tmdn20>

Four new mitochondrial genomes of the genus *Zosterops* (aves: passeriformes: zosteropidae) from East Africa with a phylogenetic evaluation of the group

Martin Husemann^{a,b,c}, Sarah Sturm^b, Manuel Curto^{d,e}, Harald Meimberg^e and Jan Christian Habel^b

^aDepartment of General Zoology, Institute of Biology, Martin-Luther University Halle-Wittenberg, Halle, Saale, Germany; ^bDepartment of Ecology and Ecosystem Management, Terrestrial Ecology Research Group, School of Life Sciences Weihenstephan, Technische Universität München, Freising, Germany; ^cCentrum Für Naturkunde, Universität Hamburg, Hamburg, Germany; ^dResearch Center in Biodiversity and Genetic Resources (CIBIO)/InBio Associated Laboratory, University of Porto, Vairão, Portugal; ^eInstitute for Integrative Nature Conservation Research, University of Natural Resources and Life Sciences (BoKu), Vienna, Austria

ABSTRACT

The white-eye birds of the genus *Zosterops* have been recognized for their high speciation rates in the past, but the relationships of the East African populations are not yet fully resolved. We sequenced and annotated mitogenomes of four populations currently assigned to three East African white-eye species, *Zosterops senegalensis*, *Z. abyssinicus* and *Z. poliogaster*. For *Z. senegalensis* specimens from two distant populations were sequenced; for the other taxa we used samples collected at one site. The mitogenomes ranged between 17,827 and 17,974 bp, in size similar to previously published mitogenomes analyzed for this genus from other geographic regions. The mitogenomes contain the classical set of 13 coding genes, two structural rRNA genes and 22 tRNA genes. We constructed a phylogeny using all complete mitogenomes currently available for the genus. The phylogeny supports an Asian or Oceanic origin of the genus *Zosterops*. The East African species represent a monophyletic clade, but the two specimens of *Zosterops senegalensis* from different regions do not group together, supporting previous hypotheses of cryptic species within the genus. The new genetic resources provided here may help to further explore the relationships and evolution of the genus.

ARTICLE HISTORY

Received 18 May 2016
Revised 1 June 2016
Accepted 4 June 2016

KEYWORDS

Illumina MiSeq; primer walking; *Zosterops abyssinicus*; *Zosterops poliogaster*; *Zosterops senegalensis*

Introduction

Mitogenomes provide important information for phylogenetic and phylogeographic studies. They are characterized by functional genes with a variety of different evolutionary rates and hence can be used for analyses of taxa with different divergent times (Curole & Kocher 1999; Jacobsen et al. 2012). They have been shown to be highly informative to resolve the basic taxonomic relationships within the Passerine birds (Barker 2014).

The Passerine bird genus *Zosterops* is an interesting target for evolutionary biology as it is characterized by high rates of speciation and a large number of endemic species, especially island endemics (Gill 1971; Moyle et al. 2009; Milá et al. 2010; Cornetti et al. 2015). In East Africa, only three species have been recognized as valid species for a long time (Moreau 1957), but more recently it has become evident that local populations are strongly differentiated from each other (genetically, phenotypically, specifically in regard of plumage coloration and bioacoustic traits) and hence should be treated as distinct taxa (Habel et al. 2013, 2014, 2015; Cox et al. 2014; Husemann et al. 2014, 2015). However, genetic resources for the group are still limited reducing phylogenetic resolution in previous studies.

More recently, several mitogenomes of *Zosterops* species have become available (*Zosterops lateralis* – Gibb et al. 2015;

Zosterops erythropleurus – Li et al. 2016; *Zosterops japonicus* – Yang et al. in press). However, so far no (mito)genome of an East African species was sequenced. Hence, we used a hybrid approach of Sanger Sequencing and Illumina MiSeq to generate whole mitochondrial genomes of four populations identified as three species of East African *Zosterops* species. We annotated the genomes and provide a phylogeny based on these and all other published mitogenome sequences of *Zosterops*.

Material and methods

We included 17 individuals of *Z. senegalensis* collected at two sites, Kakamega Forest (0°17' N; 34°50' E) (1 individual for Sanger sequencing and 7 individuals for Illumina MiSeq), and Mt. Nyeri (2°05' N, 36°42' E) (1 individual for Sanger sequencing and 8 individuals for Illumina MiSeq); 5 individuals of *Z. poliogaster* were collected in the Chyulu Hills (2°36' S; 37°50' E) (2 individuals for Sanger sequencing, 3 individuals for Illumina MiSeq); 12 individuals of *Z. abyssinicus* were collected at the foothills of the Chyulu Hills (2°25' S; 37°55' E) (with 2 individuals for Sanger sequencing and 10 individuals for Illumina MiSeq). Birds were caught with mist nets and a blood

sample was taken and stored in 99% ethanol. The birds were subsequently released unharmed.

DNA was extracted from blood using the Qiagen DNeasy blood and tissue kit (Qiagen, Hilden, Germany) using the manufacturers protocol for blood. DNA extracts are stored at the Technical University Munich under the following internal accession numbers: *Z. senegalensis* – Kakamega Forest: T41, T42, T49, T50-T54; Mt. Nyeri: T58, T71-T76, T82, T83; *Z. polioaster*: CH1, CH2-1, CH2-2, CH2-3, CH2-4; *Z. abyssinicus*: T05-T12, T17-20. PCRs were performed with standard protocols.

We initially sequenced 13 mitochondrial gene fragments using the primers provided by Amer et al. (2013). Their primer set lacked primers for the Control Region and the primers for ND5, COII and COIII did not yield good results. The lack of primer binding was further confirmed by mapping all primers against a consensus sequence derived from 41 Passerine mitogenomes obtained from Genbank (Figure 1A, Accessions: KF509924, NC020429, JQ348398, NC015114, HQ690245, NC020427, JQ423933, NC015233, HQ896034, NC015196, HQ915865, NC015198, HQ915866, NC015232, HQ896033, NC015802, JN018411, NC015074, HQ690246, NC015898, JF937590, NC020423, JQ003191, JQ003192, NC015237, HQ896037, NC015613, JF810423, NC015195, HQ915864, NC020424, JQ083495, NC020426, JQ423932, NC015824, JN108020, KF509923, NC015810, JN018413, NC015200, HQ915867). Therefore, we designed a new set of 22 primer pairs based on the consensus sequence and the preliminary assemblies for *Zosterops* derived from the initial primer set (available from the authors upon request, Figure 1A).

The newly generated sequences were trimmed and *de-novo* assembled using Geneious v. 6.1.8 (Kearse et al. 2012). For this, sequences from individuals belonging to the same populations were pooled. The resulting contigs were mapped against the published sequences of *Z. lateralis* (KC545407) to generate an initial assembly. The assemblies were further refined using Illumina MiSeq reads of specimens from identical populations (see above), which were generated for a different study with a new amplicon-based protocol that is described elsewhere (Meimberg et al. 2016). The final alignments were initially annotated using MitoAnnotator (Iwasaki et al. 2013) and Mitos (Bernt et al. 2013). The initial annotation was further refined by comparison to other published mitogenomes of *Zosterops* species (*Z. lateralis* – KC545407, *Z. erythropleurus* – KT194322, *Z. japonicus* – KT601061). Finally, we adjusted annotations of coding genes to implement stop codons, which were not implemented in the published mitogenomes for two of the genes.

We used the new mitogenomes together with published sequences to generate a phylogenetic tree with MrBayes v.3.2.6 (Ronquist et al. 2012). We included the newly generated sequences for *Z. abyssinicus* from the Chyulu Hills foothills, *Z. polioaster* from the Chyulu Hills, and sequences

for *Z. senegalensis* from Kakamega Forest and Mt. Nyeri. We further included all other available sequences for *Zosterops* species (*Z. lateralis* – KC545407, *Z. erythropleurus* – KT194322, *Z. japonicus* – KT601061) and used the consensus sequence of Passerines (see above) as outgroup. The best model of sequence evolution was determined as GTR+G by MrModeltest v.2 (Nylander 2004). We ran Mr Bayes for 10 million generations sampling every 1000 generations for a total of 10,000 sampled trees. Convergence was checked by the average split frequencies which were below 0.001. The tree was visualized using FigTree v.1.4.2 (<http://tree.bio.ed.ac.uk/software/figtree/>).

Results and discussion

We generated mitogenomic sequences for four populations of East African *Zosterops*: two populations of *Z. senegalensis* from Kakamega Forest and Mt. Nyeri, *Z. polioaster* from the Chyulu Hills and *Z. abyssinicus* from the foothills of the Chyulu Hills. The four newly generated mitogenomes varied in size between 17,827 and 17,974 bp, similar to other mitogenomes published for this genus. They contain the classical set of 13 coding genes, two structural rRNA genes and 22 tRNA genes (Figure 1B). The tRNAs for Leucine and Serine were present in two copies. Similar to other *Zosterops* species we detected two Control Regions. The mitogenomic gene order was similar across all *Zosterops* which have been analyzed so far. The GC content varied between 44.5% for *Z. senegalensis* from Kakamega forest and 45.6% for *Z. polioaster*. All newly generated sequences were deposited in NCBI GenBank (Accession Numbers: KX181885- KX181888).

Most relationships were strongly supported in our phylogenetic reconstruction (Figure 2; posterior probabilities >0.95). *Zosterops lateralis* was the basal species in the phylogeny. *Zosterops japonicus* and *Z. erythropleurus* were sister taxa and represented the sister group to the East African species. This was the only relationship with no support (pp =0.53). This may be due to limited sampling or saturation effects. The Eastern African species were monophyletic with maximum support. However, *Zosterops senegalensis* was polyphyletic with the population from Kakamega Forest being basal to all other species and the population from Mt. Nyeri being sister to *Z. abyssinicus*. *Zosterops polioaster* was the sister taxon to this grouping.

The phylogeny provides further support to previous findings in regard of the Asian or Oceanic origin of the genus (Moyle et al. 2009; Cox et al. 2014). Further, our findings support the non-monophyly of the currently established East African species of the genus, which had been suggested on the basis of morphological, acoustic, microsatellite and sequence data (Habel et al. 2013, 2014, 2015; Cox et al. 2014; Husemann et al. 2014, 2015).

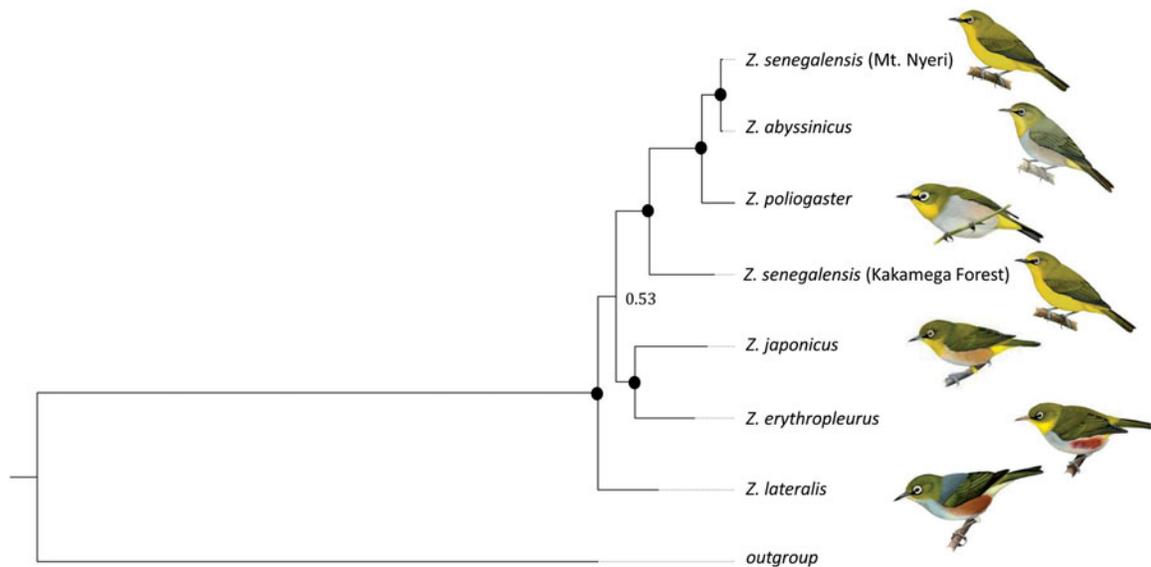


Figure 2. Bayesian phylogenetic tree generated from all available *Zosterops* mitogenomes; black circles indicate posterior probabilities of 1.

Acknowledgement

The drawings of birds for [Figure 2](#) were generously provided by the Handbook of the Birds of the World Alive (del Hoyo et al. 2014).

Disclosure statement

The authors declare no conflicts of interest. The authors alone are responsible for the content and writing of the paper.

Funding information

We would like to thank the DAAD for financial support to MH and JCH. The study was partially funded by a flexible pool grant to JCH and Wolfgang W. Weisser (TUM, Freising, Germany) of the German Centre of integrative Biodiversity Research (iDiv). This work was supported by the German Research Foundation (DFG) and the Technische Universität München within the funding programme Open Access Publishing.

References

- Amer SAM, Ahmed M, Shobrak M. 2013. Efficient newly designed primers for the amplification and sequencing of bird mitochondrial genomes. *Biosci Biotechnol Biochem.* 77:577–581.
- Barker FK. 2014. Mitogenomic data resolve basal relationships among passeriform and passerid birds. *Mol Phyl Evol.* 79:313–324.
- Bernt M, Donath A, Jühling F, Externbrink F, Florentz C, Fritzsche G, Pütz J, Middendorf M, Stadler PF. 2013. MITOS: improved de novo Metazoan mitochondrial genome annotation. *Mol Phylogenet Evol.* 69:313–319.
- Cornetti L, Valente LM, Dunning LT, Quan X, Black RA, Hébert O, Savolainen V. 2015. The genome of the “Great Speciator” provides insights into bird diversification. *Genome Biol Evol.* 7:2680–2691.
- Cox SC, Prys-Jones RP, Habel JC, Amakobe B, Day JJ. 2014. Niche divergence promotes rapid diversification of East African sky island white-eyes (Aves: Zosteropidae). *Mol Ecol.* 23:4103–4118.
- Curole J, Kocher T. 1999. Mitogenomics: digging deeper with complete mitochondrial genomes. *Trends Ecol Evol (Amst.)*. 14:394–398.
- del Hoyo J, Elliott A, Sargatal J, Christie DA, de Juana E. 2014. Handbook of the birds of the world alive. Barcelona: Lynx Edicions. Retrieved from <http://www.hbw.com/on>
- Gibb GC, England R, Hartig G, McLenachan PA, Taylor Smith BL, McComish BJ, Cooper A, Penny D. 2015. New Zealand passerines help clarify the diversification of major songbird lineages during the oligocene. *Genome Biol Evol.* 7:2983–2995.
- Gill FB. 1971. Ecology and evolution of the sympatric Mascarene white-eyes, *Zosterops borbonica* and *Zosterops olivacea*. *Auk.* 88:35–60.
- Habel JC, Cox S, Gassert F, Mulwa RK, Meyer J, Lens L. 2013. Population genetics of the East African white-eye species complex. *Conserv Genet.* 14:1019–1028.
- Habel JC, Mulwa RK, Gassert F, Rödder D, Ulrich W, Borghesio L, Husemann M, Lens L. 2014. Population signatures of large-scale, long-term disjunction and small-scale, short-term habitat fragmentation in an Afrotropical forest bird. *Heredity.* 113:205–214.
- Habel JC, Borghesio L, Newmark WD, Day JJ, Lens L, Husemann M, Ulrich W. 2015. Evolution along the Great Rift Valley: phenotypic and genetic differentiation of East African white-eyes (Aves, Zosteropidae). *Ecol Evol.* 5:4849–4862.
- Husemann M, Ulrich W, Habel JC. 2014. The evolution of contact calls in isolated and overlapping populations of two white-eye congeners in East Africa (Aves, Zosterops). *BMC Evol Biol.* 14:115.
- Husemann M, Cousseau L, Borghesio L, Lens L, Habel JC. 2015. Effects of population size and isolation on the genetic structure of the East African mountain white-eye *Zosterops polioaster* (Aves). *Biol J Linn Soc.* 114:828–836.
- Iwasaki W, Fukunaga T, Isagozawa R, Yamada K, Maeda Y, Satoh TP, Sado T, Mabuchi K, Takeshima H, Miya M, Nishida M. 2013. Mitofish and MitoAnnotator: a mitochondrial genome database of fish with an accurate and automatic annotation pipeline. *Mol Biol Evol.* 30:2531–2540.
- Jacobsen MW, Hansen MM, Orlando L, Bekkevold D, Bernatchez L, Willerslev E, Gilbert TP. 2012. Mitogenome sequencing reveals shallow evolutionary histories and recent divergence time between morphologically and ecologically distinct European whitefish (*Coregonus* spp.). *Mol Ecol.* 21:2727–2742.
- Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, et al. 2012. Geneious basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics.* 28:1647–1649.
- Li YM, Yao JY, Zhao X, Li LY, Yan SQ. 2016. Complete mitochondrial genome sequence of Chestnut-flanked white-eye (*Zosterops erythropleurus*). *Mitochondrial DNA A DNA Mapp Seq Anal.* 27:3529–3530.
- Meimberg H, Schachtler C, Curto M, Husemann M, Habel JC. 2016. A new amplicon based approach of whole genome sequencing for phylogenetic and phylogeographic analysis: an example of East African white-eyes (Aves: Zosteropidae). *Mol Phylogenet Evol.* 102:74–85. [dx.doi.org/10.1016/j.ympev.2016.05.023](https://doi.org/10.1016/j.ympev.2016.05.023)
- Milá B, Warren BH, Heeb P, Thébaud C. 2010. The geographic scale of diversification on islands: genetic and morphological divergence at a very small spatial scale in the Mascarene grey white-eye (Aves: *Zosterops borbonicus*). *BMC Evol Biol.* 10:158.

- Moreau RE. 1957. Variation in the western Zosteropidae (Aves). *Bull Br Mus.* 4:318–433.
- Moyle RG, Filardi CE, Smith CE, Diamond J. 2009. Explosive pleistocene diversification and hemispheric expansion of a ‘great speciator’. *Proc Nat Acad Sci Am.* 106:1863–1868.
- Nylander JAA. 2004. MrModeltest version 2. Program distributed by the author. Uppsala: Evolutionary Biology Centre, Uppsala University.
- Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP. 2012. MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Syst Biol.* 61:1–4.
- Yang D-C, Sun Y, Lu C-H. in press. The complete mitochondrial genome of *Zosterops japonicas*. Mitochondrial DNA. DOI: 10.3109/19401736.2015.1101581.