

RESEARCH ARTICLE

The Influence of Age and Sex on Genetic Associations with Adult Body Size and Shape: A Large-Scale Genome-Wide Interaction Study

Thomas W. Winkler¹, Anne E. Justice², Mariaelisa Graff², Llida Barata³, Mary F. Feitosa³, Su Chu⁴, Jacek Czajkowski³, Tõnu Esko^{5,6,7,8}, Tove Fall^{9,10}, Tuomas O. Kilpeläinen^{11,12}, Yingchang Lu^{13,14}, Reedik Mägi^{7,15}, Evelin Mihailov⁷, Tune H. Pers^{6,8,16}, Sina Rüeger^{17,18}, Alexander Teumer^{19,20}, Georg B. Ehret^{21,22}, Teresa Ferreira¹⁵, Nancy L. Heard-Costa^{23,24}, Juha Karjalainen²⁵, Vasiliki Lagou^{15,26}, Anubha Mahajan¹⁵, Michael D. Neinast²⁷, Inga Prokopenko^{15,26,28,29}, Jeannette Simino³⁰, Tanya M. Teslovich⁴, Rick Jansen³¹, Harm-Jan Westra^{32,33,34}, Charles C. White³⁵, Devin Absher³⁶, Tarunveer S. Ahluwalia^{11,37,38}, Shafqat Ahmad³⁹, Eva Albrecht⁴⁰, Alexessander Couto Alves⁴¹, Jennifer L. Bragg-Gresham⁴, Anton J. M. de Craen⁴², Joshua C. Bis^{43,44}, Amélie Bonnefond^{45,46,47}, Gabrielle Boucher⁴⁸, Gemma Cadby⁴⁹, Yu-Ching Cheng^{50,51}, Charleston W. K. Chiang⁵², Graciela Delgado⁵³, Ayse Demirkan⁵⁴, Nicole Dueker⁵⁵, Niina Eklund^{56,57,58}, Gudny Eiriksdottir⁵⁹, Joel Eriksson⁶⁰, Bjarke Feenstra⁶¹, Krista Fischer⁷, Francesca Frau^{62,63}, Tessel E. Galesloot⁶⁴, Frank Geller⁶¹, Anuj Goel^{15,65}, Mathias Gorski^{1,66}, Tanja B. Grammer⁵³, Stefan Gustafsson^{9,10}, Saskia Haitjema⁶⁷, Jouke-Jan Hottenga⁶⁸, Jennifer E. Huffman^{24,69}, Anne U. Jackson⁴, Kevin B. Jacobs^{70,71}, Åsa Johansson^{9,72}, Marika Kaakinen^{41,73}, Marcus E. Kleber⁵³, Jari Lahti^{74,75}, Irene Mateo Leach⁷⁶, Benjamin Lehne⁷⁷, Youfang Liu⁷⁸, Ken Sin Lo⁴⁸, Mattias Lorentzon⁶⁰, Jian'an Luan¹², Pamela A. F. Madden⁷⁹, Massimo Mangino⁸⁰, Barbara McKnight^{43,81,82}, Carolina Medina-Gomez^{83,84,85}, Keri L. Monda^{2,86}, May E. Montasser⁸⁷, Gabriele Müller⁸⁸, Martina Müller-Nurasyid^{40,89,90,91}, Ilja M. Nolte⁹², Kalliope Panoutsopoulou⁹³, Laura Pascoe⁹⁴, Lavinia Paternoster⁹⁵, Nigel W. Rayner^{15,26,93}, Frida Renström³⁹, Federica Rizzi^{62,63}, Lynda M. Rose⁹⁶, Kathy A. Ryan⁸⁷, Perttu Salo^{56,57}, Serena Sanna⁹⁷, Hubert Scharnagl⁹⁸, Jianxin Shi⁹⁹, Albert Vernon Smith^{59,100}, Lorraine Southam^{15,93}, Alena Stančáková¹⁰¹, Valgerdur Steinthorsdottir¹⁰², Rona J. Strawbridge^{103,104}, Yun Ju Sung³⁰, Ioanna Tachmazidou⁹³, Toshiko Tanaka¹⁰⁵, Gudmar Thorleifsson¹⁰², Stella Trompet^{42,106}, Natalia Pervjakova^{7,57,107,108}, Jonathan P. Tyrer¹⁰⁹, Liesbeth Vandenput⁶⁰, Sander W van der Laan⁶⁷, Nathalie van der Velde^{85,110}, Jessica van Setten¹¹¹, Jana V. van Vliet-Ostaptchouk¹¹², Niek Verweij⁷⁶, Efthymia Vlachopoulou¹¹³, Lindsay L. Waite³⁶, Sophie R. Wang^{8,32,114,115}, Zhaoming Wang^{70,71}, Sarah H. Wild¹¹⁶, Christina Willenborg^{117,118}, James F. Wilson¹¹⁹, Andrew Wong¹²⁰, Jian Yang¹²¹, Loïc Yengo^{45,46,47}, Laura M. Yerges-Armstrong⁸⁷, Lei Yu¹²², Weihua Zhang^{77,123}, Jing Hua Zhao¹², Ehm A. Andersson¹¹, Stephan J. L. Bakker¹²⁴, Damiano Baldassarre^{125,126}, Karina Banasik¹¹, Matteo Barcella⁶², Cristina Barlassina⁶², Claire Bellis^{127,128}, Paola Benaglio^{129,130}, John Blangero¹²⁷, Matthias Blüher^{131,132}, Fabrice Bonnet¹³³, Lori L. Bonnycastle¹³⁴, Heather A. Boyd⁶¹, Marcel Bruinenberg¹³⁵, Aron S Buchman¹²², Harry Campbell¹¹⁹, Yii-Der Ida Chen¹³⁶, Peter S. Chines¹³⁴, Simone Claudi-Boehm¹³⁷, John Cole^{50,138}, Francis S. Collins¹³⁴, Eco J. C. de Geus^{68,139}, Lisette C. P. G. M. de Groot¹⁴⁰, Maria Dimitriou^{93,141}, Jubao Duan^{142,143}, Stefan Enroth^{9,72}, Elodie Eury^{45,46,47}, Aiki-Eleni Farmaki¹⁴⁴, Nita G. Forouhi¹², Nele Friedrich¹⁴⁵, Pablo V. Gejman^{142,143}, Bruna Gigante¹⁴⁶, Nicola Glorioso¹⁴⁷, Alan S. Go¹⁴⁸, Omri Gottesman^{13,149}, Jürgen Gräßler¹⁵⁰, Harald Grallert^{151,152,153}, Niels Grarup¹¹, Yu-Mei Gu¹⁵⁴, Linda Broer⁸⁵, Annelies C. Ham⁸⁵, Torben Hansen^{11,155}, Tamara B. Harris^{156,157}, Catharina A. Hartman¹⁵⁸, Maija Hassinen¹⁵⁹, Nicholas Hastie⁶⁹, Andrew T. Hattersley¹⁶⁰, Andrew C. Heath⁷⁹, Anjali K. Henders¹⁶¹, Dena Hernandez¹⁶², Hans Hillege⁷⁶, Oddgeir Holmen¹⁶³, Kees



OPEN ACCESS

Citation: Winkler TW, Justice AE, Graff M, Barata L, Feitosa MF, Chu S, et al. (2015) The Influence of Age and Sex on Genetic Associations with Adult Body Size and Shape: A Large-Scale Genome-Wide Interaction Study. *PLoS Genet* 11(10): e1005378. doi:10.1371/journal.pgen.1005378

Editor: Adebowale Adeyemo, National Institute of Health, National Human Genome Research Institute, UNITED STATES

Received: November 19, 2014

Accepted: June 22, 2015

Published: October 1, 2015

Copyright: This is an open access article, free of all copyright, and may be freely reproduced, distributed, transmitted, modified, built upon, or otherwise used by anyone for any lawful purpose. The work is made available under the [Creative Commons CC0](https://creativecommons.org/licenses/by/4.0/) public domain dedication.

Data Availability Statement: The stratified genome-wide association meta-analysis results for BMI and waist-hip ratio are available from the GIANT Consortium website www.broadinstitute.org/collaboration/giant

Funding: Funding for this study was provided by the Aarne Koskelo Foundation; the Aase and Ejner Danielsens Foundation; the Academy of Finland (40758, 41071, 77299, 102318, 104781, 117787, 117844, 118590, 120315, 121584, 123885, 124243, 124282, 126925, 129269, 129293, 129378, 130326,

134309, 134791, 136895, 139635, 211497, 263836, 263924, 1114194, 24300796); the Agency for Health Care Policy Research (HS06516); the Agency for Science, Technology and Research of Singapore (A*STAR); the Ahokas Foundation; the ALF/LUA research grant in Gothenburg; the ALK-Abello A/S (Hørsholm, Denmark), Timber Merchant Vilhelm Bangs Foundation, MEKOS Laboratories Denmark; the Althingi (the Icelandic Parliament); the American Heart Association (AHA; 13POST16500011); the ANR ("Agence Nationale de la 359 Recherche"); the Ark (NHMRC Enabling Facility); the Arthritis Research UK (19542, 18030); the AstraZeneca; the Augustinus Foundation; the Australian National Health and Medical Research Council (NHMRC; 241944, 389875, 389891, 389892, 389938, 442915, 442981, 496739, 496688, 552485, 613672, 613601 and 1011506); the Australian Research Council (ARC; DP0770096 and DP1093502); the Becket Foundation; the bi-national BMBF/ANR funded project CARDomics (01KU0908A); the Biobanking and Biomolecular Resources Research Infrastructure (BBMRINL; 184.021.007, CP 32); the Biocentrum Helsinki; the Boehringer Ingelheim Foundation; the British Heart Foundation (RG/10/12/28456, SP/04/002); the Canadian Institutes for Health Research (FRCN-CCT-83028); the Cancer Research UK (C490/A10124, C490/A10119); the Center for Medical Systems Biology (CMSB; NWO Genomics); the Centers for Disease Control and Prevention and Association of Schools of Public Health (1734, S043, S3486); the Centre of Excellence Baden-Württemberg Metabolic Disorders; the Chief Scientist Office of the Scottish Government; the Clinical Research Facility at Guys & St Thomas NHS Foundation Trust; the Contrat de Projets État-Région (CPER); the Croatian Science Council (Grant no. 8875); the CVON (GENIUS); the Danish Agency for Science, Technology and Innovation; the Danish Centre for Health Technology Assessment, Novo Nordisk Inc.; the Danish Council for Independent Research (DFF 1333-00124); the Danish Diabetes Association; Danish Heart Foundation; the Danish Medical Research Council; the Danish Ministry of Internal Affairs and Health; the Danish National Research Foundation; the Danish Pharmaceutical Association; Danish Pharmacists Fund; the Danish Research Council; the Deutsche Forschungsgemeinschaft; the Diabetes Hilfs- und Forschungsfonds Deutschland (DHFD); the Dr. Robert Pflieger-Stiftung; the Dresden University of Technology Funding Grant, Med Drive; the Dutch Brain Foundation; the Dutch Diabetes Research Foundation; the Dutch Economic Structure Enhancing Fund (FES); the Dutch Kidney Foundation; the Dutch Ministry for Health, Welfare and Sports; the Dutch Ministry of Economic Affairs;

G Hovingh¹⁶⁴, Jennie Hui^{165,166,167}, Lise L. Husemoen¹⁶⁸, Nina Hutri-Kähönen^{169,170}, Pirro G. Hysi⁸⁰, Thomas Illig^{151,171,172}, Philip L. De Jager^{32,173,174}, Shapour Jalilzadeh^{15,65}, Torben Jørgensen^{168,175,176}, J. Wouter Jukema^{106,177}, Markus Juonala^{178,179,180}, Stavroula Kanoni^{93,181}, Maria Karaleftheri¹⁸², Kay Tee Khaw¹⁸³, Leena Kinnunen¹⁸⁴, Steven J. Kittner^{50,138}, Wolfgang Koenig¹⁸⁵, Ivana Kolcic¹⁸⁶, Peter Kovacs¹³¹, Nikolaj T. Krarup¹¹, Wolfgang Kratzer¹³⁷, Janine Krüger¹⁸⁷, Diana Kuh¹²⁰, Meena Kumari¹⁸⁸, Theodosios Kyriakou^{15,65}, Claudia Langenberg^{12,188}, Lars Lannfelt¹⁸⁹, Chiara Lanzani^{190,191}, Vaneet Lotay¹⁹², Lenore J. Launer¹⁵⁷, Karin Leander¹⁴⁶, Jaana Lindström¹⁹³, Allan Linneberg^{168,175,194}, Yan-Ping Liu¹⁵⁴, Stéphane Lobbens^{45,46,47}, Robert Luben¹⁹⁵, Valeriya Lyssenko^{37,196}, Satu Männistö⁵⁶, Patrik K. Magnusson¹⁹⁷, Wendy L. McArdle¹⁹⁸, Cristina Menni⁸⁰, Sigrun Merger¹³⁷, Lili Milani⁷, Grant W. Montgomery¹⁶¹, Andrew P. Morris^{15,199}, Narisu Narisu¹³⁴, Mari Nelis⁷, Ken K. Ong^{12,120,200}, Aarno Palotie^{58,93,201}, Louis Pérusse^{202,203}, Irene Pichler²⁰⁴, Maria G. Pilia⁹⁷, Anneli Pouta^{205,206}, Myriam Rheinberger⁶⁶, Rasmus Ribel-Madsen¹¹, Marcus Richards¹²⁰, Kenneth M. Rice⁸², Treva K. Rice^{30,207}, Carlo Rivolta¹³⁰, Veikko Salomaa⁵⁶, Alan R. Sanders^{142,143}, Mark A. Sarzynski²⁰⁸, Salome Scholtens⁹², Robert A. Scott¹², William R. Scott^{77,123}, Sylvain Sebert⁷³, Sebanti Sengupta⁴, Bengt Sennblad^{103,104,209}, Thomas Seufferlein¹³⁷, Angela Silveira^{103,104}, P. Eline Slagboom²¹⁰, Jan H. Smit³¹, Thomas H. Sparsø¹¹, Kathleen Stirrups^{93,181}, Ronald P. Stolk⁹², Heather M. Stringham⁴, Morris A Swertz²⁵, Amy J. Swift¹³⁴, Ann-Christine Syvänen^{9,211}, Sian-Tsung Tan^{123,212}, Barbara Thorand^{152,153}, Anke Tönjes¹³², Angelo Tremblay²⁰², Emmanouil Tsafantakis²¹³, Peter J. van der Most⁹², Uwe Völker^{20,214}, Marie-Claude Vohl^{203,215}, Judith M. Vonk⁹², Melanie Waldenberger^{91,151,152}, Ryan W. Walker^{13,14}, Roman Wennauer²¹⁶, Elisabeth Widén⁵⁸, Gonneke Willemssen⁶⁸, Tom Wilsgaard^{217,218}, Alan F. Wright⁶⁹, M. Carola Zillikens^{83,85}, Suzanne C. van Dijk⁸⁵, Natasja M. van Schoor^{139,219}, Folkert W. Asselbergs^{220,221,222}, Paul I. W. de Bakker^{111,223}, Jacques S. Beckmann¹⁷, John Beilby^{165,166}, David A. Bennett¹²², Richard N. Bergman²²⁴, Sven Bergmann^{17,130}, Carsten A. Böger⁶⁶, Bernhard O. Boehm^{137,225,226,227}, Eric Boerwinkle²²⁸, Dorret I. Boomsma⁶⁸, Stefan R. Bornstein²²⁹, Erwin P. Bottinger^{13,149}, Claude Bouchard²⁰⁸, John C. Chambers^{77,123,230}, Stephen J. Chanock^{70,81}, Daniel I. Chasman^{96,173}, Francesco Cucca^{97,231}, Daniele Cusi^{62,232}, George Dedoussis¹⁴⁴, Jeanette Erdmann^{117,118}, Johan G. Eriksson^{56,74,233}, Denis A. Evans²³⁴, Ulf de Faire¹⁴⁶, Martin Farrall^{15,65,235}, Luigi Ferrucci¹⁰⁵, Ian Ford²³⁶, Lude Franke^{25,76}, Paul W. Franks^{39,237,238}, Philippe Froguel^{45,46,47}, Ron T. Gansevoort¹²⁴, Christian Gieger^{151,152}, Henrik Grönberg¹⁹⁷, Vilmondur Gudnason^{59,100}, Ulf Gyllenstein^{9,72}, Per Hall¹⁹⁷, Anders Hamsten^{103,104,239}, Pim van der Harst^{25,76,221}, Caroline Hayward⁶⁹, Markku Heliövaara¹⁸⁴, Christian Hengstenberg^{91,240}, Andrew A Hicks²⁰⁴, Aroon Hingorani²²², Albert Hofman^{83,84}, Frank Hu^{238,241}, Heikki V. Huikuri²⁴², Kristian Hveem¹⁶³, Alan L. James²⁴³, Joanne M. Jordan⁷⁸, Antti Jula⁵⁶, Mika Kähönen^{244,245}, Eero Kajantie^{193,246}, Sekar Kathiresan^{32,247,248}, Lambertus A. L. M. Kiemeny^{64,249}, Mika Kivimäki¹⁸⁸, Paul B. Knekt²⁵⁰, Heikki A. Koistinen^{250,251,252}, Jaspal S. Kooner^{123,212,230}, Seppo Koskinen¹⁸⁴, Johanna Kuusisto¹⁰¹, Winfried Maerz^{53,98}, Nicholas G Martin¹⁶¹, Markku Laakso¹⁰¹, Timo A. Lakka^{159,253}, Terho Lehtimäki^{254,255}, Guillaume Lettre^{48,256}, Douglas F. Levinson²⁵⁷, Lars Lind²¹¹, Marja-Liisa Lokki¹¹³, Pekka Mäntyselkä^{258,259}, Mads Melbye^{61,175,260}, Andres Metspalu⁷, Braxton D. Mitchell^{51,261}, Frans L. Moll²⁶², Jeffrey C. Murray²⁶³, Arthur W. Musk²⁶⁴, Markku S. Nieminen²⁶⁵, Inger Njølstad^{217,218}, Claes Ohlsson⁶⁰, Albertine J. Oldehinkel²⁶⁶, Ben A. Oostra⁵⁴, Lyle J Palmer^{267,268}, James S. Pankow²⁶⁹, Gerard Pasterkamp⁶⁷, Nancy L. Pedersen¹⁹⁷, Oluf Pedersen¹¹, Brenda W. Penninx³¹, Markus Perola^{7,56,58}, Annette Peters^{91,151,152}, Ozren Polašek^{119,186}, Peter P. Pramstaller^{204,270}, Bruce M. Psaty^{43,44,271,272}, Lu Qi^{238,241}, Thomas Quertermous²⁶⁰, Olli T. Raitakari^{273,274}, Tuomo Rankinen²⁰⁸, Rainer Rauramaa^{159,275}, Paul M. Ridker^{96,173}, John D. Rioux^{48,256}, Fernando Rivadeneira^{83,84,85}, Jerome I. Rotter¹³⁶, Igor Rudan¹¹⁹, Hester M. den Ruijter⁶⁷, Juha Saltevo²⁷⁶, Naveed Sattar²⁷⁷, Heribert Schunkert^{91,240}, Peter E. H. Schwarz²²⁹, Alan R. Shuldiner^{87,278}, Juha Sinisalo²⁶⁵, Harold Snieder⁹², Thorkild I. A. Sørensen^{11,279,280}, Tim D. Spector⁸⁰, Jan A. Staessen^{154,281}, Bandinelli Stefania²⁸², Unnur Thorsteinsdottir^{102,283}, Michael Stumvoll^{131,132}, Jean-Claude Tardif^{48,256}, Elena Tremoli^{125,126},

the Dutch Ministry of Education, Culture and Science; the Egmont Foundation; the Else Kröner-Fresenius Stiftung (2012_A147, P48/08//A11/08); the Emil Aaltonen Foundation; the Erasmus Medical Center and Erasmus University, Rotterdam; the Estonian Ministry of Science and Education (SF0180142s08); the European Commission (223004, 2004310, DGXII, FP6-EUROSPAN, FP6-EXGENESIS, FP6-LSHG-CT-2006-018947, FP6-LSHG-CT-2006-01947, FP6-LSHM-CT-2004-503485, FP6-LSHM-CT-2006-037593, FP6-LSHM-CT-2007-037273, FP7-201379, FP7-201668, FP7-279143, FP7-305739, FP7-313010, FP7-ENGAGE-HEALTH-F4-2007-201413, FP7-EurHEALTHAgeing-277849, FP7-HEALTH-F4-2007-201550, HEALTH-2011.2.4.2-2-EU-MASCARA, HEALTH-F2-2008-201865-GEFOS, HEALTH-F7-305507 HOMAGE, LSHM-CT-2006-037593, QLG1-CT-2001-01252, QLG1-CT-2002-00896, QLG2-CT-2002-01254); the European Regional Development Fund (ERDF) and the Wissenschaftsoffensive TMO; the European Regional Development Fund to the Centre of Excellence in Genomics (EXCEGEN; 3.2.0304.11-0312); the European Research Council (ERC; 2011-StG-280559-SEPI, 2011-294713-EPLORE, 230374); the European Science Foundation (ESF; EU/QLRT-2001-01254); the EuroSTRESS project FP-006; the Finlands Slottery Machine Association; the Finnish Centre for Pensions (ETK); the Finnish Cultural Foundation; the Finnish Diabetes Association; the Finnish Diabetes Research Foundation; the Finnish Foundation for Cardiovascular Research; the Finnish Foundation for Pediatric Research; the Finnish Funding Agency for Technology and Innovation (40058/07); the Finnish Medical Society; the Finnish Ministry of Education and Culture (627:2004–2011); the Finnish Ministry of Health and Social Affairs (5254); the Finnish National Public Health Institute (current National Institute for Health and Welfare); the Finnish Special Governmental Subsidy for Health Sciences; the Finska Läkaresällskapet, Signe and Ane Gyllenberg Foundation; the Flemish League against Cancer, ITEA2 (project Care4Me); the Folkhälsan Research Foundation; the Fonds voor Wetenschappelijk Onderzoek (FWO) Vlaanderen; the Foundation for Life and Health in Finland; the Foundation for Strategic Research (SSF) and the Stockholm County Council (560283); the G. Ph. Verhagen Foundation; the Gene-diet Interactions in Obesity' project (GENDINOBI); the Genetic Association Information Network (GAIN); the GENEVA Coordinating Center (U01 HG 004446); the GenomEUtwin (EU/QLRT-2001-01254; QLG2-CT-2002-01254); the German Bundesministerium fuer Forschung und Technology (01 AK 803 A-H, 01 IG 07015 G); the German Diabetes Association; the German Ministry of Cultural Affairs; the German Federal Ministry of Education

Jaakko Tuomilehto^{193,284,285,286}, **André G. Uitterlinden**^{83,84,85}, **Matti Uusitupa**^{287,288}, **André L. M. Verbeek**⁶⁴, **Sita H. Vermeulen**^{64,289}, **Jorma S. Viikari**¹⁸⁰, **Veronique Vitart**⁶⁹, **Henry Völzke**^{19,214}, **Peter Vollenweider**²⁹⁰, **Gérard Waeber**²⁹⁰, **Mark Walker**^{32,291}, **Henri Wallaschofski**^{145,214}, **Nicholas J. Wareham**¹², **Hugh Watkins**^{15,65}, **Eleftheria Zeggini**⁹³, **CHARGE Consortium**^{292†}, **DIAGRAM Consortium**^{293†}, **GLGC Consortium**^{294†}, **Global-BPGen Consortium**^{295†}, **ICBP Consortium**^{296†}, **MAGIC Consortium**^{297†}, **Aravinda Chakravarti**²², **Deborah J. Clegg**²⁷, **L. Adrienne Cupples**^{24,35}, **Penny Gordon-Larsen**^{298,299}, **Cashell E. Jaquish**³⁰⁰, **D. C. Rao**^{3,30,207}, **Goncalo R. Abecasis**⁴, **Themistocles L. Assimes**²⁶⁰, **Inês Barroso**^{93,301,302}, **Sonja I. Berndt**⁷⁰, **Michael Boehnke**⁴, **Panos Deloukas**^{93,181,303}, **Caroline S. Fox**^{24,173}, **Leif C. Groop**^{196,304}, **David J. Hunter**^{5,238,241,305}, **Erik Ingelsson**^{9,10,15,306}, **Robert C. Kaplan**³⁰⁷, **Mark I. McCarthy**^{15,26,308}, **Karen L. Mohlke**³⁰⁹, **Jeffrey R. O'Connell**⁸⁷, **David Schlessinger**¹⁵⁷, **David P. Strachan**³¹⁰, **Kari Stefansson**^{102,283}, **Cornelia M. van Duijn**^{54,83,311}, **Joel N. Hirschhorn**^{5,6,8}, **Cecilia M. Lindgren**^{5,15}, **Iris M. Heid**^{1,40‡*}, **Kari E. North**^{312‡*}, **Ingrid B. Borecki**^{3‡*}, **Zoltán Kutalik**^{17,18,130‡*}, **Ruth J. F. Loos**^{12,13,14,313,314‡*}

1 Department of Genetic Epidemiology, Institute of Epidemiology and Preventive Medicine, University Regensburg, Regensburg, Germany, **2** Department of Epidemiology, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina, United States of America, **3** Division of Statistical Genomics, Department of Genetics, Washington University School of Medicine, St. Louis, Missouri, United States of America, **4** Center for Statistical Genetics, Department of Biostatistics, University of Michigan, Ann Arbor, Michigan, United States of America, **5** Broad Institute of the Massachusetts Institute of Technology and Harvard University, Cambridge, Massachusetts, United States of America, **6** Divisions of Endocrinology and Genetics and Center for Basic and Translational Obesity Research, Boston Children's Hospital, Boston, Massachusetts, United States of America, **7** Estonian Genome Center, University of Tartu, Tartu, Estonia, **8** Department of Genetics, Harvard Medical School, Boston, Massachusetts, United States of America, **9** Science for Life Laboratory, Uppsala University, Uppsala, Sweden, **10** Department of Medical Sciences, Molecular Epidemiology, Uppsala University, Uppsala, Sweden, **11** Novo Nordisk Foundation Center for Basic Metabolic Research, Section of Metabolic Genetics, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark, **12** MRC Epidemiology Unit, Institute of Metabolic Science, University of Cambridge, Cambridge, United Kingdom, **13** The Charles Bronfman Institute for Personalized Medicine, The Icahn School of Medicine at Mount Sinai, New York, New York, United States of America, **14** The Department of Preventive Medicine, The Icahn School of Medicine at Mount Sinai, New York, New York, United States of America, **15** Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, United Kingdom, **16** Medical and Population Genetics Program, Broad Institute of MIT and Harvard, Cambridge, Massachusetts, United States of America, **17** Swiss Institute of Bioinformatics, Lausanne, Switzerland, **18** Institute of Social and Preventive Medicine, University Hospital Lausanne (CHUV), Lausanne, Switzerland, **19** Institute for Community Medicine, University Medicine Greifswald, Greifswald, Germany, **20** Interfaculty Institute for Genetics and Functional Genomics, University Medicine Greifswald, Greifswald, Germany, **21** Department of Specialties of Internal Medicine, Geneva University Hospital, Geneva, Switzerland, **22** Center for Complex Disease Genomics, McKusick-Nathans Institute of Genetic Medicine, Johns Hopkins University School of Medicine, Baltimore, Maryland, United States of America, **23** Department of Neurology, Boston University School of Medicine, Boston, Massachusetts, United States of America, **24** National Heart, Lung, and Blood Institute, the Framingham Heart Study, Framingham, Massachusetts, United States of America, **25** University of Groningen, University Medical Center Groningen, Department of Genetics, Groningen, The Netherlands, **26** Oxford Centre for Diabetes, Endocrinology and Metabolism, University of Oxford, Churchill Hospital, Oxford, United Kingdom, **27** Department of Internal Medicine, University of Texas Southwestern Medical Center, Dallas, Texas, United States of America, **28** Hammersmith Hospital, London, United Kingdom, **29** Department of Genomics of Common Diseases, School of Public Health, Imperial College London, London, United Kingdom, **30** Division of Biostatistics, Washington University School of Medicine, St. Louis, Missouri, United States of America, **31** Department of Psychiatry and EMGO Institute for Health and Care Research, VU University Medical Center, Amsterdam, the Netherlands, **32** Program in Medical and Population Genetics, Broad Institute of Harvard and Massachusetts Institute of Technology, Cambridge, Massachusetts, United States of America, **33** Divisions of Genetics and Rheumatology, Department of Medicine, Brigham and Women's Hospital and Harvard Medical School, Boston, Massachusetts, United States of America, **34** Partners Center for Personalized Genetic Medicine, Boston, Massachusetts, United States of America, **35** Department of Biostatistics, Boston University School of Public Health, Boston, Massachusetts, United States of America, **36** HudsonAlpha Institute for Biotechnology, Huntsville, Alabama, United States of America, **37** Steno Diabetes Center A/S, Gentofte, Denmark, **38** COPSAC, Copenhagen Prospective Studies on Asthma in Childhood, Herlev and Gentofte Hospital, University of Copenhagen, Copenhagen, Denmark, **39** Department of Clinical Sciences, Genetic and Molecular Epidemiology Unit, Skåne University Hospital Malmö, Malmö, Sweden, **40** Institute of

and Research (BMBF; 03IS2061A, 03ZIK012, 01ZZ9603, 01ZZ0103, 01ZZ0403); the German National Genome Research Network (NGFN-2 and NGFN-plus); the German Research Council (SFB-1052 "Obesity mechanisms"); the Great Wine Estates of the Margaret River region of Western Australia; the Greek General Secretary of Research and Technology research grant (PENED 2003); the Gyllenberg Foundation; the Health Care Centers in Vasa, Närpes and Korsholm; the Health Fund of the Danish Health Insurance Societies; the Helmholtz Zentrum München - German Research Center for Environmental Health; the Helsinki University Central Hospital special government funds (EVO #TYH7215, #TKK2012005, #TYH2012209); the Hjartavernd (the Icelandic Heart Association); the Ib Henriksen Foundation; the Illinois Department of Public Health, and the Translational Genomics Research Institute; the INTERREG IV Oberrhein Program (Project A28); the Interuniversity Cardiology Institute of the Netherlands (ICIN; 09.001); the Italian Ministry of Health "targeted project" (ICS110.1/RF97.71); the Italian National Centre of Research InterOmics PB05_SP3; the John D and Catherine T MacArthur Foundation Research Networks on Successful Midlife Development and Socio-economic Status and Health; the Johns Hopkins University Center for Inherited Disease Research (CIDR); the Joint grant from Siemens Healthcare, Erlangen, Germany and the Federal State of Mecklenburg-West Pomerania; the Juho Vainio Foundation; the Juselius Foundation (Helsinki, Finland); the Juvenile Diabetes Research Foundation International (JDRF); the KfH Stiftung Präventivmedizin e.V.; the Knut and Alice Wallenberg Foundation; the Kuopio University Hospital; the Leenaards Foundation; the Leiden University Medical Center; the Liv och Hälsa; the Local Government Pensions Institution (KEVA); the Lokaal Gezondheids Overleg (LOGO) Leuven and Hageland; the Ludwig-Maximilians-Universität, as part of LMUinnovativ; the Lundberg Foundation; the March of Dimes Birth Defects Foundation; the Medical Research Council (G0601966; G0700931; G0000934; G0500539; G0600705; G1002319; G0701863; PrevMetSyn/SALVE; MC_U106179471; MC_UU_12019/1); the MRC centre for Causal Analyses in Translational Epidemiology (MRC CAITE); the MRC Centre for Obesity and Related Metabolic Diseases; the MRC Human Genetics Unit; the Medical Research Council of Canada; the Mid-Atlantic Nutrition and Obesity Research Center (P30 DK072488); the Ministry of the Flemish Community, Brussels, Belgium (G.0881.13 and G.0880.13); the MIUR - CNR Italian Flagship Project; the Montreal Heart Institute Foundation; the Munich Center of Health Sciences (MC Health); the Municipal Health Care Center and Hospital in Jakobstad; the Närpes Health Care Foundation; the

Genetic Epidemiology, Helmholtz Zentrum München—German Research Center for Environmental Health, Neuherberg, Germany, **41** Department of Epidemiology and Biostatistics, MRC Health Protection Agency (HPA) Centre for Environment and Health, School of Public Health, Imperial College, London, United Kingdom, **42** Department of Gerontology and Geriatrics, Leiden University Medical Center, Leiden, The Netherlands, **43** Cardiovascular Health Research Unit, University of Washington, Seattle, Washington, United States of America, **44** Department of Medicine, University of Washington, Seattle, Washington, United States of America, **45** CNRS UMR 8199, Lille, France, **46** European Genomic Institute for Diabetes, Lille, France, **47** Université de Lille 2, Lille, France, **48** Montreal Heart Institute, Montréal, Québec, Canada, **49** Centre for Genetic Origins of Health and Disease, University of Western Australia, Crawley, Western Australia, Australia, **50** VA Maryland Health Care System, Baltimore, Maryland, United States of America, **51** University of Maryland School of Medicine, Department of Medicine, Baltimore, Maryland, United States of America, **52** Department of Ecology and Evolutionary Biology, University of California, Los Angeles, Los Angeles, California, United States of America, **53** Vth Department of Medicine, Mannheim Medical Faculty, University of Heidelberg, Mannheim, Germany, **54** Genetic Epidemiology Unit, Department of Epidemiology, Erasmus University Medical Center, Rotterdam, The Netherlands, **55** University of Maryland School of Medicine, Department of Epidemiology & Public Health, Baltimore, Maryland, United States of America, **56** National Institute for Health and Welfare, Department of Chronic Disease Prevention, Helsinki, Finland, **57** National Institute for Health and Welfare, Public Health Genomics Unit, Helsinki, Finland, **58** Institute for Molecular Medicine Finland, University of Helsinki, Helsinki, Finland, **59** Icelandic Heart Association, Kopavogur, Iceland, **60** Centre for Bone and Arthritis Research, Department of Internal Medicine and Clinical Nutrition, Institute of Medicine, Sahlgrenska Academy, University of Gothenburg, Gothenburg, Sweden, **61** Department of Epidemiology Research, Statens Serum Institut, Copenhagen, Denmark, **62** Department of Health Sciences, University of Milan, Milan, Italy, **63** Filarete Foundation, Genomic and Bioinformatics Unit, Milano, Italy, **64** Radboud university medical center, Radboud Institute for Health Sciences, Department for Health Evidence, Nijmegen, The Netherlands, **65** Division of Cardiovascular Medicine, Radcliffe Department of Medicine, University of Oxford, Oxford, United Kingdom, **66** Department of Nephrology, University Hospital Regensburg, Regensburg, Germany, **67** Experimental Cardiology and laboratory of clinical chemistry, UMCU, Utrecht, The Netherlands, **68** Department of Biological Psychology, VU University, Amsterdam, The Netherlands, **69** MRC Human Genetics Unit, Institute of Genetics and Molecular Medicine, University of Edinburgh, Edinburgh, Scotland, **70** Division of Cancer Epidemiology and Genetics, National Cancer Institute, National Institutes of Health, Bethesda, Maryland, United States of America, **71** Core Genotyping Facility, SAIC-Frederick, Inc., NCI-Frederick, Frederick, Maryland, United States of America, **72** Department of Immunology, Genetics and Pathology, Uppsala University, Uppsala, Sweden, **73** Institute of Health Sciences, University of Oulu, Oulu, Finland, **74** Folkhälsan Research Centre, Helsinki, Finland, **75** Institute of Behavioural Sciences, University of Helsinki, Helsinki, Finland, **76** University of Groningen, University Medical Center Groningen, Department of Cardiology, Groningen, Netherlands, **77** Department of Epidemiology and Biostatistics, Imperial College London, London, United Kingdom, **78** Thurston Arthritis Research Center, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina, United States of America, **79** Washington University Medical School, St. Louis, Missouri, United States of America, **80** Department of Twin Research and Genetic Epidemiology, King's College London, London, United Kingdom, **81** Program in Biostatistics and Biomathematics, Division of Public Health Sciences, Fred Hutchinson Cancer Research Center, Seattle, Washington, United States of America, **82** Department of Biostatistics, University of Washington, Seattle, Washington, United States of America, **83** Netherlands Genomics Initiative (NGI)-sponsored Netherlands Consortium for Healthy Aging (NCHA), The Netherlands, **84** Department of Epidemiology, Erasmus Medical Center, Rotterdam, The Netherlands, **85** Department of Internal Medicine, Erasmus Medical Center, Rotterdam, The Netherlands, **86** The Center for Observational Research, Amgen Inc., Thousand Oaks, California, United States of America, **87** Program for Personalized and Genomic Medicine, Division of Endocrinology, Diabetes & Nutrition, Dept of Medicine, University of Maryland School of Medicine, Baltimore, Maryland, United States of America, **88** Center for Evidence Based Healthcare, University of Dresden, Medical Faculty Carl Gustav Carus, Dresden, Germany, **89** Department of Medicine I, University Hospital Grosshadern, Ludwig-Maximilians-Universität, Munich, Germany, **90** Institute of Medical Informatics, Biometry and Epidemiology, Chair of Genetic Epidemiology, Ludwig-Maximilians-Universität, Munich, Germany, **91** DZHK (German Centre for Cardiovascular Research), partnersite Munich Heart Alliance, Munich, Germany, **92** University of Groningen, University Medical Center Groningen, Department of Epidemiology, Groningen, The Netherlands, **93** Wellcome Trust Sanger Institute, Human Genetics, Hinxton, Cambridge, United Kingdom, **94** Institute of Cell & Molecular Biosciences, Newcastle University, Newcastle, United Kingdom, **95** MRC Integrative Epidemiology Unit, School of Social and Community Medicine, University of Bristol, Bristol, United Kingdom, **96** Division of Preventive Medicine, Brigham and Women's Hospital, Boston, Massachusetts, United States of America, **97** Istituto di Ricerca Genetica e Biomedica, CNR, Messorato, Italy, **98** Clinical Institute of Medical and Chemical Laboratory Diagnostics, Medical University of Graz, Graz, Austria, **99** National Cancer Institute, Bethesda, Maryland, United States of America, **100** Faculty of Medicine, University of Iceland, Reykjavik, Iceland, **101** Department of Medicine, University of Eastern Finland and Kuopio University Hospital, Kuopio, Finland,

National Alliance for Research on Schizophrenia and Depression (NARSAD); the National Cancer Institute (CA047988); the National Center for Advancing Translational Sciences (UL1TR000124); the National Center for Research Resources (U54RR020278); the National Heart, Lung and Blood Institute (NHLBI, 1RL1MH083268-01, 5R01HL087679-02, HHSN268200800007C, HHSN268201200036C, HL043851, HL080467, HL087647, HL36310, HL45670, N01HC25195, N01HC55015, N01HC55016, N01HC55018, N01HC55019, N01HC55020, N01HC55021, N01HC55022, N01HC55222, N01HC85079, N01HC85080, N01HC85081, N01HC85082, N01HC85083, N01HC85086, N02HL64278, R01HL086694, R01HL087641, R01HL087652, R01HL087676, R01HL59367, R01HL103612, R01HL105756, R01HL120393, U01HL080295); the National Human Genome Research Institute (NHGRI, U01HG004402); the National Institute for Health and Welfare (THL); the National Institute for Health Research (NIHR, RP-PG-0407-10371); the National Institute of Allergy and Infectious Diseases (NIAID); the National Institute of Child Health and Human Development (NICHD); the National Institute of Diabetes and Digestive and Kidney Disease (NIDDK-DRC, 1R01DK8925601, DK063491, R01DK089256, P30 DK072488); the National Institute of Food and Agriculture (2007-35205-17883); the National Institute of Neurological Disorders and Stroke (NINDS); the National Institute on Aging (NIA; 263-MA-410953, 263-MD-821336, 263-MD-9164, AG023629, AG13196, NO1AG12109, P30AG10161, R01AG15819, R01AG17917, R01AG023629, R01AG30146); the National Institute of Arthritis and Musculoskeletal and Skin Diseases (5-P60-AR30701, 5-P60-AR49465-03); the National Institutes of Health (NIH; 1R01DK8925601, 1RC2MH089951, 1RC2MH089995, 1Z01HG000024, 2T32 HL 007055-36, 5R01DK075681, 5R01MH63706:02, AA014041, AA07535, AA10248, AA13320, AA13321, AA13326, AG028555, AG08724, AG04563, AG10175, AG08861, DA12854, DK046200, DK091718, F32AR059469, HG002651, HHSN268200625226C, HHSN268200782096C, HL084729, MH081802, N01-AG12100, N01HG65403, R01AG011101, R01AG030146, R01D0042157-01A, R01DK062370, R01DK072193, R01DK093757, R01DK075787, R01DK075787, R01HL71981, R01MH59565, R01MH59566, R01MH59571, R01MH59586, R01MH59587, R01MH59588, R01MH60870, R01MH60879, R01MH61675, R01MH67257, R01MH81800, R01NS45012, U01066134, U01CA098233, U01DK062418, U01GM074518, U01HG004423, U01HG004436, U01HG004438, U01HL072515-06, U01HL105198, U01HL84756, U01MH79469, U01MH79470, U01NS069208-01,

102 deCODE Genetics, Amgen inc., Reykjavik, Iceland, **103** Atherosclerosis Research Unit, Department of Medicine Solna, Karolinska Institutet, Stockholm, Sweden, **104** Center for Molecular Medicine, Karolinska University Hospital, Stockholm, Sweden, **105** Translational Gerontology Branch, National Institute on Aging, Baltimore, Maryland, United States of America, **106** Department of Cardiology, Leiden University Medical Center, Leiden, The Netherlands, **107** Institute of Cell and Molecular Biology, Department of Biotechnology, University of Tartu, Tartu, Estonia, **108** University of Helsinki, Helsinki, Finland, **109** Department of Oncology, University of Cambridge, Cambridge, United Kingdom, **110** Department of Internal Medicine, Section of Geriatric Medicine, Academic Medical Center, Amsterdam, The Netherlands, **111** Department of Medical Genetics, University Medical Center Utrecht, Utrecht, Netherlands, **112** University of Groningen, University Medical Center Groningen, Department of Endocrinology, Groningen, The Netherlands, **113** Transplantation Laboratory, Haartman Institute, University of Helsinki, Helsinki, Finland, **114** Division of Endocrinology, Boston Children's Hospital, Boston, Massachusetts, United States of America, **115** Divisions of Genetics and Endocrinology and Program in Genomics, Boston's Children's Hospital, Boston, Massachusetts, United States of America, **116** Centre for Population Health Sciences, Usher Institute of Population Health Sciences and Informatics, University of Edinburgh, Edinburgh, Scotland, **117** DZHK (German Centre for Cardiovascular Research), partner site Hamburg/Kiel/Lübeck, Lübeck, Germany, **118** Institut für Integrative und Experimentelle Genomik, Universität zu Lübeck, Lübeck, Germany, **119** Centre for Global Health Research, Usher Institute of Population Health Sciences and Informatics, University of Edinburgh, Edinburgh, Scotland, **120** MRC Unit for Lifelong Health & Ageing at UCL, London, United Kingdom, **121** Queensland Brain Institute, The University of Queensland, Brisbane, Queensland, Australia, **122** Rush Alzheimer's Disease Center, Rush University Medical Center, Chicago, Illinois, United States of America, **123** Ealing Hospital NHS Trust, Middlesex, United Kingdom, **124** University of Groningen, University Medical Center Groningen, Department of Medicine, Groningen, Netherlands, **125** Centro Cardiologico Monzino, IRCCS, Milan, Italy, **126** Dipartimento di Scienze Farmacologiche e Biomolecolari, Università di Milano, Milan, Italy, **127** Department of Genetics, Texas Biomedical Research Institute, San Antonio, Texas, United States of America, **128** Genomics Research Centre, Institute of Health and Biomedical Innovation, Queensland University of Technology, Brisbane, Queensland, Australia, **129** Department of Pediatrics, University of California San Diego, La Jolla, California, United States of America, **130** Department of Medical Genetics, University of Lausanne, Lausanne, Switzerland, **131** University of Leipzig, IFB Adiposity Diseases, Leipzig, Germany, **132** University of Leipzig, Department of Medicine, Leipzig, Germany, **133** University Rennes 1, Rennes, France, **134** Medical Genetics and Metabolic Genetics Branch, National Human Genome Research Institute, NIH, Bethesda, Maryland, United States of America, **135** University of Groningen, University Medical Center Groningen, The LifeLines Cohort Study, Groningen, The Netherlands, **136** Los Angeles BioMedical Research Institute at Harbor-UCLA Medical Center, Torrance, California, United States of America, **137** Department of Internal Medicine I, Ulm University Medical Centre, Ulm, Germany, **138** University of Maryland School of Medicine, Department of Neurology, Baltimore, Maryland, United States of America, **139** EMGO Institute for Health and Care Research, VU University Medical Center, Amsterdam, The Netherlands, **140** Department of Human Nutrition, Wageningen University, Wageningen, The Netherlands, **141** Department of Dietetics-Nutrition, Harokopio University, Athens, Greece, **142** NorthShore University HealthSystem, Evanston, Illinois, United States of America, **143** University of Chicago, Chicago, Illinois, United States of America, **144** Department of Nutrition and Dietetics, School of Health Science and Education, Harokopio University, Athens, Greece, **145** Institute of Clinical Chemistry and Laboratory Medicine, University Medicine Greifswald, Greifswald, Germany, **146** Division of Cardiovascular Epidemiology, Institute of Environmental Medicine, Karolinska Institutet, Stockholm, Sweden, **147** Hypertension and Related Disease Centre, AOU-University of Sassari, Sassari, Italy, **148** Kaiser Permanente, Division of Research, Oakland, California, United States of America, **149** The Department of Medicine, The Icahn School of Medicine at Mount Sinai, New York, New York, United States of America, **150** Department of Medicine III, Pathobiochemistry, University of Dresden, Dresden, Germany, **151** Research Unit of Molecular Epidemiology, Helmholtz Zentrum München—German Research Center for Environmental Health, Neuherberg, Germany, **152** Institute of Epidemiology II, Helmholtz Zentrum München—German Research Center for Environmental Health, Neuherberg, Germany, **153** German Center for Diabetes Research (DZD), Neuherberg, Germany, **154** Research Unit Hypertension and Cardiovascular Epidemiology, KU Leuven Department of Cardiovascular Sciences, University of Leuven, Leuven, Belgium, **155** Faculty of Health Sciences, University of Southern Denmark, Odense, Denmark, **156** Laboratory of Epidemiology and Population Sciences, National Institute on Aging, Bethesda, Maryland, United States of America, **157** National Institute on Aging, National Institutes of Health, Bethesda, Maryland, United States of America, **158** University of Groningen, University Medical Center Groningen, Department of Psychiatry, Groningen, The Netherlands, **159** Kuopio Research Institute of Exercise Medicine, Kuopio, Finland, **160** Institute of Biomedical & Clinical Science, University of Exeter, Exeter, United Kingdom, **161** QIMR Berghofer Medical Research Institute, Brisbane, Queensland, Australia, **162** Laboratory of Neurogenetics, National Institute on Aging, National Institutes of Health, Bethesda, Maryland, United States of America, **163** Department of Public Health and General Practice, Norwegian University of Science and Technology, Trondheim, Norway, **164** Department Vascular Medicine, Academic Medical Center, Amsterdam, The

UL1RR025005); the NIHR Biomedical Research Centre based at Guy's and St Thomas' NHS Foundation Trust; the NIHR Cambridge Biomedical research Centre; the Netherlands Heart Foundation (2001 D 032); the Netherlands Organisation for Scientific Research (NWO); Geestkracht program grant 10-000-1002; 050-060-810; 100-001-004; 175.010.2003.005; 175.010.2005.011; 175.010.2007.006; 261-98-710; 40-0056-98-9032; 400-05-717; 452-04-314; 452-06-004; 480-01-006; 480-04-004; 480-05-003; 480-07-001; 481-08-013; 60-60600-97-118; 904-61-090; 904-61-193; 911-03-012; 985-10-002; Addiction-31160008; GB-MW 940-38-011; SPI 56-464-14192); the Netherlands Organization for the Health Research and Development (ZonMw; 91111025); the Nordic Center of Excellence in Disease Genetics; the Nordic Centre of Excellence on Systems biology in controlled dietary interventions and cohort studies, SYSDIET (070014); the Northern Netherlands Collaboration of Provinces (SNN); the Novo Nordisk Foundation; the Office of Research and Development, Medical Research Service, and the Baltimore Geriatrics Research, Education, and Clinical Center of the Department of Veterans Affairs; the Ollqvist Foundation; the Paavo Nurmi Foundation; the Pahlssons Foundation; the Päivikki and Sakari Sohlberg Foundation; the Perklén Foundation; the Republic of Croatia Ministry of Science, Education and Sports research (108-1080315-0302); the Research Centre for Prevention and Health, the Capital Region of Denmark; the Research Foundation of Copenhagen County; the Research Institute for Diseases in the Elderly (014-93-015; RIDE2); the Reynold's Foundation; the Rotterdam Oncologic Thoracic Study Group, Erasmus Trust Fund, Foundation against Cancer; the Royal Swedish Academy of Science; the Russian Foundation for Basic Research (NWO-RFBR 047.017.043); the Rutgers University Cell and DNA Repository cooperative agreement (NIMH U24 MH068457-06); the Samfundet Folkhälsan; the Sigrid Juselius Foundation; the Social Insurance Institution of Finland, Kuopio, Tampere and Turku University Hospital Medical Funds (9M048, 9N035); the Social Ministry of the Federal State of Mecklenburg-West Pomerania; the Société Francophone du 358 Diabète (SFD); the South Tyrolean Sparkasse Foundation; the Stichting Nationale Computerfaciliteiten (National Computing Facilities Foundation, NCF); the Strategic Cardiovascular Programme of Karolinska Institutet and the Stockholm County Council (560183); the Susan G. Komen Breast Cancer Foundation; the Swedish Cancer Society; the Swedish Cultural Foundation in Finland; the Swedish Diabetes Association; the Swedish Diabetes Foundation (grant no. 2013-024); the Swedish Foundation for Strategic

Netherlands, **165** Pathwest Laboratory Medicine of Western Australia, Nedlands, Western Australia, Australia, **166** School of Pathology and Laboratory Medicine, University of Western Australia, Nedlands, Western Australia, Australia, **167** School of Population Health, University of Western Australia, Nedlands, Western Australia, Australia, **168** Research Centre for Prevention and Health, Glostrup Hospital, Glostrup, Denmark, **169** Department of Pediatrics, University of Tampere School of Medicine, Tampere, Finland, **170** Department of Pediatrics, Tampere University Hospital, Tampere, Finland, **171** Hannover Unified Biobank, Hannover Medical School, Hannover, Germany, **172** Institute of Human Genetics, Hannover Medical School, Hanover, Germany, **173** Harvard Medical School, Boston, Massachusetts, United States of America, **174** Program in Translational NeuroPsychiatric Genomics, Department of Neurology, Brigham and Women's Hospital, Boston, Massachusetts, United States of America, **175** Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark, **176** Faculty of Medicine, University of Aalborg, Aalborg, Denmark, **177** Interuniversity Cardiology Institute of the Netherlands, Utrecht, The Netherlands, **178** Division of Medicine, Turku University Hospital, Turku, Finland, **179** Murdoch Children's Research Institute, Parkville, Victoria, Australia, **180** Department of Medicine, University of Turku, Turku, Finland, **181** William Harvey Research Institute, Barts and The London School of Medicine and Dentistry, Queen Mary University of London, London, United Kingdom, **182** Echinops Medical Centre, Echinops, Greece, **183** Clinical Gerontology Unit, Addenbrooke's Hospital, Cambridge, United Kingdom, **184** Department of Health, National Institute for Health and Welfare, Helsinki, Finland, **185** Department of Internal Medicine II—Cardiology, University of Ulm Medical Center, Ulm, Germany, **186** Department of Public Health, Faculty of Medicine, University of Split, Split, Croatia, **187** Department of Medicine A, University Medicine Greifswald, Greifswald, Germany, **188** Department of Epidemiology and Public Health, UCL, London, United Kingdom, **189** Department of Public Health and Caring Sciences, Geriatrics, Uppsala University, Uppsala, Sweden, **190** Chair of Nephrology, Università Vita Salute San Raffaele, Segrate (Milan), Italy, **191** Genomics of Renal Disease and Hypertension Unit, IRCCS San Raffaele Scientific Institute, Segrate (Milan), Italy, **192** Department of Biological Sciences, University of Calgary, Calgary, Alberta, Canada, **193** Diabetes Prevention Unit, National Institute for Health and Welfare, Helsinki, Finland, **194** Department of Clinical Experimental Research, Rigshospitalet, Glostrup, Denmark, **195** Strangeways Research Laboratory Wort's Causeway, Cambridge, United Kingdom, **196** Lund University Diabetes Centre and Department of Clinical Science, Diabetes & Endocrinology Unit, Lund University, Malmö, Sweden, **197** Department of Medical Epidemiology and Biostatistics, Karolinska Institutet, Stockholm, Sweden, **198** School of Social and Community Medicine, University of Bristol, Bristol, United Kingdom, **199** Department of Biostatistics, University of Liverpool, Liverpool, United Kingdom, **200** Department of Paediatrics, University of Cambridge, Cambridge, United Kingdom, **201** Massachusetts General Hospital, Center for Human Genetic Research, Psychiatric and Neurodevelopmental Genetics Unit, Boston, Massachusetts, United States of America, **202** Department of Kinesiology, Laval University, Québec City, Québec, Canada, **203** Institute of Nutrition and Functional Foods, Laval University, Québec City, Québec, Canada, **204** Center for Biomedicine, European Academy Bozen/Bolzano (EURAC), Bolzano, Italy, Affiliated Institute of the University of Lübeck, Lübeck, Germany, **205** Department of Children, Young People and Families, National Institute for Health and Welfare, Helsinki, Finland, **206** Department of Obstetrics and Gynecology, Medical Research Center Oulu, Oulu University Hospital and University of Oulu, Oulu, Finland, **207** Department of Psychiatry, Washington University School of Medicine, St. Louis, Missouri, United States of America, **208** Human Genomics Laboratory, Pennington Biomedical Research Center, Baton Rouge, Louisiana, United States of America, **209** Science for Life Laboratory, Karolinska Institutet, Stockholm, Sweden, **210** Department of Molecular Epidemiology, Leiden University Medical Center, Leiden, The Netherlands, **211** Department of Medical Sciences, Uppsala University, Uppsala, Sweden, **212** National Heart and Lung Institute, Imperial College London, London, United Kingdom, **213** Anogia Medical Centre, Anogia, Greece, **214** DZHK (German Centre for Cardiovascular Research), partner site Greifswald, Greifswald, Germany, **215** School of Nutrition, Laval University, Québec City, Québec, Canada, **216** Department of Clinical Chemistry, Ulm University Medical Centre, Ulm, Germany, **217** Department of Clinical Medicine, Faculty of Health Sciences, University of Tromsø, Tromsø, Norway, **218** Department of Community Medicine, Faculty of Health Sciences, University of Tromsø, Tromsø, Norway, **219** VUMC, Department of Epidemiology and Biostatistics, Amsterdam, The Netherlands, **220** Department of Cardiology, Division Heart and Lungs, University Medical Center Utrecht, Utrecht, Netherlands, **221** Durrer Center for Cardiogenetic Research, Interuniversity Cardiology Institute Netherlands-Netherlands Heart Institute, Utrecht, The Netherlands, **222** Institute of Cardiovascular Science, University College London, London, United Kingdom, **223** Department of Epidemiology, University Medical Center, Utrecht, The Netherlands, **224** Diabetes and Obesity Research Institute, Cedars-Sinai Medical Center, Los Angeles, California, United States of America, **225** Imperial College London, London, United Kingdom, **226** Lee Kong Chian School of Medicine, Singapore, Singapore, **227** Nanyang Technological University, Singapore, Singapore, **228** Human Genetics Center and Institute of Molecular Medicine, University of Texas Health Science Center, Houston, Texas, United States of America, **229** Department of Medicine III, University of Dresden, Medical Faculty Carl Gustav Carus, Dresden, Germany, **230** Imperial College Healthcare NHS Trust, London, United Kingdom, **231** University of Sassari, Sassari, Italy, **232** Institute of Biomedical Technologies, National Institute of Research, Segrate-Milano, Italy,

Research (SSF; ICA08-0047); the Swedish Heart-Lung Foundation (20120197); the Swedish Medical Research Council (K2007-66X-20270-01-3, 2012-1397); the Swedish Ministry for Higher Education; the Swedish Research Council (8691, M-2005-1112, 2009-2298); the Swedish Society for Medical Research; the Swiss National Science Foundation (31003A-143914, 3200B0105993, 3200B0-118308, 33CSO-122661, 33CS30-139468, 33CS30-148401); SystemsX.ch (51RTP0_151019); the Tampere Tuberculosis Foundation; the TEKES (70103/06, 40058/07); the The Paul Michael Donovan Charitable Foundation; the Torsten and Ragnar Söderberg Foundation; the Umeå Medical Research Foundation; the United Kingdom NIHR Cambridge Biomedical Research Centre; the Universities and Research of the Autonomous Province of Bolzano, South Tyrol; the University Hospital of Regensburg (ReForm A, ReForm C); the University Hospital Oulu, Biocenter, University of Oulu, Finland (75617); the University Medical Center Groningen; the University of Groningen; the University of Maryland General Clinical Research Center (M01RR16500, AG000219); the University of Tartu (SP1GVARENG); the University of Tromsø, Norwegian Research Council (185764); the Västerbottens Intervention Programme; the Velux Foundation; the VU University Institute for Health and Care Research (EMGO+) and Neuroscience Campus Amsterdam (NCA); the Wellcome Trust (064890, 068545/Z/02, 076113/B/04/Z, 077016/Z/05/Z, 079895, 084723/Z/08/Z, 086596/Z/08/Z, 088869/B/09/Z, 089062, 090532, 098017, 098051, 098381); the Western Australian DNA Bank (NHMRC Enabling Facility); the Yrjö Jahnsson Foundation (56358); and the Zorg Onderzoek Nederland-Medische Wetenschappen, KWF Kankerbestrijding, Stichting Centraal Fonds Reserves van voormalig Vrijwillige Ziekenfondsverzekeringen. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. More details of acknowledgements can be found in S2 Text.

Competing Interests: I have read the journals policy and we have the following conflicts: KSte, UT, VSte and GT are employed by deCODE Genetics/Amgen inc. IB and spouse own stock in Incyte Ltd and GlaxoSmithKline. JMJ serves as consultant for Trinity Partners, Inc., Samumed, Flexion, and is deputy editor for Osteoarthritis & Cartilage and serves on board of directors of American College of Rheumatology. PV and GWa received an unrestricted grant from GlaxoSmithKline to build the CoLaus study. BMP serves on the data and safety monitoring board (DSMB) for a clinical trial of a device funded by the manufacturer (Zoll LifeCor) and serves on the Steering Committee of the Yale Open Data Access Project funded by Johnson & Johnson.

233 Department of General Practice and Primary Health Care, University of Helsinki, Helsinki, Finland, **234** Rush Institute for Healthy Aging and Department of Internal Medicine, Rush University Medical Center, Chicago, Illinois, United States of America, **235** Durrer Center for Cardiogenetic Research, Amsterdam, The Netherlands, **236** Robertson Center for Biostatistics, University of Glasgow, Glasgow, United Kingdom, **237** Department of Public Health & Clinical Medicine, Umeå University Hospital, Umeå, Sweden, Umeå, Sweden, **238** Department of Nutrition, Harvard School of Public Health, Boston, Massachusetts, United States of America, **239** Department of Medicine, Karolinska Institutet, Stockholm, Sweden, **240** Deutsches Herzzentrum München, Technische Universität München, München, Germany, **241** Channing Division of Network Medicine, Department of Medicine, Brigham and Women's Hospital and Harvard Medical School, Boston, Massachusetts, United States of America, **242** Medical Research Center Oulu, Oulu University Hospital, and University of Oulu, Oulu, Finland, **243** Department of Pulmonary Physiology and Sleep Medicine, Sir Charles Gairdner Hospital, Nedlands, Western Australia, **244** Department of Clinical Physiology, Tampere University Hospital, Tampere, Finland, **245** Department of Clinical Physiology, University of Tampere School of Medicine, Tampere, Finland, **246** Children's Hospital, Helsinki University Hospital and University of Helsinki, Helsinki, Finland, **247** Cardiovascular Research Center and Cardiology Division, Massachusetts General Hospital, Boston, Massachusetts, United States of America, **248** Center for Human Genetics Research, Massachusetts General Hospital, Boston, Massachusetts, United States of America, **249** Radboud university medical center, Radboud Institute for Health Sciences, Department of Urology, Nijmegen, The Netherlands, **250** National Institute for Health and Welfare, Helsinki, Finland, **251** University of Helsinki and Helsinki University Central Hospital, Department of Medicine and Abdominal Center: Endocrinology, Helsinki, Finland, **252** Minerva Foundation Institute for Medical Research, Helsinki, Finland, **253** Department of Physiology, Institute of Biomedicine, University of Eastern Finland, Kuopio Campus, Kuopio, Finland, **254** Department of Clinical Chemistry, University of Tampere School of Medicine, Tampere, Finland, **255** Department of Clinical Chemistry, Fimlab Laboratories and School of Medicine, University of Tampere, Tampere, Finland, **256** Department of Medicine, Université de Montréal, Montréal, Québec, Canada, **257** Stanford University, Stanford, California, United States of America, **258** Primary Health Care Unit, Institute of Public Health and Clinical Nutrition, School of Medicine, University of Eastern Finland, Kuopio, Finland, **259** Primary Health Care Unit, Kuopio University Hospital, Kuopio, Finland, **260** Department of Medicine, Stanford University School of Medicine, Stanford, California, United States of America, **261** Geriatrics Research and Education Clinical Center, Baltimore Veterans Administration Medical Center, Baltimore, Maryland, United States of America, **262** Department of Surgery, University Medical Center Utrecht, Utrecht, Netherlands, **263** Department of Pediatrics, University of Iowa, Iowa City, Iowa, United States of America, **264** Department of Respiratory Medicine, Sir Charles Gairdner Hospital, Nedlands, Western Australia, Australia, **265** HUCH Heart and Lung Center, Division of Cardiology, Helsinki University Central Hospital, Helsinki, Finland, **266** University of Groningen, University Medical Center, Interdisciplinary Center Psychopathology and Emotion Regulation, Groningen, The Netherlands, **267** School of Public Health, University of Adelaide, Adelaide, South Australia, Australia, **268** Robinson Research Institute, University of Adelaide, Adelaide, South Australia, Australia, **269** Division of Epidemiology and Community Health, School of Public Health, University of Minnesota, Minneapolis, Minnesota, United States of America, **270** Department of Neurology, General Central Hospital, Bolzano, Italy, **271** Departments of Epidemiology and Health Services, University of Washington, Seattle, Washington, United States of America, **272** Group Health Research Institute, Group Health Cooperative, Seattle, Washington, United States of America, **273** Department of Clinical Physiology and Nuclear Medicine, Turku University Hospital, Turku, Finland, **274** Research Centre of Applied and Preventive Cardiovascular Medicine, University of Turku, Turku, Finland, **275** Department of Clinical Physiology and Nuclear Medicine, Kuopio University Hospital, Kuopio, Finland, **276** Department of Medicine, Central Finland Central Hospital, Jyväskylä, Finland, **277** BHF Glasgow Cardiovascular Research Centre, University of Glasgow, Glasgow, United Kingdom, **278** Geriatric Research and Education Clinical Center, Veterans Administration Medical Center, Baltimore, Maryland, United States of America, **279** MRC Integrative Epidemiology Unit, School of Social and Community Medicine, University of Bristol, Bristol, United Kingdom, **280** Institute of Preventive Medicine, Bispebjerg and Frederiksberg Hospital, The Capital Region, Frederiksberg, Denmark, **281** R & D VitaK Group, Maastricht University, Maastricht, The Netherlands, **282** Geriatric Unit, Azienda Sanitaria Firenze (ASF), Florence, Italy, **283** Faculty of Medicine, University of Iceland, Reykjavik, Iceland, **284** Centre for Vascular Prevention, Danube-University Krems, Krems, Austria, **285** Instituto de Investigación Sanitaria del Hospital Universitario La Paz (IdiPAZ), Madrid, Spain, **286** Diabetes Research Group, King Abdulaziz University, Jeddah, Saudi Arabia, **287** Department of Public Health and Clinical Nutrition, University of Eastern Finland, Finland, **288** Research Unit, Kuopio University Hospital, Kuopio, Finland, **289** Department of Human Genetics, Radboud university medical center, Nijmegen, The Netherlands, **290** Department of Internal Medicine, University Hospital Lausanne (CHUV) and University of Lausanne, Lausanne, Switzerland, **291** Institute of Cellular Medicine, Newcastle University, Newcastle, United Kingdom, **292** The Cohorts for Heart and Aging Research in Genomic Epidemiology Consortium, **293** The DIAbetes Genetics Replication And Meta-analysis Consortium, **294** The Global Lipids Genetics Consortium, **295** The Global Blood Pressure Genetics Consortium, **296** The International Consortium for Blood Pressure, **297** The Meta-

Analyses of Glucose and Insulin-related traits Consortium, **298** Department of Nutrition, Gillings School of Global Public Health, University of North Carolina, Chapel Hill, North Carolina, United States of America, **299** Carolina Population Center, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina, United States of America, **300** National Heart, Lung, and Blood Institute, National Institute of Health, Bethesda, Maryland, United States of America, **301** NIHR Cambridge Biomedical Research Centre, Institute of Metabolic Science Addenbrooke's Hospital, Cambridge, United Kingdom, **302** University of Cambridge Metabolic Research Laboratories, Institute of Metabolic Science Addenbrooke's Hospital, Cambridge, United Kingdom, **303** Princess Al-Jawhara Al-Brahim Centre of Excellence in Research of Hereditary Disorders (PACER-HD), King Abdulaziz University, Jeddah, Saudi Arabia, **304** Finnish Institute for Molecular Medicine (FIMM), Helsinki University, Helsinki, Finland, **305** Department of Epidemiology, Harvard School of Public Health, Boston, Massachusetts, United States of America, **306** Department of Medicine, Division of Cardiovascular Medicine, Stanford University School of Medicine, Stanford, California, United States of America, **307** Department of Epidemiology and Population Health, Albert Einstein College of Medicine, Bronx, New York, United States of America, **308** Oxford NIHR Biomedical Research Centre, Oxford, United Kingdom, **309** Department of Genetics, University of North Carolina, Chapel Hill, North Carolina, United States of America, **310** Population Health Research Institute, St George's, University of London, London, United Kingdom, **311** Center for Medical Systems Biology, Leiden, The Netherlands, **312** Carolina Center for Genome Sciences and Department of Epidemiology, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina, United States of America, **313** The Genetics of Obesity and Related Metabolic Traits Program, The Icahn School of Medicine at Mount Sinai, New York, New York, United States of America, **314** The Mindich Child Health and Development Institute, The Icahn School of Medicine at Mount Sinai, New York, New York, United States of America

☉ These authors contributed equally to this work.

‡ These authors jointly supervised this work.

¶ Memberships of CHARGE Consortium, DIAGRAM Consortium, GLGC Consortium, Global-BPGen Consortium, ICBP Consortium, MAGIC Consortium are provided in [S2 Text](#).

* iris.heid@klinik.uni-regensburg.de (IMH); kari_north@unc.edu (KEN); iborecki@wustl.edu (IBB); zoltan.kutalik@unil.ch (ZK); Ruth.loos@mssm.edu (RJFL)

Abstract

Genome-wide association studies (GWAS) have identified more than 100 genetic variants contributing to BMI, a measure of body size, or waist-to-hip ratio (adjusted for BMI, WHR_{adjBMI}), a measure of body shape. Body size and shape change as people grow older and these changes differ substantially between men and women. To systematically screen for age- and/or sex-specific effects of genetic variants on BMI and WHR_{adjBMI} , we performed meta-analyses of 114 studies (up to 320,485 individuals of European descent) with genome-wide chip and/or Metachip data by the Genetic Investigation of Anthropometric Traits (GIANT) Consortium. Each study tested the association of up to ~2.8M SNPs with BMI and WHR_{adjBMI} in four strata (men $\leq 50y$, men $>50y$, women $\leq 50y$, women $>50y$) and summary statistics were combined in stratum-specific meta-analyses. We then screened for variants that showed age-specific effects (G x AGE), sex-specific effects (G x SEX) or age-specific effects that differed between men and women (G x AGE x SEX). For BMI, we identified 15 loci (11 previously established for main effects, four novel) that showed significant (FDR $<5\%$) age-specific effects, of which 11 had larger effects in younger ($<50y$) than in older adults ($\geq 50y$). No sex-dependent effects were identified for BMI. For WHR_{adjBMI} , we identified 44 loci (27 previously established for main effects, 17 novel) with sex-specific effects, of which 28 showed larger effects in women than in men, five showed larger effects in men than in women, and 11 showed opposite effects between sexes. No age-dependent effects were identified for WHR_{adjBMI} . This is the first genome-wide interaction meta-analysis to report convincing evidence of age-dependent genetic effects on BMI. In addition, we confirm the sex-specificity of genetic effects on WHR_{adjBMI} . These results may provide

further insights into the biology that underlies weight change with age or the sexually dimorphism of body shape.

Author Summary

Adult body size and body shape differ substantially between men and women and change over time. More than 100 genetic variants that influence body mass index (measure of body size) or waist-to-hip ratio (measure of body shape) have been identified. While there is evidence that some genetic loci affect body shape differently in men than in women, little is known about whether genetic effects differ in older compared to younger adults, and whether such changes differ between men and women. Therefore, we conducted a systematic genome-wide search, including 114 studies (>320,000 individuals), to specifically identify genetic loci with age- and or sex-dependent effects on body size and shape. We identified 15 loci of which the effect on BMI was different in older compared to younger adults, whereas we found no evidence for loci with different effects in men compared to women. The opposite was seen for body shape as we identified 44 loci of which the effect on waist-to-hip ratio differed between men and women, but no difference between younger and older adults were observed. Our observations may provide new insights into the biology that underlies weight change with age or the sexual dimorphism of body shape.

Introduction

Body size and shape are independent risk factors for morbidity and mortality [1–6]. They change as people grow older and these changes differ substantially between men and women [7–12]. Subtle sexual dimorphisms are already apparent during early childhood, but differences become more apparent during puberty due, at least in part, to the increasing influence of sex steroid hormones [12–14]. After puberty, sex-differences are largely maintained over the adult life-course. As women age a decline in sex steroid hormones, which coincides with menopause, affects their body shape and composition, resulting in a more android fat distribution [8, 12, 15]. When younger, women tend towards an hourglass body shape with gynoid fat distribution, storing proportionally more fat at thighs and hip than around the waist [12, 16, 17]. At a later age, often after menopause, women's fat storage shifts more upwards around the waist [12, 16, 17]. In men, changes in body fat distribution are subtler than in women, showing a slow but steady increase in waist circumference with age [12]. Thus, after the menopause, the sex-differences in body shape between men and women decrease [12].

This intricate interplay between age and sex on body size and shape is driven by underlying biological processes, involving environmental and genetic factors [7–12, 15]. Elucidating sex- and age-specific genetic effects on body size and shape may provide insights into the biological processes that are involved in the regulation of body weight and fat distribution.

More than 100 genetic loci have been identified for body mass index (BMI), a measure for body size, and for waist-to-hip ratio adjusted for BMI (WHR_{adjBMI}), a measure of body shape, most of which were identified through our own work in the Genetic Investigation of ANthropometric Traits (GIANT) Consortium [18, 19]. In a recent sex-stratified genome-wide association meta-analysis (up to 133,723 individuals in discovery stage), we searched for variants with sex-specific effects on BMI and WHR_{adjBMI} and identified several loci for which the association

with $\text{WHR}_{\text{adjBMI}}$ differed between men and women, whereas no such loci were observed for BMI [10]. However, so far, no GWAS efforts have aimed to identify genetic loci that contribute to differences in body size and shape observed in younger versus older adults, particularly across the menopausal period in women.

We conducted a genome-wide search for loci that exhibit age- and/or sex-specific differences in BMI and $\text{WHR}_{\text{adjBMI}}$. For this, we utilized study-specific genome-wide association statistics separately by sex and by two age groups in each of the studies participating in the GIANT consortium. The two age groups focus on those below and above 50 years of age, as this cut-off coincides with the average age at which women transition through menopause and experience changes in body fat distribution [20–25]. We hypothesize that genetic loci may contribute to the observed differences in body size/shape before age 50y and after age 50y, and that these differences may be sex-specific.

Results

Stratified GWAS identifies age- and sex-specific loci for BMI and $\text{WHR}_{\text{adjBMI}}$

Our total sample comprised up to 320,485 adults ($\geq 18\text{y}$) of European ancestry from 114 studies with genome-wide array data imputed to the HapMap reference or genotyped Illumina Metachip array data including up to 2.8 million autosomal variants. Details on study-specific analyses, genotyping methods and phenotypic descriptives are given in **S1–S3 Tables**. To systematically search for genetic loci that influence body size or shape in an age- and sex-specific manner, we first conducted study-specific GWA analyses for BMI and $\text{WHR}_{\text{adjBMI}}$ by four strata (men $\leq 50\text{y}$, men $> 50\text{y}$, women $\leq 50\text{y}$, women $> 50\text{y}$), and subsequently performed stratified meta-analyses (comprising up to 50,095 men $\leq 50\text{y}$, 93,201 men $> 50\text{y}$, 70,692 women $\leq 50\text{y}$, and 106,497 women $> 50\text{y}$) and derived pooled stratum-specific association results ($P_{\text{men} \leq 50}$, $P_{\text{men} > 50}$, $P_{\text{women} \leq 50}$, $P_{\text{women} > 50}$) for each trait. This strategy allowed us to test for three types of interactions: (1) SNPs that demonstrate age-specific effects (SNP x AGE, P_{agediff}), (2) SNPs that show sex-specific effects (SNP x SEX, P_{sexdiff}), and (3) SNPs that show age-specific effects that differ between men and women (SNP x AGE x SEX, $P_{\text{agesexdiff}}$). We first performed genome-wide screens using an *a priori* filter; i.e. we examined interaction effects on SNPs that showed evidence of an overall main-effect association ($P_{\text{Overall}} < 10^{-5}$). This screen is known to have better power to identify loci with age- or sex-specific effects that are directionally concordant [10, 26]. In a second screen, we examined interaction effects for all SNPs, irrespective of their main-effect association, which allows identification of loci with opposite effect direction in older vs younger adults or in men vs women.

As such, 15 loci with age-specific effects for BMI and 44 loci with sex-specific effects for $\text{WHR}_{\text{adjBMI}}$ reached significance after accounting for multiple testing (controlling false-discovery rate, $\text{FDR} < 5\%$) (**Figs 1** and **S1**). No loci were identified with evidence for three way SNP x AGE x SEX interaction.

In addition to the stratum-specific meta-analyses, we performed (a) a *main effect* meta-analysis that combined the four pooled effect estimates (one from each stratum), providing results for the *overall association* (P_{Overall}), assuming effects in age- and sex-groups are the same, and (b) a *joint (main + interaction) meta-analysis approach* (P_{joint}) allowing for simultaneous testing of overall association, SNP-by-age and SNP-by-sex interactions [27]. These two screens revealed 83 novel loci of which the association with BMI or $\text{WHR}_{\text{adjBMI}}$ reached genome-wide significance ($P < 5 \times 10^{-8}$) (**S2 Fig**). This extended discovery is enabled through power augmentation achieved by simultaneously testing main and interaction effects, and/or by accounting for potentially different effects of age and sex on the respective phenotype in the four strata.

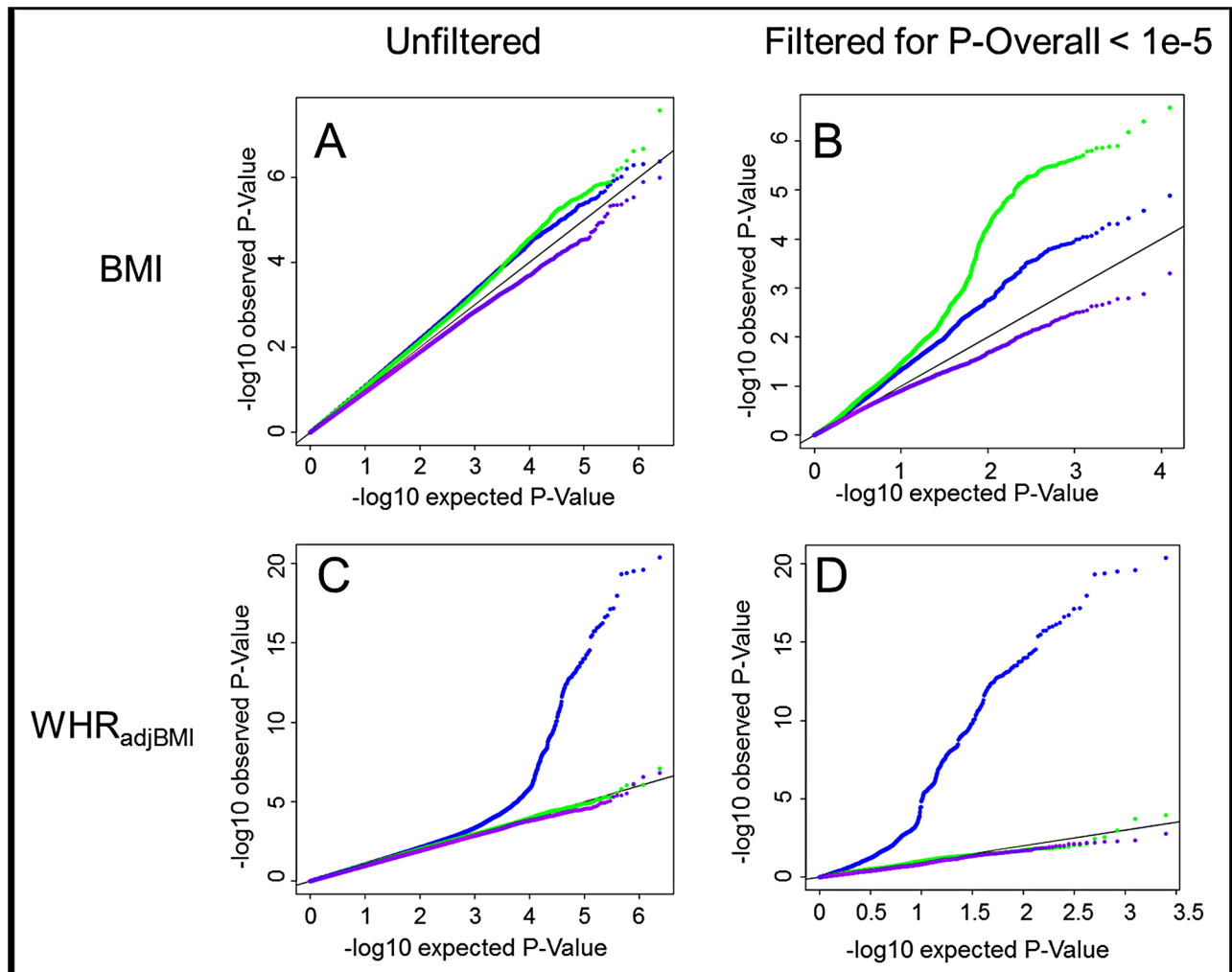


Fig 1. Interaction QQ plots. Quantile-Quantile plots showing P-Values for age-difference ($P_{agediff}$, green), sex-difference ($P_{sexdiff}$, blue) and age- and sex-difference ($P_{agesexdiff}$, purple). For BMI the P-Values are depicted for all SNPs genome-wide (A) as well as for a limited subset of SNPs that survived pre-filtering on the overall association with BMI, $P_{Overall} < 1 \times 10^{-5}$ (B). For WHR_{adjBMI} the P-Values are depicted for all SNPs genome-wide (C) as well as for a limited subset of SNPs that survived pre-filtering on the overall association with WHR_{adjBMI} , $P_{Overall} < 1 \times 10^{-5}$ (D).

doi:10.1371/journal.pgen.1005378.g001

BMI-novel loci with differential effects in younger and older individuals

Among the 15 loci with significantly different effects (at 5% FDR) on BMI in the younger versus the older individuals, four were novel (near *COBLL1*, *DDC*, *SLC22A3* and *CBLN4*) and 11 were previously established as BMI loci in large-scale *main effect* GWA meta-analyses (near *NEGR1*, *TNNI3K*, *SEC16B*, *TMEM18*, *ADCY3*, *AC016194.1*, *TCF7L2*, *STK33*, *FTO*, *MC4R*, *APOC1*) (S3 Fig and Tables 1 and S4) [19, 28]. Eleven of the 15 age-dependent BMI loci (73%, $P_{binomial} = 0.06$ for divergence from 50%) showed stronger effects in the younger than in the older group, while the four remaining loci had effects that were more pronounced in the older than in the younger group (Figs 2 and S4). We did not identify BMI-associated loci that showed effects in opposite direction between the younger versus the older group, nor did we find any sex-specific BMI effects. A sensitivity analysis excluding studies with self-report BMI found similar results (S5 Fig).

Table 1. Fifteen BMI loci showing significant age-differences in adults ≤ 50 y compared to adults >50 y. The table shows the age-group specific (sex-combined) results, ordered by largest to smallest effect in adults ≤ 50 y. All loci were detected by the screen on age-difference that included the a-priori filter on $P_{Overall} < 10^{-5}$. The age- and sex-specific results (four strata) and more detailed information on the loci are given in [S4 Table](#).

SNP	Novel Locus ^a	Nearest Gene	Chr	Pos	Alleles ^b EA/OA	EAF	Age ≤ 50 y			Age > 50 y			
							β	P	N	β	P	N	$P_{Agediff}$
rs9936385		<i>FTO</i>	16	52376670	C/T	39%	0.093	4.5E-95	115,354	0.073	1.0E-97	197,478	1.6E-04
rs2867125		<i>TMEM18</i>	2	612827	C/T	83%	0.086	6.1E-49	112,934	0.051	2.3E-30	195,579	4.0E-07
rs12955983		<i>MC4R</i>	18	56023969	G/A	28%	0.068	1.7E-41	114,448	0.038	2.0E-23	196,590	6.7E-07
rs6737082		<i>ADCY3</i>	2	24991544	C/A	47%	0.046	6.3E-20	92,191	0.022	5.4E-09	162,112	4.7E-05
rs2821248		<i>NEGR1</i>	1	72348148	A/G	83%	0.042	8.4E-12	106,067	0.017	1.9E-04	188,322	6.2E-04
rs1514174		<i>TNNI3K</i>	1	74765651	C/T	43%	0.039	3.0E-15	92,120	0.012	1.7E-03	161,764	2.8E-06
rs591120		<i>SEC16B</i>	1	176169376	C/G	20%	0.033	4.9E-14	115,337	0.014	2.8E-05	197,481	3.1E-04
rs11908421	yes	<i>CBLN4</i>	20	53813074	T/C	81%	0.033	8.7E-08	92,575	0.007	1.2E-01	162,284	4.3E-04
rs4947644	yes	<i>DDC</i>	7	50586370	T/C	51%	0.030	7.7E-10	91,980	0.009	1.7E-02	158,555	2.5E-04
rs10840060		<i>STK33</i>	11	8456621	C/A	50%	0.029	3.8E-11	110,697	0.011	2.0E-03	187,808	4.0E-04
rs1459180		Intergenic	8	77144822	G/T	58%	0.027	3.1E-09	112,913	0.009	1.6E-02	190,729	6.0E-04
rs17747324		<i>TCF7L2</i>	10	114742493	T/C	77%	0.004	4.8E-01	111,572	0.031	2.6E-13	193,773	4.7E-05
rs3127574	yes	<i>SLC22A3</i>	6	160711360	C/G	51%	0.001	7.9E-01	113,057	0.019	2.3E-08	195,472	6.8E-04
rs3769885	yes	<i>COBLL1</i>	2	165300636	A/G	48%	-0.001	9.1E-01	107,703	0.020	3.9E-09	192,513	1.1E-04
rs4420638		<i>APOC1</i>	19	50114786	A/G	82%	-0.007	3.6E-01	83,196	0.040	8.9E-12	152,014	2.1E-07

Chr: Chromosome; Pos: position; EAF: Effect Allele Frequency; EA: Effect allele; OA: Other allele

^a 'Yes' if the locus is mentioned as BMI locus for the first time

^b Effect allele is according to the BMI increasing allele according to the associated sex.

doi:10.1371/journal.pgen.1005378.t001

WHR_{adjBMI}—additional genetic loci contribute to differences between men and women

Unlike for BMI, no WHR_{adjBMI}-associated loci with significant difference between the age-groups were observed. Yet, 44 loci showed significantly different effects on WHR_{adjBMI} between women and men of which 17 loci were novel (near *TTN*, *IRS1*, *CDH10*, *IQGAP2*, *SIMI*, *ISPD*, *KLF14*, *SGCZ*, *PTPRD*, *RXRA*, *GANAB*, *SLC2A3*, *LEMD3*, *GPNPAT1*, *RPS6KA5*, *CECR2*, *HMGXB4*) and 27 loci had been previously established in *main-effect* GWA meta-analyses for WHR_{adjBMI} ([S6 Fig](#) and [Tables 2](#) and [S5](#)). Of the 27 previously established WHR_{adjBMI} loci, sex-differences had already been reported for 17 loci [[10](#), [29](#)] [[18](#)]. Our genome-wide screen established sex-specific effects for an additional 10 of the previously established loci with a main-effect on WHR_{adjBMI} (near *GORAB*, *LY86*, *ITPR2*, *PIGU*, *EYA2*, *KCNJ2*, *MEIS*, *EYA1*, *CCDC92*, *NSD1*). Of the 44 sex-specific loci, 11 loci showed opposite effect directions in women versus men and 33 showed a significant effect in one and a smaller or no effect in the other sex. Consistent with previous observations, almost all of these 33 loci (28 out of the 33, $P_{binomial} = 3.3 \times 10^{-5}$) showed more pronounced effects in women than in men ([Figs 3](#) and [S7](#)). Again, a sensitivity analysis excluding studies with self-report waist and hip circumference found similar results ([S8 Fig](#)).

No evidence for loci with simultaneous age- and sex-specific effects

We searched for loci with sex-specific effects on WHR_{adjBMI} that differ between the two age-groups and for loci with age-specific effect on BMI that differ between men and women by testing a three-way interaction (SNP x AGE x SEX, $P_{agesexdiff}$). We first tested for this three-way interaction in the 59 SNPs identified with an age-difference (15 loci for BMI) or a sex-difference (44 loci for WHR_{adjBMI}), as described above. However, none of these 59 loci showed a

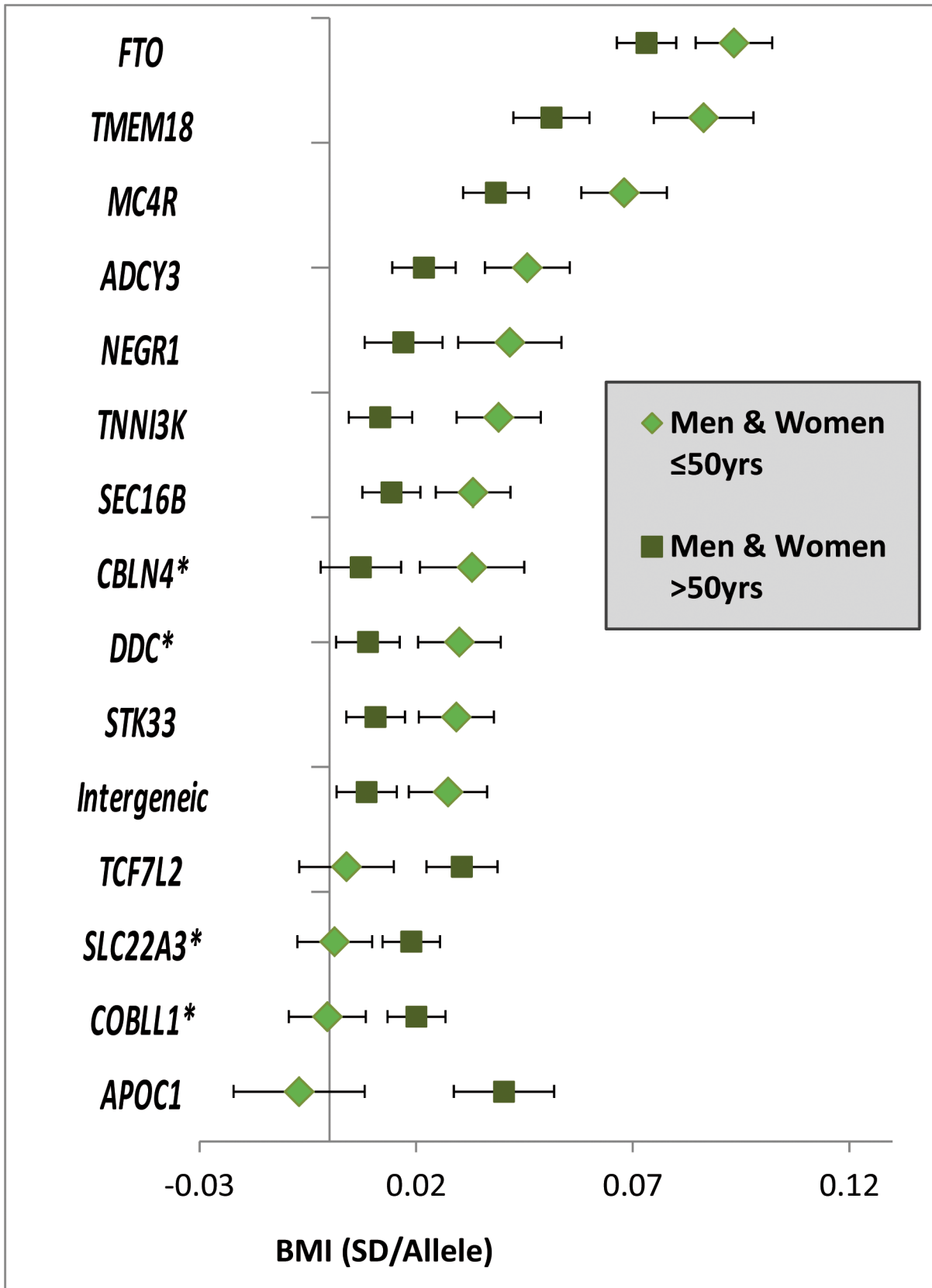


Fig 2. Age-dependent BMI loci. Effect estimates (beta \pm 95CI) per standard deviation in BMI and risk allele for loci showing age-differences in men & women \leq 50y compared to men & women $>$ 50y. Loci are ordered by greater magnitude of effect in men & women \leq 50y compared to men & women $>$ 50y. (95%CI: 95% confidence interval; BMI: body mass index; SD: standard deviation, *Newly identified loci).

doi:10.1371/journal.pgen.1005378.g002

significant three-way interaction ($P_{age \times sex \times diff} > 0.00084 = 0.05/59$, Bonferroni corrected) (**S4 and S5 Tables**). When screening for the three-way interaction genome-wide, no such loci were identified (at 5% FDR) (**Fig 1**).

Detecting loci with age- or/and sex-interaction requires extremely large sample sizes

We analytically computed the statistical power of our screens to identify SNP \times AGE, SNP \times SEX or SNP \times AGE \times SEX interaction effects, assuming a total sample size of 300,000 individuals distributed across four equally sized strata and considering a range of effect size configurations informed by previous observations (**S9, S10 and S11 Figs**). For example, for a medium genetic effect on BMI ($R^2 = 0.037\%$ as observed previously for a locus near *MAP2K5* [28]), our screens had (i) sufficient power to identify genetic loci with two-way SNP \times AGE or SNP \times SEX interactions (i.e. loci with effect in one stratum and not in the other, so-called *pure two-way interaction*, power = 86%, or loci with effect in both strata, but with opposite effect direction, power = 99%), (ii) sufficient power to detect *extreme three-way interaction* SNP \times AGE \times SEX, typically involving a biologically-unlikely scenario with opposite effect directions across both AGE and SEX (power = 99%), but (iii) insufficient power to identify loci with biologically more plausible three-way interactions (in the range of R^2 of 0.01–0.05%), i.e., loci that have an effect in only one stratum and not in the other three strata, *1-stratum interaction*, power = 2%, or those with a similar effect in three strata and not in the fourth, *3-strata interaction*, power = 21% (**Fig 4**). Identification of loci with medium 1-stratum ($R^2 = 0.037\%$ in one stratum and $R^2 = 0$ in the other three strata) or 3-strata ($R^2 = 0.037\%$ in three strata and $R^2 = 0$ in one stratum) interaction effects with a power of 80%, would require a total sample size of 750,000 or 600,000 individuals, respectively.

Reducing the multiple testing burden by applying a filter on the overall meta-analysis to first identify SNPs with main effects ($P_{Overall} < 10^{-5}$) improved the statistical power to identify loci with specific interaction scenarios: (i) loci with pure two-way interaction effects (e.g. 30% power increase to detect SNP \times AGE with $R^2 = 0.037\%$ and $R^2 = 0$ in the two strata), or (ii) loci with 3-strata interaction effects (e.g. 21% power increase for loci with $R^2 = 0.037\%$ in three strata and $R^2 = 0$ in one stratum) (**Figs 4 and S9**).

With our sample size of 300,000 subjects and equally sized strata we had 80% power to detect (i) 1-stratum interaction with $R^2 = 0.09\%$ in one stratum ($R^2 = 0$ in the other three strata), (ii) 3-strata interaction with $R^2 = 0.07\%$ in three strata ($R^2 = 0$ in one stratum), or (iii) pure two-way interaction with $R^2 = 0.03\%$ in one stratum ($R^2 = 0\%$ in the other stratum).

In summary, this analysis suggests that our study is sufficiently powered to detect even subtle two-way interaction effects, and would certainly include effect-sizes that would be considered biologically or clinically important. While even more subtle interactions may be occurring, it appears likely that in this effort, we have detected the most important age- and sex- interactions for body size and shape.

Association of identified loci with other traits

To examine whether the age- and sex-specific effects of the identified BMI and WHR_{adjBMI} loci translate into similar age- and sex-effects on obesity-related cardiometabolic traits, we gathered results from the ICBP, CHARGE and Global-BPGen consortia (age-specific and sex-specific

Table 2. Forty-four WHR_{adjBMI} loci showing significant sex-differences. The table shows the sex-specific (age-group combined) results, ordered by largest, positive effect in women to largest, negative effect in women. The age- and sex-specific results (four strata), more detailed information on the loci and on the screens for which they were detected are given in [S5 Table](#).

SNP	NovelLocus ^a	Novel Sexdiff ^b	NearestGene	Chr	Pos	Alleles ^c EAOA	Women				Men			
							EAF	β	P	N	β	P	N	P _{Sexdiff}
rs2820443			LYPLAL1	1	217820132	T/C	72%	0.063	5.2E-36	111,691	0.000	9.8E-01	93,780	1.1E-18
rs998584			VEGFA	6	43865874	A/C	48%	0.060	3.1E-32	109,533	0.015	5.0E-03	87,177	5.0E-10
rs6717858			COBLL1	2	165247907	T/C	59%	0.054	2.7E-31	110,110	-0.009	7.2E-02	90,259	4.2E-21
rs4616635			ADAMTS9	3	64677315	C/G	72%	0.049	4.0E-23	114,021	0.008	1.5E-01	93,679	7.5E-09
rs2811434			PLXND1	3	130822305	T/G	79%	0.046	8.8E-14	91,914	-0.005	4.7E-01	66,742	3.0E-08
rs1936811			RSPO3	6	127425553	T/A	61%	0.043	5.6E-17	91,862	0.015	1.4E-02	67,436	2.0E-04
rs10743579	yes		ITPR2	12	26352412	A/C	25%	0.043	1.4E-13	91,035	0.020	2.3E-03	67,178	8.7E-03
rs6958350			NFE2L3	7	25838458	T/C	25%	0.038	4.8E-14	114,759	0.016	4.9E-03	91,294	2.1E-03
rs1443512			HOXC13	12	52628951	A/C	24%	0.038	3.0E-13	114,486	0.016	5.8E-03	88,811	3.8E-03
rs7830933			NKX2-6	8	23659269	A/G	77%	0.037	4.4E-13	116,052	-0.004	5.3E-01	93,504	4.8E-08
rs11057396	yes		CDC92	12	122985015	A/C	67%	0.037	1.2E-10	78,489	0.005	4.6E-01	53,789	2.6E-04
rs1294404	yes		LY86	6	6680021	A/G	61%	0.035	3.0E-14	116,324	0.016	1.4E-03	92,668	4.3E-03
rs9687846			MAP3K1	5	55897651	A/G	19%	0.035	1.9E-09	116,005	0.000	9.8E-01	93,710	3.5E-05
rs6018158	yes		EYA2	20	44971841	T/C	41%	0.033	7.5E-11	93,476	0.012	3.9E-02	67,612	5.0E-03
rs745578	yes		EYA1	8	72628878	A/G	24%	0.033	2.9E-08	92,963	0.010	1.6E-01	67,179	9.3E-03
rs1045241			TNFAIP8	5	118757185	C/T	71%	0.031	4.8E-11	116,314	0.000	9.3E-01	93,754	3.4E-06
rs7492628	yes		RPS6KA5	14	90616889	G/C	30%	0.031	2.3E-08	91,645	0.007	2.5E-01	66,029	3.9E-03
rs17819328			PPARG	3	12464342	G/T	43%	0.031	8.5E-11	109,626	0.004	4.3E-01	88,650	7.9E-05
rs12443634			CMP	16	80081775	A/C	29%	0.031	3.2E-08	93,188	-0.009	1.8E-01	66,051	2.4E-06
rs4656767	yes		GORAB	1	168646351	A/C	71%	0.029	5.3E-09	115,682	0.006	2.8E-01	91,023	1.3E-03
rs13029520	yes		MEIS1	2	66626466	T/C	40%	0.028	3.5E-08	86,851	0.007	2.1E-01	62,091	6.9E-03
rs2092029	yes		HMGXB4	22	33982241	C/T	33%	0.028	1.4E-07	91,409	0.004	5.4E-01	63,601	2.7E-03
rs6971365	yes		KLF14	7	130083021	C/T	30%	0.027	2.8E-08	116,043	-0.006	2.6E-01	92,416	2.9E-06
rs9991328	yes		FAM13A	4	89932144	T/C	49%	0.027	1.2E-09	111,934	0.007	1.4E-01	92,564	1.7E-03
rs7917772			SFXN2	10	104477433	A/G	62%	0.027	6.2E-09	113,982	0.001	8.0E-01	90,756	1.3E-04
rs2956993	yes		GANAB	11	62162738	G/T	38%	0.026	1.9E-08	111,837	0.004	4.2E-01	90,047	1.2E-03
rs8066985	yes		KCNJ2	17	65964940	A/G	51%	0.026	5.4E-09	114,268	0.005	3.5E-01	93,518	8.0E-04
rs17185536	yes		SIM1	6	100727652	C/T	78%	0.024	4.3E-05	88,603	-0.017	1.9E-02	62,861	5.5E-06
rs9648211	yes		ISPD	7	16056277	A/G	57%	0.023	3.6E-06	93,196	-0.011	6.9E-02	67,611	6.5E-06
rs3805389			NMU	4	56177507	A/G	28%	0.023	7.1E-06	110,897	-0.013	1.9E-02	88,609	1.1E-06
rs3088050	yes		NSD1	5	176659241	A/G	21%	0.010	7.8E-02	112,933	0.036	3.0E-09	91,432	1.1E-03
rs6088735			EDEM2	20	33209337	C/T	77%	0.009	1.0E-01	114,266	0.035	6.9E-10	90,782	4.0E-04
rs7307410	yes		LEMD3	12	63828845	C/G	26%	-0.005	3.6E-01	89,227	0.033	5.0E-07	65,085	7.5E-06
rs6088552	yes		PIGU	20	32690152	G/A	37%	-0.007	1.0E-01	116,320	0.022	8.7E-06	92,396	7.2E-06
rs972303	yes		CDH10	5	24391312	T/C	75%	-0.008	1.6E-01	87,302	0.032	1.9E-06	64,371	4.1E-06
rs4898764	yes		GMPNAT1	14	52334821	G/A	53%	-0.013	2.6E-03	114,264	0.016	8.5E-04	90,762	4.2E-06
rs17470444	yes		SGCZ	8	14852373	A/G	71%	-0.014	1.4E-02	86,472	0.029	1.0E-05	61,247	4.0E-07
rs2069664	yes		IQGAP2	5	75952190	G/A	53%	-0.015	3.3E-03	88,448	0.019	9.1E-04	65,083	5.7E-06
rs741361	yes		SLC2A3	12	7966952	A/G	60%	-0.016	1.7E-03	89,766	0.022	2.7E-04	64,771	8.6E-07
rs2673140	yes		IRS1	2	226868111	G/A	38%	-0.017	2.0E-04	114,393	0.018	4.1E-04	92,271	1.7E-07

(Continued)

Table 2. (Continued)

SNP	NovelLocus ^a	Novel Sexdiff ^b	NearestGene	Chr	Pos	Alleles ^c EA/OA	EAF	Women			Men			
								β	P	N	β	P	N	$P_{Sexdiff}$
rs2042995	yes	yes	TTN	2	179266611	C/T	23%	-0.018	2.3E-03	66,222	0.022	1.4E-03	91,408	6.2E-06
rs10881574	yes	yes	RXRA	9	136345043	C/T	7%	-0.032	2.2E-03	88,999	0.045	4.0E-04	62,482	1.6E-06
rs7042428	yes	yes	PTPRD	9	8252414	A/G	98%	-0.053	4.3E-03	99,878	0.073	2.9E-04	79,410	2.5E-06
rs17809093	yes	yes	CECR2	22	16370258	G/C	4%	-0.062	8.7E-04	67,968	0.071	6.5E-04	47,894	1.1E-06

Chr: Chromosome; Pos: position; EAF: Effect Allele Frequency; EA: Effect allele; OA: Other allele

^a 'Yes' if the locus is mentioned as WHR_{adjBMI} locus for the first time

^b 'Yes' if the sex-difference in the effect on WHR_{adjBMI} is reported for the first time

^c Effect allele is according to the WHR_{adjBMI} increasing allele according to the associated sex.

doi:10.1371/journal.pgen.1005378.t002

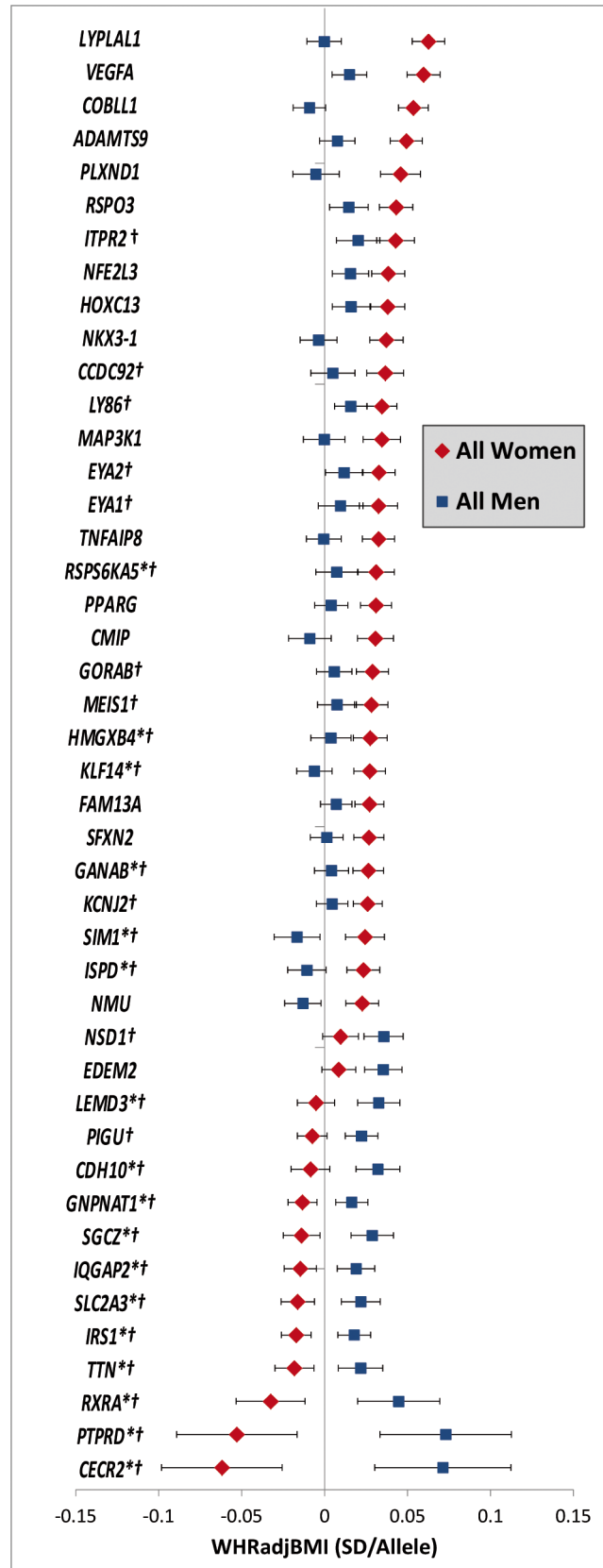


Fig 3. Sex-dependent WHR_{adjBMI} loci. Effect estimates (beta ± 95CI) per standard deviation in WHR_{adjBMI} and risk allele for loci showing sex-differences in women compared to men. Loci are ordered by greater magnitude of effect in women compared to men. (95%CI: 95% confidence interval; SD: standard deviation. *Newly identified loci. † Newly identified sex-differences)

doi:10.1371/journal.pgen.1005378.g003

effects in blood pressure) [30], Global Lipids Genetics Consortium (GLGC) (sex-specific effects in lipids) [31], DIAGRAM (sex-specific effects for type 2 diabetes) [32] and MAGIC (sex-specific effects of glycemic traits, personal communication) [33] (S6–S10 Tables). Only CHARGE, Global-BPGen and ICBP had previously performed GWAS searching for age-specific effects on blood pressure [34]. None of the 15 age-specific BMI-associated loci influenced blood pressure in an age-specific manner ($P_{SNP \times AGE} > 0.0033 = 0.05/15$) (S6 Table). Eight of the 44 sexually dimorphic WHR_{adjBMI} loci show directionally consistent female-specific effects in other traits (S10 Table), but none attained significant sex-difference ($P_{sexdiff} > 0.0011 = 0.05/44$).

In addition, we performed a systematic search in the National Human Genome Research Institute (NHGRI) GWAS Catalog (www.genome.gov/gwastudies) to examine previously reported GWAS-associations for potential age- or sex-specificity for the loci we identified for BMI and WHR_{adjBMI}, respectively [35]. While no associations have been reported that corroborate the sex- or age-specificity of our findings, largely because few sex-stratified and no age-stratified genome-wide studies have been performed to date (this study is among the first ones), many main-effect associations with a wide range of traits and disease have been reported for our age- or sex-specific BMI or WHR_{adjBMI} loci (S11 and S12 Tables). For example, the four loci that showed a larger effect in the older group are known for their association with type 2 diabetes (T2D, near *TCF7L2* and *COBLL1*) or with coronary artery disease (CAD, near *SLC22A3* and *APOC1*). The fact that disease status may correlate both with age and obesity traits may confound our age- or sex-specific findings. To reduce this possibility we repeated

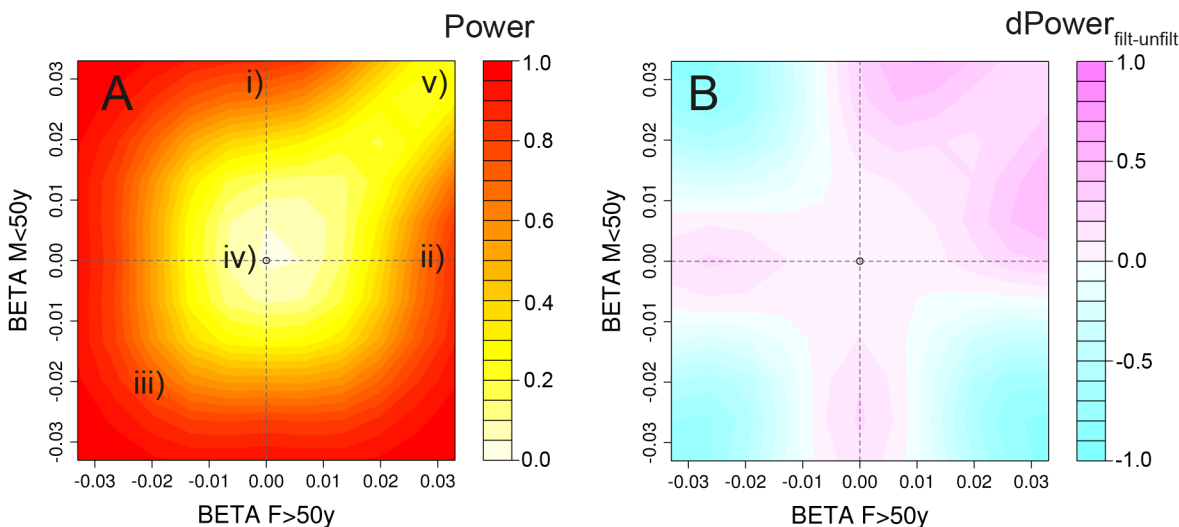


Fig 4. Power heatplots. Power for the combination of screens and gain through a priori filtering for varying configurations of effect sizes across the 4 strata. The figures illustrate (A) the power to detect age-difference, sex-difference or age-sex-difference in at least one of our scans (on $P_{agediff}$, $P_{sexdiff}$ and $P_{agesexdiff}$, with and without a priori filtering); and (B) a power comparison, comparing approaches with and without a priori filtering on $P_{Overall} < 1 \times 10^{-5}$. We here assume four equally sized strata and a total sample size of $N = 300,000$ (comparable to the sample size in our BMI analyses). We set $b_{F \leq 50y} = 0.033$ (corresponding to a known and mean BMI effect in *MAP2K5* region with $R^2 = 0.037\%$), $b_{M > 50y} = 0$, and vary $b_{F > 50y}$ and $b_{M \leq 50y}$ on the axes. This strategy allows us to cover the most interesting and plausible interaction effects: Two-way interactions, such as (i) pure age-difference ($b_{\leq 50y} = 0.033$, $b_{> 50y} = 0$) and (ii) pure sex-difference ($b_F = 0.033$, $b_M = 0$); and three-way interactions, such as (iii) extreme three-way interaction with opposite direction across AGE and SEX, (iv) 1-strata interaction ($b_{F \leq 50y} = 0.033$, $b_{F > 50y} = b_{M \leq 50y} = b_{M > 50y} = 0$), and (v) 3-strata interaction ($b_{F \leq 50y} = b_{F > 50y} = b_{M \leq 50y} = 0.033$, $b_{M > 50y} = 0$).

doi:10.1371/journal.pgen.1005378.g004

the meta-analyses restricted to population-based samples (excluding all case-control studies) and observed similar effect sizes compared to the original meta-analysis (S13 and S14 Tables).

Age-specific effects of BMI loci extend across the life course

We then examined whether the age-specific effects of the 15 BMI loci extend to younger ages and across the life course by performing look-ups in (i) a GWAS for birth weight [36] and for childhood obesity [37] from the Early Growth Genetics (EGG) Consortium, (ii) a GWAS for BMI of individuals aged 16–25 years [38], and (iii) a GWAS for weight change during adulthood (personal communication).

We found no evidence of association with *birth weight* (N = 26,836) for any of our 15 age-dependent BMI-associated loci (S15 Table) [36]. In contrast, we observed nominal significant associations with risk of *childhood obesity* (N = 13,648) for 10 of the 11 variants with stronger effect on BMI in the younger adults (Tables 3 and S16). The four loci that only showed association with BMI in the older adults were not associated with childhood obesity risk (S16 Table) [37].

Furthermore, nine of the 11 variants with stronger effect on BMI in the younger adults (18–50y) showed directionally consistent association with increased BMI in the youngest 16–25y age-group (N = 29,880, Tables 3 and S17). A more detailed experimental examination of effect sizes across the three age-groups did not reveal significant trends (S12 Fig, S17 Table, and S1 Text).

Finally, we speculated that a higher genetic BMI effect in the younger adults would translate into weight loss and a higher genetic BMI effect in the older adults would translate into weight gain with increasing age (Methods). Five of the 15 loci with age-specific effects on BMI showed a nominal significant association accompanied by the hypothesized direction on *weight change* (N = 39,041, Tables 3 and S18).

In summary, the age-dependency of the 15 loci is supported by directionally consistent enrichment of nominal significant associations ($P < 0.05$) with *childhood obesity*, with BMI in the 16–25y age-group and with *weight changes* across adulthood (P_{Binomial} ranging from 2.4×10^{-5} to 1.0×10^{-15} , Table 3).

Table 3. Enrichment analyses using look-up data for the 15 age-group specific BMI loci. The look-up data is taken from the EGG consortium for birth weight and for childhood obesity, and from personal communication for weight change trajectories. More details including SNP specific effect sizes or odds ratios and association P-Values on the look-up trait can be found in S15 Table (for birth weight), S16 Table (for childhood obesity) and S18 Table (for weight change).

Look-up data set	Sample size	#SNPs tested	#SNPs concordant with the $\leq 50y$ vs $>50y$ association pattern	P_{binomial}^a	Loci with expected association pattern
Birth weight	26,836	11	0 ^b	>0.99	-
Childhood obesity	13,648	11	10 ^b	1.0×10^{-15}	<i>FTO, TMEM18, MC4R, ADCY3, NEGR1, TNNI3K, SEC16B, CBLN4, DDC, STK33</i>
16–25y age-group	29,880	11	9 ^b	2.0×10^{-13}	<i>FTO, TMEM18, MC4R, ADCY3, NEGR1, TNNI3K, SEC16B, CBLN4, Intergenic</i>
Weight change	39,041	15	5 ^c	2.4×10^{-5}	<i>FTO, STK33, TCF7L2, SLC22A3, APOC1</i>

^a One-sided binomial P-values that test for enrichment of nominal significant and directionally consistent association in the look-up data.

^b For the BMI increasing alleles of the 11 SNPs with stronger effect on BMI in $\leq 50y$, we expect to see a nominal significant association with increased birth weight, increased risk for childhood obesity and increased BMI in the 16–25y age-group.

^c For the BMI increasing alleles of the 11 SNPs with stronger effect on BMI in $\leq 50y$, we expect to see a nominal significant association with negative effect on weight change (weight loss), and for the BMI increasing alleles of the four SNPs with stronger effect on BMI in $>50y$, we expect to see a nominal significant association with positive effect on weight change (weight gain) (see Methods for details).

eQTL analysis

eQTLs in humans. We performed sex-specific *cis* eQTL analyses in lymphoblastoid cell lines of the combined Groningen and EGCUT studies (1,450 men and 910 women) [39, 40] for the 44 SNPs showing sex-specific effects for $\text{WHR}_{\text{adjBMI}}$ to determine whether there is evidence to support sex-specific regulatory effects of the index variants on adjacent gene expression. Two SNP-gene associations displayed significant differences in genetic effects on expression between men and women ($\text{FDR}(P_{\text{Sexdiff}}) < 5\%$ with and without initial filtering on overall expression effects): rs6088552–*ACSS2* and rs6088735–*MYH7B* (S19 Table). While both SNPs were associated with $\text{WHR}_{\text{adjBMI}}$ in men-only (and no effect in women), the first SNP showed no effect on gene expression in men but was associated with gene expression in women, and the second SNP rs6088735 was associated with gene expression in both sexes, but higher in men and lower in women. The two loci were located at only 519kb from each other (rs6088552 near *PIGU*, rs6088735 near *EDEM2*, at chr20:33–34Mb, $r^2 = 0.07$), each showing independent sex-specific associations with $\text{WHR}_{\text{adjBMI}}$ and each also showing independent sex-specific association with the expression of two different genes (*ACSS2* and *MYH7B*, respectively) (S13 Fig). *ACSS2* (acyl-CoA synthetase short-chain family member 2) is a cytosolic enzyme, transcribed by SREB-proteins, that catalyzes the production of acetyl-CoA for use in both lipid synthesis and energy generation acids [41]. *MYH7B* (myosin, heavy chain 7B, cardiac muscle, beta) encodes a heavy chain subunit for slow-twitch myosin, largely expressed in heart and skeletal muscle tissue, and is involved in ATP-hydrolysis.

Age-stratified analysis were not performed for EGCUT as the study participants were relatively young (mean age: 37y), with too few individuals in the >50y age-group. Instead, we examined association between the 15 age-specific loci and gene expression using data from 3,489 unrelated individuals ($N = 2,531$ for <50y, $N = 958$ for ≥ 50 y) from the NESDA and NTR cohorts [42, 43]. No SNP showed a significant age-specific effect on gene expression ($\text{FDR}(P_{\text{agediff}}) > 5\%$ for all SNP-gene expression combinations).

eQTLs in mice. We compared expression of genes harboured by the identified loci in inguinal and gonadal fat in age-matched male, female or ovariectomized female (OVX) C57/BL6 mice maintained on a high-fat (HF) diet [44].

For genes located in the 15 age-specific BMI-associated loci, we compared expression in OVX female mice with the expression in the other male and female mice, but no differences in gene expression were observed.

For genes located in the 44 sex-specific $\text{WHR}_{\text{adjBMI}}$ -associated loci, we compared expression in female mice (OVX and non-OVX) with the expression in male mice. The expression of two genes reached significance ($P < 6.4 \times 10^{-4} = 0.05/(39 \times 2)$), corrected for testing 39 genes with homologous regions, and two tissues). The expression of *IQGAP2*, which regulates cell adhesion and motility, (rs2069664) was higher ($P = 2.3 \times 10^{-7}$) in gonadal fat tissue of male compared to female mice, whereas the expression of *TP53INP2*, a co-factor for the thyroid hormone receptor, (rs6088552) was higher ($P = 2.3 \times 10^{-6}$) in inguinal fat tissue of male compared to female mice. *TP53INP2* is located in the same chromosomal region for which we found evidence for sex-specific associations with the expression of *ACSS2* and *MYH7B* in humans. Interestingly, *Tp53inp2* has also been named the DOR (Diabetes and Obesity Related) gene, as its expression is substantially reduced in skeletal muscle of obese diabetic fa/fa Zucker rats [45]. Muscle-specific overexpression of *Tp53inp2* in mice leads to reduced muscle mass, whereas a deletion leads to muscle hypertrophy [46]. *TP53INP2* expression was markedly reduced in muscle from individuals with type 2 diabetes and in rodent diabetes models [46].

Pathway analyses

We applied pathway analyses to gain insight into mechanisms that might be involved in the age- and sex-specific difference in body size and body shape. We assumed that loci even with moderate evidence for age- or sex-difference for BMI and WHR_{adjBMI} , respectively, are enriched for genes that contribute to the age-specific BMI association or sex-specific WHR_{adjBMI} association (**Methods**). We used the DEPICT software to perform gene set enrichment and gene expression analyses [47] (**S20** and **S21 Tables** and **S1 Text**), and QIAGEN's Ingenuity Pathway Analysis (IPA, QIAGEN Redwood City, www.qiagen.com/ingenuity) tool for pathway analysis and functional annotation (**S22–S26 Tables** and **S1 Text**). Both the DEPICT and the IPA analyses identify the possible influence of sex-specific WHR_{adjBMI} loci in androgen biosynthesis, a hormone known to decrease the storage of lipids in adipose tissue [48]. Additionally, PPAR α /RXR α activation, the most significant canonical pathway for loci with a greater effect on WHR_{adjBMI} in women, may be inhibited in the presence of estrogen, thus decreasing the breakdown of lipids through competitive receptor binding [49]. To fully understand the possible age- and sex-specific regulatory effects these identified genes may have in the identified pathways, gene sets, and biological functions, further analyses are needed.

Heritability and explained variance analyses

To assess whether the age-group differences observed for BMI and the sex-differences observed for WHR_{adjBMI} extend to the contribution of all 2.5M variants (narrow-sense heritability), we calculated heritability using the GCTA method [50] in several large studies ($N =$ up to 29,232 individuals) for all, for women and men, for the younger and older adult groups. The variance explained by the 2.5M variants was 21% for BMI and 10% for WHR_{adjBMI} , with no significant difference between age groups for BMI ($P_{agediff} = 0.19$) or between men and women for WHR_{adjBMI} ($P_{sexdiff} = 0.48$) (**S27 Table**).

To further investigate differences between subgroups, we calculated the variance explained in the discovery data set for subsets of SNPs based on varying thresholds of overall association on BMI or WHR_{adjBMI} (**S14 Fig**). When we included only SNPs that reached genome-wide significance for BMI ($P_{Overall} < 5 \times 10^{-8}$), the variance explained in the younger adults (3.4%) was significantly larger than in the older (2.45%) adults. As we increased the significance threshold and included more SNPs with less significant overall association, the difference between the two age groups reduced and became non-significant once SNPs with a $P_{Overall} > 3 \times 10^{-5}$ were included. We observed similar significant differences in explained variance for WHR_{adjBMI} between men and women, with the most pronounced difference for genome-wide significant SNPs ($P_{Overall} < 5 \times 10^{-8}$, women 1.60%; men: 0.70%) that reduced and became non-significant for SNPs with a $P_{Overall} > 1 \times 10^{-5}$. Consistent with the observed interactions, we found no difference in explained variance between men and women for BMI or between the younger and the older group for WHR_{adjBMI} at any $P_{Overall}$ cut-off (**S14 Fig**).

Family-based heritability estimates, from the Family Heart Study ($N = 1,810$, 454 families), showed similar (but non-significant) trends for younger versus older adults for BMI (60% vs 45%, $P_{agediff} = 0.24$), for women and men for WHR_{adjBMI} (43% vs 38%, $P_{sexdiff} = 0.68$) (**S27 Table**).

Collectively, these observations are consistent with the results of our genome-wide search, showing that genetic variants contribute more to BMI variation in younger than in older adults and more to WHR_{adjBMI} variation in women than in men. These differences are most pronounced when we test genome-wide significant SNPs only, while differences are minimized as more SNPs with weaker associations are included.

Joint testing of main- and interaction effects yield novel loci for BMI and WHR_{adjBMI}

Our stratified analysis approach also offered an opportunity for discovery of novel variants influencing BMI and WHR_{adjBMI} by (i) using a joint 4df test of the main SNP effect in the presence of interaction [27] and (ii) by overall meta-analysis of the 4 strata. Both approaches increase statistical power to detect a main effect if there is evidence of heterogeneity across the strata. Of the 164 loci that reached genome-wide significance for BMI ($P < 5 \times 10^{-8}$), 73 are novel (S28 Table and S1, S2 and S15 Figs). Of the 73 loci, 45 were only identified in the overall test and 26 were identified in both tests. The remaining two loci were only identified in the joint test and either displayed evidence for difference between men and women (near *CXXC5*, $P_{sexdiff} = 2.7 \times 10^{-5}$) or between age-groups (near *DDC*, $P_{agediff} = 6.2 \times 10^{-4}$) suggesting that its identification may have been aided by allowing for interaction. We identified 53 loci with significant associations with WHR_{adjBMI} , of which 10 were novel (S29 Table and S1, S2 and S16 Figs). It can be speculated that the yield of novel SNP associations for BMI was greater than that of WHR_{adjBMI} , because age-dependent effects have not been sought systematically before, whereas sex-specific screens have been performed previously [10].

Discussion

Our genome-wide search for age- and sex-specific loci in up to 320,485 adults of European ancestry identified 15 loci that were associated with BMI in an age-dependent manner, with predominantly larger effects in the younger than in the older adults. Notably, despite sufficient statistical power, we did not identify BMI-associated loci with sex-dependent effects. The largest association study on BMI [19] identified two SNPs with different impact on BMI in men and women: rs543874 (*SEC16B*) and rs6091540 (*ZFP64*). While these SNPs show more modest trends towards sex-different effect ($P_{sexdiff} = 2.4 \times 10^{-4}$ and 1.3×10^{-4} , respectively) in our study, they were not picked up by our analysis due to the different pre-filtering strategy. In contrast to BMI and consistent with previous observations for WHR_{adjBMI} , we identified 44 WHR_{adjBMI} associated loci with sex-specific effects of which the majority have a larger effect in women compared with men. No age-specific WHR_{adjBMI} loci were discovered.

Our work is the first large-scale genome-wide association study to interrogate the influence of both age and sex, simultaneously, on genetic effects for BMI and WHR_{adjBMI} . While our meta-analysis had sufficient power to identify SNP-by-age or SNP-by-sex interactions, we only discovered loci influenced by age for BMI. Studies that followed up on previously established BMI loci in longitudinal and cross-sectional designs support our findings regarding the age-dependency of the majority of these loci [38, 51–57]. Indeed, for 11 of the 15 loci identified in our study, the effect on BMI was 1.5 to 3.5 times smaller in the older adults than in the younger adults, which may reflect a greater culmination of environmental and lifestyle factors on adiposity in older adults that overwhelm the genetic effects. While none of these loci were associated with birth weight, all—but one—were nominally associated with increased risk of childhood obesity. Results from a GWAS on BMI in 16-to-25 year-olds [58] provide preliminary evidence that some loci exert their largest effects relatively early in life, whereas others become more pronounced in young adulthood. Notwithstanding the predominance of BMI loci with larger genetic effects in younger individuals we identified four loci with stronger genetic effects in older adults. Interestingly, these four loci have been previously associated with either type 2 diabetes [32] or coronary artery disease [59]. Sensitivity analyses precluded potential ascertainment bias introduced by disease studies in the older group. These loci may influence BMI through mechanisms that are distinct from other BMI-associated loci; mechanisms that may be more closely related to processes more directly involved in the pathogenesis

obesity-related diseases. Furthermore, the directional consistent genetic effects of our loci on weight change during adult life from longitudinal studies supports our finding.

Indeed, the stratification into age-groups may introduce a cohort effect that implies a different genetic or environmental make-up of cohorts with older vs younger adults. For example, the obesogenic environment that has fueled the obesity epidemic that westernized societies have experienced during the past 30 years may have affected older individuals differently than younger individuals. To examine the contribution of such cohort effects and to obtain more accurate age-dependent effect estimates, large-scale genetic longitudinal studies would be required that measure BMI at multiple time points with individuals born across a wide range of birth years.

While our study provides some first insights into age-dependent genetic effects, in particular before and after menopause, more data from larger studies with longitudinal data spanning from childhood through late adulthood are desirable to accurately assess the influence of these loci on BMI across the life course. Indeed, identifying the time of life when variants affect body weight the most may help us determine the mechanisms of their influence on body weight and potential for intervention.

In contrast to the observations for BMI, our genome-wide interaction analyses did not identify loci with age-dependent effects for WHR_{adjBMI} but there was strong novel evidence for sex-influenced effects in 44 loci. For 27 of the 44 loci, the sexual dimorphism is reported for the first time, with 17 being completely novel associations for WHR_{adjBMI} . Due to increased sample size and optimized SNP selection approaches, we more than doubled the number of loci with established sex-difference for WHR_{adjBMI} [10, 18, 29]. The 44 loci divide into 11 loci with opposite effects between men and women, 28 loci with a stronger effect in women and five loci with a stronger effect in men. This is the first report to highlight loci with opposite effects and the enrichment of women-specific WHR_{adjBMI} associations is consistent with previous findings.

We examined whether the sex-dependent effects on WHR_{adjBMI} were mediated through sex-specific effects on the expression of genes located within these loci, using data available from eQTL analyses in humans and mice. Of particular interest is a region at chromosome 20q11.22 in which two independent WHR_{adjBMI} lead SNPs near *PIGU* and near *EDEM2* showed independent sex-specific associations with the expression of *ACSS2* and *MYH7B*, respectively, in humans. While we found no direct evidence of sex-specific action of *ACSS2* or *MYH7B*, based on current knowledge, both proteins seem to be involved in peripheral energy metabolism. In addition, we observed that the expression of *Tp53inp2* (Tumor Protein 53 Inducible Nuclear Protein 2), of which the human *TP53INP2* ortholog is also located in the *PIGU* locus, had significantly higher expression levels in the inguinal fat of male than female mice. This observation is consistent with a previous study, showing that *Tp53inp2* expression in white adipose tissue is significantly higher in male than in female mice [60]. The authors speculated that this sex-specificity might be due to differences in fat distribution with females storing proportionally more fat in subcutaneous/inguinal and males more in intra-abdominal depots [60]. Taken together, the sex-specific association with WHR_{adjBMI} of two independent loci at chr20q11.22 may be mediated through any or all three genes for which we found sex-specific expression. While all three genes are good candidates, experimental follow up will be needed to pinpoint the causal gene(s) and to elucidate the function and sex-specificity.

Our broad-sense (family-based analyses) or narrow-sense (GCTA including all 2.5M variants) heritability estimates showed no difference in explained variance between men or women, or between younger and older adults for either outcome. However, when considering subsets of variants displaying overall significant associations ($P_{Overall} < 1 \times 10^{-5}$), we observed a significant difference between age- but not sex- groups for BMI, with a larger explained

variance among the younger than the older adults, and between sex- but not age groups for WHR_{adjBMI} , with a larger explained variance in women than in men. These observations further corroborate the predominance of age-dependent loci for BMI and sex-dependent loci for WHR_{adjBMI} identified through a genome-wide screen.

Even though our study is likely the largest GxE and the first GxE_1xE_2 interaction GWAS meta-analysis ever conducted, we did not detect loci with sex-specific effects for BMI (SNP x SEX), age-specific effects for WHR_{adjBMI} (SNP x AGE) or three-way interactions effects (SNP x AGE x SEX). Three-way interactions are biologically plausible when considering that sex-specific effects might be exerted through hormones and that the hormonal status particularly of women changes at menopause (i.e., around the age of 50 years). This would result in a 1-stratum interaction (i.e., genetic effect only present in younger women) or a 3-strata interaction (i.e., genetic effect present in all but in younger women). While our study had sufficient power (power > 80%) to identify any kind of two-way interaction (SNP x SEX or SNP x AGE) even for effects as small as those observed for established BMI or WHR_{adjBMI} loci, our power was limited specifically for the biologically plausible three-way effects (1-stratum or 3-strata-interaction). To detect subtle effects appearing in only one of the four strata will require specialized study designs or alternative approaches. We provide a detailed analytical perspective on the power to detect different interaction signals that may inform other studies aiming at detecting interaction effects.

We acknowledge that our power estimations are expressed as a function of previously observed explained variances, incorporating measurement error. As measurement error increases, the variance of the phenotype increases and—because the genetic effect is not affected—the explained variance of the genetic variants decreases. While a random measurement error in the dependent variable of a linear regression model would not lead to a biased effect size estimate, such an error would increase the standard errors of the effect size estimates compared to a measurement error free outcome. Under the alternative hypothesis, this results in smaller statistical power. This would imply, for our analysis, that we have potentially missed some true associations, which could have been detected with smaller measurement error.

With the growing sample-size and thus statistical power, measurement error is often larger than the variant-wise effect size estimate for many human traits currently under investigation in large-scale GWAS. Thus, an individual variant's effect may not have clinical significance by itself in predictive models. However, its ultimate significance should be evaluated in the context of the biological mechanism it reveals along with other discovered variants, and the potential of such a mechanism as a therapeutic target; this is yet to be determined. In order to discover more disease-associated genetic variants, reducing measurement error by repeated and/or more accurate measurements is a viable alternative to only increasing sample size—especially when the measurement error relative to the outcome variability is high.

For technical reasons, variants on the X-chromosome were not screened. Yet, an interesting hypothesis is that sex-linked variants contribute to a sex-dependent architecture of body size and shape, both of which exhibit obvious sexual dimorphism. These analytic challenges are being addressed currently, and exploration of X-linked variation is warranted. Further, we have included only individuals of European-ancestry and thus cannot report on the generalizability of our findings to other race or ethnic groups. While we examined age-dependent effects by binning individuals below and above age 50 years—an average age of menopause—it is possible that modeling of age as a continuous trait might have had superior power. This approach poses more complex harmonization issues that should be addressed in a follow-up study. In addition, we recognize that environmental modifiers may further influence the effect of trait-related loci, and that some of the interactions we identified may be proxies for interactions with other environmental factors that are correlated with either age or sex.

In summary, our findings further distinguish the genetics of BMI from the genetics of WHR_{adjBMI} . Previously described aspects of distinction include the enrichment of neural pathways versus insulin-related pathways and sexual consistency versus sexual dimorphism, respectively [61, 62]. Our findings suggest that genetic BMI effects can change by age possibly depicting different mechanisms of genetic BMI effects that either increase or decrease during adult age. The knowledge of such mechanisms might guide the development of more effective intervention programs that are desperately sought after.

Methods

Anthropometric phenotypes

The anthropometric traits examined are body mass index (BMI, kg/m^2), which is a measure of body mass and a surrogate for total body fat, and waist-to-hip-ratio adjusted for BMI (WHR_{adjBMI}), which is a measure of body fat distribution. Traits were transformed before analyses; we first created age- (and BMI) adjusted residuals (including age and age^2 into the regression for BMI, and additionally BMI for WHR_{adjBMI}) for each of the four strata separately (men $\leq 50y$, men $> 50y$, women $\leq 50y$, and women $> 50y$) and subsequently applied an inverse normal transformation.

Study-specific analyses

We included up to 92 studies (totalling up to 21,989 men $\leq 50y$, 74,324 men $> 50y$, 41,386 women $\leq 50y$, and 88,625 women $> 50y$) with genome-wide genotyping chip data using either Affymetrix or Illumina arrays. To enable meta-analyses across different SNP panels, each study group performed genotype imputation using HapMap II CEU (build 21 or 22) via MACH [63], IMPUTE [64] or BimBam [65] yielding ~ 2.8 Million SNPs. In addition, we included 22 studies (up to 28,106, 18,877, 29,306, 17,872 individuals for each of the strata, respectively) for BMI and WHR_{adjBMI} that were genotyped using the custom iSELECT MetaboChip array containing $\sim 195K$ SNPs designed to support large-scale follow-up of putative associations with metabolic and cardiovascular traits [66].

In each study, SNP associations were tested separately by age-group and sex (men $\leq 50y$, men $> 50y$, women $\leq 50y$ and women $> 50y$) for autosomal variants. The additive genetic effect for each SNP on each phenotype was estimated via linear regression using MACH2QTL [67], SNPTTEST [64], ProbABEL [68], GenABEL [69], Merlin [70], PLINK [71] or QUICKTEST [72]. For studies with a case-control design, cases and controls were analysed separately. See [S1](#), [S2](#) and [S3 Tables](#) for study specific genotyping, imputation, analysis, quality control and phenotypic descriptive information. In total we gathered association data from up to 92 studies with imputed GWAS data and 22 studies genotyped on the MetaboChip array for BMI including up to 320,485 individuals and 64 studies with imputed GWAS data and 20 studies genotyped on the MetaboChip array for WHR_{adjBMI} including up to 216,654 individuals.

All studies were conducted according to the principles expressed in the Declaration of Helsinki. The studies were approved by the local Review Boards and all study participants provided written informed consent for the collection of samples and subsequent analysis.

Quality control of study-specific aggregated data

All study-specific files were processed in the meta-analysis centers through a standardized quality-control (QC) pipeline [73]. This involved QC checks on file completeness, range of test statistics, allele frequencies, trait transformation and population stratification as well as filtering on low quality data. Briefly, we excluded monomorphic SNPs, SNPs with $MAF \cdot N \leq 3$

(minor allele frequency multiplied by sample size), imputed SNPs with poor imputation quality: $r2_hat < 0.3$ in MACH, observed/expected dosage variance < 0.3 in BIMBAM, $proper_info < 0.4$ in IMPUTE, information < 0.8 in PLINK [64, 65, 67, 71]; genotyped SNPs with low call-rate ($< 95\%$), and genotyped SNPs that were out of Hardy-Weinberg equilibrium (HWE, P-Value testing for HWE $< 10^{-5}$). To increase the overlap in the number of SNPs between imputed GWAS and MetaboChip data, we transferred all SNP identifiers to unique SNP names consisting of chromosomal and base position, e.g. using chr1:217820132 instead of rs2820443 in the meta-analysis. Sex- and age-specific standard errors and P-values from each participating study were genomic-control (GC) corrected using study- and strata-specific lambda factors [74], whereas the lambdas were estimated from all genome-wide available SNPs for imputed GWAS and form a subset of 4,427 QT-interval SNPs for MetaboChip studies.

The meta-analyses

Generally, beta-estimates and standard errors were meta-analyzed using an inverse-variance weighted fixed effect model as implemented in METAL [75].

We meta-analyzed effect estimates and standard errors from all available studies in each of the four strata separately, yielding $b_{M \leq 50y}$, $b_{M > 50y}$, $b_{F \leq 50y}$, $b_{F > 50y}$ and $SE_{M \leq 50y}$, $SE_{M > 50y}$, $SE_{F \leq 50y}$, $SE_{F > 50y}$. By meta-analyzing $b_{M \leq 50y}$ and $b_{M > 50y}$ we obtained the effect and standard error for men (b_M, SE_M) and women (b_F, SE_F). Similar meta-analyses yielded the age group-specific association statistics, $b_{\leq 50y}$ and $b_{> 50y}$ with standard errors $SE_{\leq 50y}$ and $SE_{> 50y}$. Meta-analysis of all four strata provided the overall association effect estimate $b_{overall}$, standard error $SE_{overall}$ and P-value $P_{overall}$. A joint meta-analysis based on the pooled stratum-specific estimates was performed according to Aschard et al [27].

After the meta-analyses, we performed an additional quality control step on the meta-analytic results: We only included SNPs (i) being available in at least half of the maximum sample size in all strata; and (ii) having chromosome and position annotation in dbSNP.

Genome-wide screening approaches to detect interaction effects

Our study aimed at discovering SNPs with (1) *age-different* effects, (2) *sex-different* effects, and (3) *age-dependent sex-different* effects or *sex-dependent age-different* effects.

To find *age-different* genetic effects, we computed age-difference P-values ($P_{agediff}$) by testing for difference between the age group-specific meta-analyzed beta-estimates $b_{\leq 50y}$ and $b_{> 50y}$ using

$$t_{age} = \frac{b_{\leq 50y} - b_{> 50y}}{\sqrt{SE_{\leq 50y}^2 + SE_{> 50y}^2 - 2r_{age} \cdot SE_{\leq 50y} \cdot SE_{> 50y}}}$$

The correlation r_{age} between $b_{\leq 50y}$ and $b_{> 50y}$ computed as the Spearman rank correlation coefficient across all SNPs for BMI and WHR_{adjBMI} was 0.123 and 0.049, respectively. The analogous test statistic for *sex-different* effects was

$$t_{sex} = \frac{b_M - b_F}{\sqrt{SE_M^2 + SE_F^2 - 2r_{sex} \cdot SE_M \cdot SE_F}}$$

with corresponding P-value ($P_{sexdiff}$). The Spearman correlation r_{sex} was 0.121 or 0.047 for BMI and WHR_{adjBMI} , respectively.

To test for the three-way interaction of age- and sex-differences, we introduced for the first time a test of difference between age groups in the sex-difference, which is mathematical equivalent to a test of difference between sexes in the age group-difference using the age-sex-

difference statistic as

$$t_{agesex} = \frac{(b_{M \leq 50y} - b_{F \leq 50y}) - (b_{M > 50y} - b_{F > 50y})}{\sqrt{SE_{M \leq 50y}^2 + SE_{F \leq 50y}^2 + SE_{M > 50y}^2 + SE_{F > 50y}^2}},$$

with the corresponding P-value ($P_{agesexdiff}$).

To maximize statistical power we did not split our samples (artificially) into discovery and replication sets, but meta-analyzed all studies together and verified the absence of cross-study heterogeneity. We screened genome-wide for $P_{agediff}$, $P_{sexdiff}$, and $P_{agesexdiff}$ for each of the two traits (BMI, WHR_{adjBMI}). These screens have ideal power to detect effects that are of opposite direction across the four strata (S9 Fig). However, searching for effects that are prominent in one or some strata, but not existent or directionally consistent and less pronounced in other strata profits from an a priori filter on the overall association ($P_{overall} < 10^{-5}$) as shown previously [10, 26] (Fig 4). The rationale behind this filter is that SNPs with unequal effects in the different strata have non-zero overall effect when tested in all strata combined. This is true unless these effects are the same magnitude, but in opposite direction (i.e. cancel out in the combined analysis). Hence filtering on overall association P-value possibly enriches our selection with SNPs showing interaction effects. For BMI and WHR_{adjBMI} 7,382 and 2,014 SNPs passed this filter.

For each trait and for each of the 6 approaches ($P_{agediff}$, $P_{sexdiff}$, $P_{agesexdiff}$, with and without a priori filtering), we controlled the False Discovery Rate (FDR) at 5% to account for the multiple testing [76]. Importantly, controlling the FDR of each single analysis at 5% implies a global FDR control at 5% for the ensemble of discoveries resulting from all the different approaches together.

Sensitivity analyses using population-based studies only

To ensure the association of none of our age- or sex-specific loci were driven by ascertainment bias through inclusion of case-series of individuals with type 2 diabetes or coronary artery disease, we performed additional meta-analyses restricted to population-based (i.e. no ascertainment bias) studies and compared the effect-sizes between the original meta-analyses and the meta-analyses restricted to population-based studies.

Sensitivity analyses excluding studies with self-reported BMI or WHR

Self-reported BMI or WHR may cause systematic measurement error that might lead to biased effect estimates. Few of our studies assessed BMI and WHR by self-report in the sense that they told study participants how to measure BMI and WHR for themselves. In order to ensure that the age- or sex-differences of our identified loci was not driven by the few studies that used self-report data (13 of our 114 studies), which may introduce bias [77–79], we conducted sensitivity meta-analyses limited to studies that measured anthropometric phenotypes (S5 and S8 Figs).

Power computations

To illustrate the strength and characteristics of the various screens outlined, we analytically computed power by scan (S9 Fig) and for all scans combined (Figs 4, S10 and S11), for varying configurations of effect size combinations and directions across the four strata. More specifically, we assumed equally sized strata, a total sample size approximately corresponding to the maximum sample size of our study and modelled three categories of SNPs explaining realistic fractions of the phenotypic variance, i.e. small, medium and large effects from Sneliotes et al

[28] and from Heid et al [29]. The power shown in any of the heatplots was calculated based on a fixed effect in women ≤ 50 y (set to the known effect), a fixed effect in men > 50 y (set to 0), and varying effects in women > 50 y and men ≤ 50 y (varying from negative to positive magnitude of the known effect). This strategy allowed us to depict power for most important interaction effects (i.e. for pure sex-difference, pure age-difference, 1-strata interaction and 3-strata interaction) in a single heatplot (see legend of Fig 4).

Genome-wide screening approaches to detect main effects accounting for interaction

To identify novel genetic association for BMI and $\text{WHR}_{\text{adjBMI}}$, we screened (i) the P_{Overall} gathered from a four-way meta-analysis of the stratified results and (ii) the P_{Joint} gathered from a four-way joint meta-analysis of the stratified results according to Aschard et al [27]. We used a genome-wide significance level ($P < 5 \times 10^{-8}$) for both approaches to correct for the multiple testing and compared the detected regions to previously established loci using a 500kb distance criterion.

Establishing enrichment for sex-specific or age-dependent genetic effects

For $\text{WHR}_{\text{adjBMI}}$, we counted among the sex-different associations (disregarding the opposite effect loci) how many were significantly stronger in men or women. To test whether the observed counts represent significant imbalances between sexes we compared them to the expected binomial distribution (with $p = 0.5$). Similar exercise was done for age-specific associations for BMI.

Lookup of age- and sex-specific associations with other phenotypes

Age-group specific association results of the identified loci were requested for blood-pressure measures (diastolic and systolic blood pressure, mean arterial pressure and pulse pressure) from the Global-BPGen consortium [30]. The provided effect size and standard error estimates for six age bins (20–29, 30–29, . . . , 70–79 years) were combined to derive SNP x AGE interaction effect sizes and P-Values (S6 Table) using meta-regression [34].

Sex-specific associations of the identified loci were requested for lipid traits (HDL-C, LDL-C, Total Cholesterol and Triglycerides) from the Global Lipids Genetics Consortium [31], for type 2 diabetes (T2D) from the DIAGRAM consortium [32], for glycemic traits (fasting insulin, fasting glucose, HOMA-B, HOMA-IR) from the Meta-Analyses of Glucose and Insulin-related traits (MAGIC) Consortium [33] (personal communication), and for blood-pressure measures (diastolic and systolic blood pressure) from the Global-BPGen consortium [30] (S7, S8 and S9 Tables). The provided men- and women-specific estimates were used to derive sex-difference P-Values.

NHGRI GWAS catalog lookups

To further investigate the identified genetic variants in this study and to gain additional insight into their functionality and possible pleiotropic effects, we searched for previous SNP-trait associations nearby our lead SNPs. PLINK was used to find all SNPs within 500 kb of any of our lead SNPs using 1000 Genomes Project Pilot I genotype data from the CEPH (Utah residents with ancestry from northern and western Europe) population (CEU) [80, 81]. To identify previous associations, all SNPs within the specified regions were compared with the NHGRI (National Human Genome Research Institute) catalog for overlap and distances between the

two SNPs were obtained using SAS, Version 9.2 [citation info below for SAS and PLINK] [82]. The NHGRI's (National Human Genome Research Institute) GWAS catalog contains only the top 30 most significant SNP-trait associations from recent GWAS published results from studies with at least 100,000 SNPs with resulting P-values of less than $P < 1 \times 10^{-5}$ [82]. For previous GWAS results not reported in the Catalog when accessed on 10/15/2014, additional SNP-trait associations were pulled from the literature and compared to our lead SNPs using the same PLINK output file to obtain distance and r^2 values [83–91]. All previous associations within 500 kb and with an $r^2 > 0.1$ with our lead SNP that reached genome-wide significance in the previous publication were retained for further interrogation.

Association of age-specific BMI loci with birth weight and childhood obesity

Summary statistics from a genome-wide association meta-analysis previously performed by EGG Consortium (www.egg-consortium.org) were used to examine whether the 15 age-specific BMI loci associate with birth weight and/or childhood obesity risk. Birth weight (BW) had been transformed to z-scores. Association between each SNP and the birth weight was tested using linear regression assuming an additive genetic model, with sex and, where available, gestational age as covariables [36]. In the genome-wide association meta-analysis for childhood obesity risk, cases were defined as having an age- and sex-specific BMI > 95th percentile, and controls as having an age- and sex-specific BMI < 50th percentile in children of European ancestry. SNP associations were assessed in a case-control design assuming an additive genetic model [37].

Comparison of effect sizes for age-dependent BMI loci with younger individuals aged 16–25 years

We compared the effect sizes for 15 loci with age related differences in BMI for each of the age strata (≤ 50 y and > 50 y) in men and women combined with the BMI in young adults ages 16–25 years [58]. Nine out of the 14 studies included in the young adult analysis had overlapping samples with the current sample, although the BMI measurements utilized were different (i.e. adolescence/early adulthood versus middle-aged to older adulthood). We used t-tests to compare effect estimates (β) from the younger adults aged 16–25 years (A) to each of our age strata (≤ 50 y or > 50 y) (B) adjusting for the correlation due to overlapping samples such that:

$$t_{diff} = \frac{b_A - b_B}{\sqrt{SE_A^2 + SE_B^2 - 2r \cdot SE_A \cdot SE_B}},$$

where SE = standard error and r = Spearman correlation coefficient between the effect estimates genome-wide. We calculated the Spearman correlation r between our study and GIANT using the combined stages from both studies. The significance level (P-value) was based on a two-tailed t-test.

Look-up of age-dependent BMI loci for weight change across adulthood

We also evaluated the 15 BMI loci showing age-dependent results from genome-wide analyses with weight change across adulthood. Using growth curves generated from multiple measures of weight in individuals between the ages of 20 and 65 years, weight change trajectories were calculated by sex using age as both a random and fixed effect. For each of the 15 loci showing age-differences in BMI, we observationally compared the direction of the effect estimate in the weight change results with the direction of effect seen between our adults aged 18–50 years and

adults >50 years. While assuming constant height across adulthood and no cohort effect between the two age-groups, we hypothesized that for loci where we find a stronger effect for BMI in the adults ages 18–50 years compared to adults >50 years, the direction of effect estimate in the weight change data would be negative. For the loci where we found a stronger effect for BMI in the adults >50 years compared to the adults ages 18–50 years we hypothesized that the direction of effect estimate in the weight change data would be positive.

Expression QTL analyses in human tissue

We examined transcript expression of genes nearby (\pm 1 Mb) the 44 identified WHR_{adjBMI} SNP in lymphoblastoid human cell lines available in 2,360 human samples from the EGCUT and Groningen cohorts (910 women and 1,450 men) [39, 40]. We computed sex-specific associations between each of the 44 variants and all genes in their 1 Mb vicinity and tested the men- and women-specific eQTLs for sex-difference ($FDR_{sexdiff} < 5\%$ calculated with/without initial filter on overall expression effect $FDR_{overall} < 20\%$).

We next examined whether the 15 SNPs identified to be age-dependently associated with BMI impact nearby (\pm 1 Mb) transcripts differently in younger (<50y) than in older individuals (\geq 50y). As such, we analyzed human whole blood transcription in 3,489 unrelated individuals from NESDA and NTR cohorts [42, 43], which were divided in a \geq 50y group (N = 958) and a <50y group (N = 2,531). Cis-eQTL analysis for the 15 SNPs was conducted for the two groups separately and age-group specific eQTLs were compared for age-difference ($FDR_{agediff} < 5\%$ calculated with/without initial filter on overall expression effect $FDR_{overall} < 20\%$).

Expression QTL analyses of adipose tissues in high-fat-diet-induced obese mice

We performed a microarray analysis on data from an experiment previously published [44]. Briefly, 21 male, 21 female, and 21 ovariectomized (OVX) female C57/BL6 mice were fed from day 21 for 12 weeks on an high fat diet (45% calories from fat; Research Diets, Inc., New Brunswick, NJ). All mice (male, female, and OVX) were exposed to sham or OVX surgery. Animals were sacrificed and tissues collected during the first 2h of the beginning of the light cycle after a 12h fast.

GeneChip microarray (Affymetrix, Santa Clara, CA) was performed according to manufacturer's instructions on 7 independent pooled samples (3 mice per pooled sample) per experimental group (male, female, OVX) from gonadal adipose tissue (GWAT) and inguinal adipose tissue (IWAT) fat pads. AMC Project Report Version 12 (6/27/07) GeneChip Operating System parameters α_1 and α_2 were set to 0.05 and 0.065, respectively. Normalized expression values from the Affymetrix identifier were analyzed with the online software server Genesifter (VizX Labs, Inc., Seattle, WA, USA). For comparisons of microarray data sets, multiple t-tests were used to identify genes with at least a twofold difference in gene expression (with Benjamini and Hochberg correction; $P < 0.05$) and at least an expression level of 100. Genes populated from the GWAS studies were compared to this list of genes that met the minimum criteria of expression, fold difference, and p-value. Those identified as being statistically significant were further validated by qPCR.

Pathway analyses

DEPICT. We used a recently developed pathway enrichment method, DEPICT [47]. The methodology first selects all lead SNPs below a certain threshold with respect to a target P-value (available genome-wide). We tested multiple hypotheses corresponding to different

lead SNP selection scenarios. First, we selected SNPs with $P_{sexdiff} < 0.001$. Second, focusing on SNPs with concordant effect size direction (CED), but different magnitude we added a marginal filter to boost power by selecting SNPs with $P_{sexdiff} < 0.01$ and $P_{overall} < 0.01$. In case of CED, SNPs with stronger effect in women may fall into separate pathways from SNPs with stronger effect in men. Hence, we have derived gender-specific sex-difference P-values ($P_{sexdiff_F}$, $P_{sexdiff_M}$). We then looked for women-specific pathway enrichment by selecting SNPs with $P_{sexdiff_F} < 0.01$ and $P_{overall} < 0.01$ (given the CED framework). Similarly, we created a separate list for men-specific SNPs by a filter of $P_{sexdiff_M} < 0.01$ and $P_{overall} < 0.01$. All above lists were also created for age-dependent BMI associations by replacing $P_{sexdiff}$ by $P_{agediff}$, $P_{sexdiff_F}$ by $P_{agediff_younger}$ and $P_{sexdiff_M}$ by $P_{agediff_older}$.

For each of the eight SNP lists, lead SNPs were identified. For each lead SNP locus a target region is defined as the smallest interval containing all SNPs with $LD > 0.5$ with the lead SNP of the locus. All genes encompassed in the target regions represent the “GWAS genes”, thereby assuming that either the lead SNP is in LD with a functional coding SNP within a gene or that the lead SNP marks a cis-acting regulatory region. We then used the following pre-defined gene sets and pathways: Gene Ontology (GO), Reactome, InWeb protein complexes, Mouse knock-out phenotypes. Gene sets were re-annotated based on co-expression in a large collection (80,000) of gene expression compendium from GEO. Then, for each gene-set the pairwise similarity between GWAS genes was calculated and compared to that of matching sets of non-GWAS genes to assess significance of enrichment.

DEPICT also generated a prioritized set of genes at each locus. Briefly, genes within associated loci that are functionally similar to genes from other associated loci are the more likely candidates. DEPICT prioritizes genes in three steps: gene scoring, bias adjustment, and false discovery rate estimation. In the scoring step, the method quantifies the similarity of a given gene to genes from other associated loci. The bias adjustment step controls confounding factors that may bias the gene scores, e.g. gene length. In the last step, experiment-wide false discovery rates are estimated by repeating the scoring and bias adjustment steps 20 times based on top SNPs from pre-computed null GWAS.

Ingenuity Pathway Analysis (IPA). Significantly associated loci were further explored using QIAGEN’s Ingenuity Pathway Analysis (IPA, QIAGEN Redwood City, www.qiagen.com/ingenuity) to determine if there was an over-arching functional or disease relationship among these loci and their associated genes using the age and sex specific SNP lists described above. IPA uses publicly available databases (e.g. NHGRI GWAS Catalog, NCBI databases, KEGG) and proprietary databases of gene/protein interaction, expression, and function to identify possible pathways, networks, and overlapping functions of genes. For our analysis, IPA identified potential genes as those genes with coding regions within 2kb upstream or 0.5kb downstream of our list of input dbSNP ids that can unambiguously be mapped to these ids. To perform the analyses, only Ingenuity Knowledge Base genes were used, both direct and indirect relationships that are observed or predicted in mammals (humans, mice, and rats) are strictly considered. All canonical pathways and functional/disease categories and annotations that were statistically significant ($P < 0.05$ using the Fisher’s exact test) are reported; however, those that meet significance for multiple test correction (Benjamin-Hochberg corrected $P < 0.05$) are highlighted in the table. Only the top ten predicted networks containing up to 140 genes or endogenous molecules were requested. Only those networks with a score of greater than 2 (Fisher’s Exact Test result of $P < 0.01$) are considered significant [92].

Estimation of heritability

We estimated the broad heritability (H^2) of BMI and $\text{WHR}_{\text{adjBMI}}$ within the Family Heart Study (FHS) to assess how much of each trait's total phenotypic variance may be genetic. A random sample of 1,810 individuals (454 families) was used for this analysis. The sample was stratified by age and sex into 9 groups (all, all ≤ 50 y, all > 50 y, men, women, men ≤ 50 y, men > 50 y, women ≤ 50 y, women > 50 y) to assess how each trait's genetic variance may differ across strata. Within each group, BMI and $\text{WHR}_{\text{adjBMI}}$ were adjusted for age, age², genotyping chips (Illumina 560K, 1,000,000K, 610K), 10 principal components and 3 study centers. Residuals for BMI and $\text{WHR}_{\text{adjBMI}}$ were ranked and an inverse normal transformation was applied. Subsequently, SOLAR was used to estimate the H^2 of BMI and $\text{WHR}_{\text{adjBMI}}$ within each group ([S28 Table](#)).

Genome-wide Complex Trait Analysis for proportion of variance explained

To explore the contribution of all common (genotyped) SNPs genome-wide to each trait of interest, BMI and $\text{WHR}_{\text{adjBMI}}$, we estimated the variance explained by all the autosomal SNPs in the combined ARIC, KORA S3/S4, CoLaus, EGCUT and SHIP studies within each of the sex and age strata, using the method proposed by Yang et al [93] and implemented in the Genome-wide Complex Trait Analysis software package (GCTA <http://www.complextaitgenomics.com/software/gcta/>). Each phenotypic trait was transformed in the same form as was used for all meta-analyses.

Estimation of explained variance

We estimated the age-group and sex-specific polygenic variance explained by various subsets of SNPs that were based on varying thresholds of overall association (P_{Overall}) with BMI or $\text{WHR}_{\text{adjBMI}}$. First, each subset of SNPs was clumped into independent regions using a physical distance criterion < 500 kb and for each region the most significantly overall associated SNP (i.e. top SNP) was taken further. For each top SNP, the explained variance was calculated according to

$$r^2 = \frac{1}{1 + \frac{N}{(\Phi^{-1}(\frac{\alpha}{2}))^2}} - \frac{1 - r^2}{N}$$

for each age-group and for each sex separately [94]. Finally, the variance explained by the subset of SNPs was obtained by summing up the single SNP-specific explained variances. The overall association threshold was varied from 1×10^{-8} to 0.1.

Search for biological and functional knowledge of the identified association regions

We examined whether SNPs known to provide reliable tags for Copy-Number-Variations (CNVs) in subjects of European-descent (combining four catalogues including 60,167 CNV-tagging SNPs as described previously [95]) correlated with our lead SNPs. We also performed several online database searches to establish whether known variants within a 500kb-window on both sides of each lead SNP, that are in high linkage disequilibrium ($r^2 > 0.8$) with our lead SNPs (using SNAP Proxy search [96]), might have putative or predicted function. (i) We searched the SIFT database [97] to determine whether any of these SNPs were predicted to affect protein function. (ii) We used Annovar [98] to investigate predicted and putative

function in several functional classes, including splicing regulation, stop codons, polyphen predictions. (iii) We used the regulome database (<http://regulome.stanford.edu/>) to search for known and predicted regulatory elements (DNAase hypersensitivity, binding sites of transcription factors, and promoter regions) in the intergenic regions of our age-specific BMI and sex-specific WHR_{adjBMI} loci. Additionally, we searched for estrogen, androgen or progesterone receptor motifs around our sex-specific WHR_{adjBMI} loci. Source of these data include public datasets from GEO, the ENCODE project, and published literature [99].

Supporting Information

S1 Fig. Workflow and overview of results. The numbers stated are the number of identified independent loci for the respective analysis. Given in brackets is the number of the identified loci that are novel loci for the trait, i.e. have not been previously reported for association with the trait.

(TIF)

S2 Fig. QQ plots for overall association and joint test P-Values for both traits. QQ-plots for BMI (A) and WHR_{adjBMI} (B) depicting overall association P-Values (red) and joint test P-Values (blue) for all SNPs and after excluding previously published BMI or WHR_{adjBMI} associated regions ($P_{Overall}$: magenta; P_{Joint} : cyan).

(TIF)

S3 Fig. Locuszoom plots for 15 loci associated with BMI that are different between men and women ≤ 50 y and men and women > 50 y. Each plot highlights the most significant SNP for age-differences and illustrates p-values for age-differences ($P_{agediff}$), sex-differences ($P_{sexdiff}$), all strata combined ($P_{Overall}$), and the joint test (P_{Joint}). The figure is sorted according to [Table 1](#). The plots are based on GrCh37 build positions and annotations.

(TIF)

S4 Fig. Scatterplot of effect estimates (beta) for loci showing age-differences in BMI, contrasting loci with larger effect estimates in men & women ≤ 50 years (light green diamonds) and loci with larger effects in men & women > 50 years (dark green squares).

(TIF)

S5 Fig. Sensitivity meta-analysis for the 15 age-specific BMI loci-excluding 13 studies that used self-report data for BMI and comparing the age-difference effects to the originally observed age-difference.

(TIF)

S6 Fig. Locuszoom plots for 44 loci associated with WHR_{adjBMI} that are different between men and women. Each plot highlights the most significant SNP for sex-differences and illustrates p-values for age-differences ($P_{agediff}$), sex-differences ($P_{sexdiff}$), all strata combined ($P_{Overall}$), and the joint test (P_{Joint}). The figure is sorted according to [Table 2](#). The plots are based on GrCh37 build positions and annotations.

(TIF)

S7 Fig. Scatterplot of effect estimates (beta) for loci showing sex-differences in waist-to-hip ratio adjusted for BMI (WHR_{adjBMI}), organized by loci with larger effect estimates in women compared to men (red circles), larger effect estimates in men compared to women (blue squares) and opposite effect estimates between men and women (green triangles).

(TIF)

S8 Fig. Sensitivity meta-analysis for the 44 sex-differential WHR_{adjBMI} loci—excluding two self-report studies and comparing the sex-difference effects to the originally observed sex-difference.

(TIF)

S9 Fig. Power by AGE x SEX scan. The figures illustrate the power of scanning $P_{sexdiff}$ (A: unfiltered, B: pre-filtered on $P_{Overall}$), $P_{agediff}$ (C: unfiltered, D: pre-filtered on $P_{Overall}$), and $P_{agesexdiff}$ (E: unfiltered, F: pre-filtered on $P_{sexdiff}$ or on $P_{agediff}$). We assume four equally sized strata, a total sample size of $N = 300,000$ (comparable to the sample size in our BMI analyses). To investigate varying scenarios of interaction effects, we set (i) $b_{F<50y} = 0.033$, a median BMI effect near *MAP2K5* from Speliotes et al. ($R^2 = 0.037\%$), (ii) $b_{M>50y} = 0$, and (iii) vary $b_{F>50y}$ and $b_{M<50y}$ on the x- and y-axes respectively.

(TIF)

S10 Fig. Power of the AGE x SEX approaches for BMI for varying allele frequencies and varying modelled effect sizes. The figure shows the power to detect age-difference, sex-difference or age x sex-difference in at least one of our scans and for varying scenarios of effect size combinations between the 4 strata. We assume four equally sized strata and a total sample size of $N = 300,000$ (comparable to the sample size in our BMI analyses). Furthermore, for each plot we (i) set $b_{F<50y}$ to a known BMI effect sizes from Speliotes et al. paper (using a small (*PTPB2*), medium (*NEGR1*) and the largest (*FTO*) effect size), (ii) set $b_{M>50y} = 0$, and (iii) vary $b_{F>50y}$ and $b_{M<50y}$ on the axes.

(TIF)

S11 Fig. Power of the AGE x SEX approaches for WHR_{adjBMI} for varying allele frequencies and varying modelled effect sizes. The figure shows the power to detect age-difference, sex-difference or age x sex-difference in at least one of our scans and for varying scenarios of effect size combinations between the 4 strata. We assume four equally sized strata and a total sample size of $N = 200,000$ (comparable to the sample size in our WHR_{adjBMI} analyses). Furthermore, for each plot we (i) set $b_{F<50y}$ to a known WHR_{adjBMI} effect sizes from Heid et al. paper (using a small (*CPEB4*), medium (*LYPLAL1*) and the largest (*RSPO3*) effect size), (ii) set $b_{M>50y} = 0$, and (iii) vary $b_{F>50y}$ and $b_{M<50y}$ on the axes.

(TIF)

S12 Fig. Differences in effect estimates ($\beta \pm SE$) between young adults, adults $\leq 50y$, and adults $> 50y$ for BMI loci selected for age-differences. Loci are ordered according to trends in absolute magnitude of effect: 1) where the absolute magnitude of effect is largest in adolescent/youngest adults (ages 16–25y)¹, 2) where absolute magnitude is largest in adults ($\leq 50y$), and 3) where absolute magnitude is largest in older adults ($> 50y$). BMI: Body mass index; SE: standard error; Details for men and women ages 16–25 have been described elsewhere (Graff et al.: “Genome-wide analysis of BMI in adolescents and young adults reveals additional insight into the effects of genetic loci over the life course.” Human Molecular Genetics 2013).

(TIF)

S13 Fig. The most significant SNPs, rs6088552 and rs6088735, for sex-differences with WHR_{adjBMI} each identified to be a sex-different cis-eQTL for the *ACSS2* and *MYH7B* genes, respectively, on chromosome 20. WHR_{adjBMI} : waist-to-hip ratio adjusted for body-mass index; eQTL: expression quantitative trait loci. Sex-specific associations were computed to identify cis eQTL signals that were likely to be coincident with the WHR_{adjBMI} using human eQTL in lymphoblastoid cells.

(TIF)

S14 Fig. Total stratum-specific explained variance by SNPs meeting varying thresholds of overall association for BMI (A: sex-specific; B: age-group specific) and for WHR_{adjBMI} (C: sex-specific; D: age-specific).

(TIF)

S15 Fig. Locuszoom plots for 73 novel loci associated with BMI that were either identified by the joint 4df test or by the overall (age-group and sex—combined) analysis. Each plot highlights the most significant SNP for the combined effect ($P_{Overall}$) or for the joint test (P_{Joint}) and illustrates p-values for age-differences ($P_{Agediff}$), sex-differences ($P_{Sexdiff}$) and P_{Joint} or $P_{Overall}$ respectively^a. The figure is sorted according to chromosome and position. The plots are based on GrCh37 build positions and annotations. For three loci we identified two different SNPs that met the significance threshold for the scan of $P_{Overall}$ and P_{Joint} . For each set we plotted the SNP with the lowest P-value based on the scan it was identified for. These loci and the SNP plotted are as follows: 1) rs7421089 – Selected for P_{Joint} and rs10804189 – Selected for $P_{Overall}$ ->rs10804189 is plotted, 2) rs1557765 – Selected for $P_{Overall}$ and rs7928810 – Selected for P_{Joint} -> rs7928810 is plotted, and 3) rs11181001 – Selected for P_{Joint} & rs1405552 – Selected for $P_{Overall}$ -> rs1405552 is plotted.

(TIF)

S16 Fig. Locuszoom plots for 10 novel loci associated with WHR_{adjBMI} that were either identified by the joint 4df test or by the overall (sex-combined) analysis. Each plot highlights the most significant SNP for the combined effect ($P_{Overall}$) or for the joint test (P_{Joint}) and illustrates p-values for age-differences ($P_{Agediff}$), sex-differences ($P_{Sexdiff}$) and P_{Joint} or $P_{Overall}$ respectively. The figure is sorted according to chromosome and position. The plots are based on GrCh37 build positions and annotations.

(TIF)

S1 Table. Study design, number of individuals and sample quality control for genome-wide association study cohorts.

(XLSX)

S2 Table. Information on genotyping methods, quality control of SNPs, imputation, and statistical analysis for genome-wide association study cohorts.

(XLSX)

S3 Table. Study-specific descriptive statistics of study cohorts. ** There were significant differences in the number of subjects available for different phenotypes. In this case, separate summary statistics were provided.

(XLSX)

S4 Table. Stratum-specific results and extended details for the 15 age-specific BMI loci. The table is ordered according to [Table 1](#).

(XLSX)

S5 Table. Stratum-specific results and extended details for the 44 sex-specific WHR_{adjBMI} loci. The table is ordered according to [Table 2](#).

(XLSX)

S6 Table. Age-specific associations of age-dependent BMI SNPs with blood pressure (BP). Abbreviations: Effect Allele (EA), Other Allele (OA), SNP-by-age interaction effect ($b_{SNP \times AGE}$), SNP-by-age interaction effect standard error ($SE_{SNP \times AGE}$), SNP-by-age interaction P-value ($P_{SNP \times AGE}$).

(XLSX)

S7 Table. Sex-specific associations of sexually dimorphic WHR_{adjBMI} SNPs with lipid traits (GLGC). Abbreviations: Effect Allele (EA), Other Allele (OA), High Density Lipoprotein Cholesterol (HDL), Low Density Lipoprotein (LDL), Total Cholesterol (TC), Triglycerides (TG). (XLSX)

S8 Table. Sex-specific associations of sexually dimorphic WHR_{adjBMI} SNPs with Type 2 Diabetes (T2D, DIAGRAM) and glycemic traits (MAGIC). Abbreviations: Effect Allele (EA), Other Allele (OA), Odds Ratio (OR). (XLSX)

S9 Table. Sex-specific associations of sexually dimorphic WHR_{adjBMI} SNPs with blood pressure (BP) measures. Abbreviations: Effect Allele (EA), Other Allele (OA), Odds Ratio (OR). (XLSX)

S10 Table. Remarkable women-specific associations in the WHR_{adjBMI} lookup data. The table shows SNPs that meet a Bonferroni-corrected significance level ($<0.05/44$) for its sex-specific association with the lookup trait and no association with the lookup trait in the other sex. Only women-specific loci displayed similar patterns in the look-up data. None of the opposite effect direction loci showed opposite effects (requesting $P < 0.05$ in both sexes) with a look-up trait. (XLSX)

S11 Table. Previously-identified associations listed in the NHGRI GWAS Catalog that lie within 500 kb and $r^2 > 0.1$ to our lead BMI SNPs. (XLSX)

S12 Table. Previously-identified associations listed in the NHGRI GWAS Catalog that lie within 500 kb and $r^2 > 0.1$ to our lead WHR_{adjBMI} SNPs. (XLSX)

S13 Table. BMI loci showing significant age-differences in adults $\leq 50y$ compared to adults $>50y$. Analysis was restricted to non-case control studies. (XLSX)

S14 Table. WHR_{adjBMI} loci showing significant sex-differences. Analysis was restricted to non-case control studies. (XLSX)

S15 Table. Effect estimates of BMI loci selected for age-differences and birthweight from 26,836 participants in the EGG consortium. (XLSX)

S16 Table. Odds ratios of BMI loci selected for age-differences and childhood obesity from 5,530 cases and 8,318 controls in the EGG consortium. (XLSX)

S17 Table. Differences in effect estimates between young adults, adults $\leq 50y$, and adults $>50y$ for BMI loci selected for age-differences. (XLSX)

S18 Table. Effect estimates for weight change trajectories in adults between the ages of 20 and 65 years of age in loci showing effect size differences in BMI by age. (XLSX)

S19 Table. Top hits of the human sex-specific eQTL lookup of the sex-specific WHR_{adjBMI} associated SNPs. Presented are SNPs with significant sex-differences in eQTL effects, selected according to $FDR(P\text{-Sexdiff}) < 5\%$, with and without initial filtering on overall expression effects ($FDR(P\text{-Overall}) < 20\%$).

(XLSX)

S20 Table. List of tissues in which DEPICT identified significant expression ($FDR < 0.1$) of genes from age-specific BMI associated loci in at least one of the four approaches.

(XLSX)

S21 Table. Gene sets enriched ($FDR < 0.1$) for harboring SNPs with sex-different effect on WHR_{adjBMI} identified by DEPICT.

(XLSX)

S22 Table. All significant function or disease annotations identified in IPA for the younger adult-specific BMI loci. Functions that remain significant after B-H ($p < 0.05$) correction are marked in bold.

(XLSX)

S23 Table. All significant function or disease annotations identified in IPA for the older adult-specific BMI loci. Functions that remain significant after B-H correction are marked in bold.

(XLSX)

S24 Table. All significant canonical pathways identified in IPA for the women-specific WHR_{adjBMI} loci. Pathways that remain significant after B-H correction are marked in bold.

(XLSX)

S25 Table. All significant function or disease annotations identified in IPA for the women-specific WHR_{adjBMI} loci. Functions that remain significant after B-H correction are marked in bold.

(XLSX)

S26 Table. All significant function or disease annotations identified in IPA for the men-specific WHR_{adjBMI} loci. Functions that remain significant after B-H correction are marked in bold.

(XLSX)

S27 Table. Proportion of genetic to phenotypic variance explained for BMI, and WHR_{adjBMI} estimated using GCTA and Heritability estimated using SOLAR.

(XLSX)

S28 Table. BMI main or joint (main+interaction, 4df) effect findings compared to results from the GIANT BMI group [19].

(XLSX)

S29 Table. WHR_{adjBMI} main or joint (main+interaction, 4df) effect findings compared to results from the GIANT WAIST group [18].

(XLSX)

S1 Text. Supplementary note.

(DOCX)

S2 Text. Consortia members and extended acknowledgments.

(DOCX)

Author Contributions

Conceived and designed the experiments: IBB IMH KEN RJFL TWW ZK. Performed the experiments: AEJ LBa MGr TWW ZK. Analyzed the data: AEJ LBa MGr RM TWW ZK. Contributed reagents/materials/analysis tools: AEJ LBa MGr TWW ZK. Wrote the paper: AEJ IBB IMH KEN LBa MGr MFF RJFL TWW ZK. Contributed genome-wide association study results: AAH AB ACA ACHa ACHe ACS AD AEF AEJ AFW AG AGU AHa AHo AJ ÅJ AJMdc AJO AJS AKH ALJ ALMV AMa AMe APa APe APM APo ARSa ARSh ASB ASG ASt ATe ATH ATö ATr AUJ AVS AWM BAO BDM BF BL BMK BMP BOB BSt BT BWP CAB CAH CBa CBe CBo CG CHa CHe CLang CM CMG CML CMvD CO CR CSF CW CWKC DA DAB DAE DC DCR DFL DH DIB DIC DJH DPS DS EA EB EE EI EJCdG EK EM EPB ETs EV EW EZ FB FC FF FG FH FLM FRe FRiv FRiz FSC FWA GB GC GDed GDel GE GL GM GP GRA GT GWa GWi GWM HAB HAK HC HGra HGrö HH HMdR HMS HScha HSchu HSn HV HVH HWal HWat IB IBB IF IK IMH IML IMN IN IPi IPr IR IT JAS JBe JBl JCB JCC JCM JCo JCT JcZ JD JDR JEH JErd JEri JFw JG JGE JH JHS JHZ JIR JJH JKr JKu JLa JLBG JLu JMJ JMV JNH JPT JROC JSB JSh JSin JSK JSP JSV JT JvS JvVO JWJ JY KAR KBJ KEN KKO KLMoh KLMon KMR KP KSL KSti KSti KTK LAC LALMK LBa LBr LCG LCPGMdG LFe LJL LJP LK LLa LLi LLW LM LMR LMYA LPas LPat LPé LQ LS LV LYe LYu MASa MASw MBa MBl MBo MBr MCV MCZ MD MEK MEM MF MFF MGo MGP MGr MHa MHe MJ MKä MKaa MKar MLa MLL MLo MMa MMe MN MP MRh MS MSN MWald MWalk ND NE NF NGI NGM NH NHK NJW NLHC NLP NMvS NP NS NV NvdV NWR OG OPo OTR PAFM PB PBK PD PEHS PES PF PGH PH PIWdB PjvdM PK PKM PLdJ PMR PPP PS PSC PV PvdH PVG PWF RAS RJFL RL RM RNB RPS RR RTG RWW SA SB SCvD SE SG SH SHV SHW SIB SJ SJC SJK SJLB SKan SKat SKo SL SMä SRB SRW Ssa SSc SSeb ST STT SWvdL TAL TBG TBH TDS TE TEG TFa TH TIAS TK TKR TL TLA TMT TQ TR TSA TT TW UG UT UV VG VLö VLy VSa VSt VV WLMA WM WRS WZ YCC YDIC YJS YLi YLu YMG YPL ZK ZW. Contributed MetaboChip association study results: AB AHa AHi AL AMe ASi AW BG BOB BSe CLang CLanz DB DK EAA EE EI EM ETr FB FRe IB IMH JHZ JLi JLu JSa KB KF KGH KH KKO KL KSti KTK LLB LLH LM LYe MGo MKi MKu MMN MRi MU NGF NGr NJW NLP NN NTK OH OPe PD PF PM PWF RAS RJFL RJS RL RRM RW SA SCB SKan SL SMe TE TFa TH THS TI TJ TS TSA TWW UdF WKo WKr. Contributed SNP look-up data: AC CCW CEJ DCR GBE IPr JSim LAC NLHC RM SSen TFe VLä. Performed Study-specific GCTA analyses: ATe MGr SR TE TWW ZK. Performed study-specific quality control: AEJ JcZ LBa MFF MGr RM SC TE TFa TOK TWW YLu ZK. Performed DEPICT Gene Pathway analyses: JKa JNH LFr THP. Performed IPA Gene Pathway analyses: AEJ. Performed Expression Quantitative trait loci (eQTL) analyses: HJW LFr RJ TE. Performed Mouse microarray expression analyses: DJC MDN. Contributed Look-up information and analyses for young adults ages 16–25 years: KEN LAC MGr PGL SIB. GIANT Steering committee: CMvD CSF DJH DPS DS EI GRA IB IBB IMH JNH JROC KEN KLMoh LCG MBo MIMC PD RCK RJFL SIB TLA UT.

References

1. Vazquez G., et al., Comparison of body mass index, waist circumference, and waist/hip ratio in predicting incident diabetes: a meta-analysis. *Epidemiol Rev*, 2007. 29: p. 115–28. PMID: [17494056](#)
2. Pischon T., et al., General and abdominal adiposity and risk of death in Europe. *N Engl J Med*, 2008. 359(20): p. 2105–20. doi: [10.1056/NEJMoa0801891](#) PMID: [19005195](#)
3. Mokdad A.H., et al., Prevalence of obesity, diabetes, and obesity-related health risk factors, 2001. *JAMA*, 2003. 289(1): p. 76–9. PMID: [12503980](#)
4. Must A., et al., The disease burden associated with overweight and obesity. *JAMA*, 1999. 282(16): p. 1523–9. PMID: [10546691](#)

5. Yusuf S., et al., Obesity and the risk of myocardial infarction in 27,000 participants from 52 countries: a case-control study. *Lancet*, 2005. 366(9497): p. 1640–9. PMID: [16271645](#)
6. Canoy D., et al., Body fat distribution and risk of coronary heart disease in men and women in the European Prospective Investigation Into Cancer and Nutrition in Norfolk cohort: a population-based prospective study. *Circulation*, 2007. 116(25): p. 2933–43. PMID: [18071080](#)
7. De Mello J.J., et al., Gender Differences In The Evaluation Of Adult Body Composition. *Medicine and Science in Sports and Exercise*, 2005. 37: p. S299–S299.
8. Kirchengast S., Gender Differences in Body Composition from Childhood to Old Age: An Evolutionary Point of View. *Journal of Life Sciences*, 2010. 2(1): p. 1–10.
9. Legato M.J., Beyond women's health the new discipline of gender-specific medicine. *Med Clin North Am*, 2003. 87(5): p. 917–37, vii. PMID: [14621324](#)
10. Randall J.C., et al., Sex-stratified Genome-wide Association Studies Including 270,000 Individuals Show Sexual Dimorphism in Genetic Loci for Anthropometric Traits. *Plos Genetics*, 2013. 9(6): p. e1003500. doi: [10.1371/journal.pgen.1003500](#) PMID: [23754948](#)
11. Walter A.A., et al., Sarcopenia Indices: Age- And Gender-related Differences In Body Composition, Strength, And Muscle Quality. *Medicine and Science in Sports and Exercise*, 2012. 44: p. 98–98.
12. Wells J.C., Sexual dimorphism of body composition. *Best Pract Res Clin Endocrinol Metab*, 2007. 21(3): p. 415–30. PMID: [17875489](#)
13. Loomba-Albrecht L.A. and Styne D.M., Effect of puberty on body composition. *Curr Opin Endocrinol Diabetes Obes*, 2009. 16(1): p. 10–5. PMID: [19115520](#)
14. Rogol A.D., Roemmich J.N., and Clark P.A., Growth at puberty. *J Adolesc Health*, 2002. 31(6 Suppl): p. 192–200. PMID: [12470915](#)
15. Rosenbaum M. and Leibel R.L., Clinical review 107: Role of gonadal steroids in the sexual dimorphisms in body composition and circulating concentrations of leptin. *J Clin Endocrinol Metab*, 1999. 84(6): p. 1784–9. PMID: [10372664](#)
16. Kuk J.L., et al., Age-related changes in total and regional fat distribution. *Ageing Res Rev*, 2009. 8(4): p. 339–48. doi: [10.1016/j.arr.2009.06.001](#) PMID: [19576300](#)
17. Mott J.W., et al., Relation between body fat and age in 4 ethnic groups. *American Journal of Clinical Nutrition*, 1999. 69(5): p. 1007–1013. PMID: [10232643](#)
18. Shungin D., et al., New genetic loci link adipose and insulin biology to body fat distribution. *Nature*, 2015. 518(7538): p. 187–96. doi: [10.1038/nature14132](#) PMID: [25673412](#)
19. Locke A.E., et al., Genetic studies of body mass index yield new insights for obesity biology. *Nature*, 2015. 518(7538): p. 197–206. doi: [10.1038/nature14177](#) PMID: [25673413](#)
20. Abdunour J., et al., The effect of the menopausal transition on body composition and cardiometabolic risk factors: a Montreal-Ottawa New Emerging Team group study. *Menopause*, 2012. 19(7): p. 760–7. doi: [10.1097/gme.0b013e318240f6f3](#) PMID: [22395454](#)
21. Douchi T., et al., Precedence of bone loss over changes in body composition and body fat distribution within a few years after menopause. *Maturitas*, 2003. 46(2): p. 133–138. PMID: [14559384](#)
22. Morita Y., et al., Precedence of the shift of body-fat distribution over the change in body composition after menopause. *Journal of Obstetrics and Gynaecology Research*, 2006. 32(5): p. 513–516. PMID: [16984520](#)
23. Bromberger J.T., et al., Prospective study of the determinants of age at menopause. *Am J Epidemiol*, 1997. 145(2): p. 124–33. PMID: [9006309](#)
24. Gold E.B., et al., Factors associated with age at natural menopause in a multiethnic sample of midlife women. *Am J Epidemiol*, 2001. 153(9): p. 865–74. PMID: [11323317](#)
25. Gold E.B., et al., Factors Related to Age at Natural Menopause: Longitudinal Analyses From SWAN. *Am J Epidemiol*, 2013. 178(1): p. 70–83. doi: [10.1093/aje/kws421](#) PMID: [23788671](#)
26. Kooperberg C. and Leblanc M., Increasing the power of identifying gene x gene interactions in genome-wide association studies. *Genet Epidemiol*, 2008. 32(3): p. 255–63. doi: [10.1002/gepi.20300](#) PMID: [18200600](#)
27. Aschard H., et al., Genome-wide meta-analysis of joint tests for genetic and gene-environment interaction effects. *Hum Hered*, 2010. 70(4): p. 292–300. doi: [10.1159/000323318](#) PMID: [21293137](#)
28. Speliotes E.K., et al., Association analyses of 249,796 individuals reveal 18 new loci associated with body mass index. *Nat Genet*, 2010. 42(11): p. 937–48. doi: [10.1038/ng.686](#) PMID: [20935630](#)
29. Heid I.M., et al., Meta-analysis identifies 13 new loci associated with waist-hip ratio and reveals sexual dimorphism in the genetic basis of fat distribution. *Nature Genetics*, 2010. 42(11): p. 949–60. doi: [10.1038/ng.685](#) PMID: [20935629](#)

30. Newton-Cheh C., et al., Genome-wide association study identifies eight loci associated with blood pressure. *Nat Genet*, 2009. 41(6): p. 666–76. doi: [10.1038/ng.361](https://doi.org/10.1038/ng.361) PMID: [19430483](https://pubmed.ncbi.nlm.nih.gov/19430483/)
31. Teslovich T.M., et al., Biological, clinical and population relevance of 95 loci for blood lipids. *Nature*, 2010. 466(7307): p. 707–13. doi: [10.1038/nature09270](https://doi.org/10.1038/nature09270) PMID: [20686565](https://pubmed.ncbi.nlm.nih.gov/20686565/)
32. Morris A.P., et al., Large-scale association analysis provides insights into the genetic architecture and pathophysiology of type 2 diabetes. *Nat Genet*, 2012. 44(9): p. 981–90. doi: [10.1038/ng.2383](https://doi.org/10.1038/ng.2383) PMID: [22885922](https://pubmed.ncbi.nlm.nih.gov/22885922/)
33. Scott R.A., et al., Large-scale association analyses identify new loci influencing glycemic traits and provide insight into the underlying biological pathways. *Nat Genet*, 2012. 44(9): p. 991–1005. doi: [10.1038/ng.2385](https://doi.org/10.1038/ng.2385) PMID: [22885924](https://pubmed.ncbi.nlm.nih.gov/22885924/)
34. Simino J., et al., Gene-Age Interactions in Blood Pressure Regulation: A Large-Scale Investigation with the CHARGE, Global BPgen, and ICBP Consortia. *Am J Hum Genet*, 2014. 95(1): p. 24–38. doi: [10.1016/j.ajhg.2014.05.010](https://doi.org/10.1016/j.ajhg.2014.05.010) PMID: [24954895](https://pubmed.ncbi.nlm.nih.gov/24954895/)
35. Hindroff, L.A., et al., *A Catalog of Published Genome-Wide Association Studies Available at www.genome.gov/gwastudies*. 2010.
36. Horikoshi M., et al., New loci associated with birth weight identify genetic links between intrauterine growth and adult height and metabolism. *Nat Genet*, 2013. 45(1): p. 76–82. doi: [10.1038/ng.2477](https://doi.org/10.1038/ng.2477) PMID: [23202124](https://pubmed.ncbi.nlm.nih.gov/23202124/)
37. Bradfield J.P., et al., A genome-wide association meta-analysis identifies new childhood obesity loci. *Nat Genet*, 2012. 44(5): p. 526–31. doi: [10.1038/ng.2247](https://doi.org/10.1038/ng.2247) PMID: [22484627](https://pubmed.ncbi.nlm.nih.gov/22484627/)
38. Graff M., et al., The influence of obesity-related single nucleotide polymorphisms on BMI across the life course: the PAGE study. *Diabetes*, 2013. 62(5): p. 1763–7. doi: [10.2337/db12-0863](https://doi.org/10.2337/db12-0863) PMID: [23300277](https://pubmed.ncbi.nlm.nih.gov/23300277/)
39. Leitsalu L., et al., Cohort Profile: Estonian Biobank of the Estonian Genome Center, University of Tartu. *Int J Epidemiol*, 2014.
40. Fehrmann R.S., et al., Trans-eQTLs reveal that independent genetic variants associated with a complex phenotype converge on intermediate genes, with a major role for the HLA. *PLoS Genet*, 2011. 7(8): p. e1002197. doi: [10.1371/journal.pgen.1002197](https://doi.org/10.1371/journal.pgen.1002197) PMID: [21829388](https://pubmed.ncbi.nlm.nih.gov/21829388/)
41. Luong A., et al., Molecular characterization of human acetyl-CoA synthetase, an enzyme regulated by sterol regulatory element-binding proteins. *J Biol Chem*, 2000. 275(34): p. 26458–66. PMID: [10843999](https://pubmed.ncbi.nlm.nih.gov/10843999/)
42. Wright F.A., et al., Heritability and genomics of gene expression in peripheral blood. *Nat Genet*, 2014. 46(5): p. 430–7. doi: [10.1038/ng.2951](https://doi.org/10.1038/ng.2951) PMID: [24728292](https://pubmed.ncbi.nlm.nih.gov/24728292/)
43. Jansen R., et al., Sex differences in the human peripheral blood transcriptome. *BMC Genomics*, 2014. 15: p. 33. doi: [10.1186/1471-2164-15-33](https://doi.org/10.1186/1471-2164-15-33) PMID: [24438232](https://pubmed.ncbi.nlm.nih.gov/24438232/)
44. Grove K.L., et al., A microarray analysis of sexual dimorphism of adipose tissues in high-fat-diet-induced obese mice. *Int J Obes (Lond)*, 2010. 34(6): p. 989–1000.
45. Baumgartner B.G., et al., Identification of a novel modulator of thyroid hormone receptor-mediated action. *PLoS One*, 2007. 2(11): p. e1183. PMID: [18030323](https://pubmed.ncbi.nlm.nih.gov/18030323/)
46. Sala D., et al., Autophagy-regulating TP53INP2 mediates muscle wasting and is repressed in diabetes. *J Clin Invest*, 2014. 124(5): p. 1914–27. doi: [10.1172/JCI72327](https://doi.org/10.1172/JCI72327) PMID: [24713655](https://pubmed.ncbi.nlm.nih.gov/24713655/)
47. Pers T.H., et al., Biological interpretation of genome-wide association studies using predicted gene functions. *Nat Commun*, 2015. 6: p. 5890. doi: [10.1038/ncomms6890](https://doi.org/10.1038/ncomms6890) PMID: [25597830](https://pubmed.ncbi.nlm.nih.gov/25597830/)
48. Veilleux A., et al., Glucocorticoid-induced androgen inactivation by aldo-keto reductase 1C2 promotes adipogenesis in human preadipocytes. *Am J Physiol Endocrinol Metab*, 2012. 302(8): p. E941–9. doi: [10.1152/ajpendo.00069.2011](https://doi.org/10.1152/ajpendo.00069.2011) PMID: [22275760](https://pubmed.ncbi.nlm.nih.gov/22275760/)
49. Yoon M., The role of PPARalpha in lipid metabolism and obesity: focusing on the effects of estrogen on PPARalpha actions. *Pharmacol Res*, 2009. 60(3): p. 151–9. doi: [10.1016/j.phrs.2009.02.004](https://doi.org/10.1016/j.phrs.2009.02.004) PMID: [19646654](https://pubmed.ncbi.nlm.nih.gov/19646654/)
50. Yang J., et al., Common SNPs explain a large proportion of the heritability for human height. *Nat Genet*, 2010. 42(7): p. 565–9. doi: [10.1038/ng.608](https://doi.org/10.1038/ng.608) PMID: [20562875](https://pubmed.ncbi.nlm.nih.gov/20562875/)
51. Hardy R., et al., Life course variations in the associations between FTO and MC4R gene variants and body size. *Hum Mol Genet*, 2010. 19(3): p. 545–52. doi: [10.1093/hmg/ddp504](https://doi.org/10.1093/hmg/ddp504) PMID: [19880856](https://pubmed.ncbi.nlm.nih.gov/19880856/)
52. Hertel J.K., et al., FTO, type 2 diabetes, and weight gain throughout adult life: a meta-analysis of 41,504 subjects from the Scandinavian HUNT, MDC, and MPP studies. *Diabetes*, 2011. 60(5): p. 1637–44. doi: [10.2337/db10-1340](https://doi.org/10.2337/db10-1340) PMID: [21398525](https://pubmed.ncbi.nlm.nih.gov/21398525/)
53. den Hoed M., et al., Genetic susceptibility to obesity and related traits in childhood and adolescence: influence of loci identified by genome-wide association studies. *Diabetes*, 2010. 59(11): p. 2980–8. doi: [10.2337/db10-0370](https://doi.org/10.2337/db10-0370) PMID: [20724581](https://pubmed.ncbi.nlm.nih.gov/20724581/)

54. Graff M., et al., Estimation of genetic effects on BMI during adolescence in an ethnically diverse cohort: The National Longitudinal Study of Adolescent Health. *Nutr Diabetes*, 2012. 2: p. e47. doi: [10.1038/nutd.2012.20](https://doi.org/10.1038/nutd.2012.20) PMID: [23168566](https://pubmed.ncbi.nlm.nih.gov/23168566/)
55. Murphy R.A., et al., Candidate Gene Association Study of BMI-Related Loci, Weight, and Adiposity in Old Age. *J Gerontol A Biol Sci Med Sci*, 2013. 68(6): p. 661–6. doi: [10.1093/gerona/gls227](https://doi.org/10.1093/gerona/gls227) PMID: [23160366](https://pubmed.ncbi.nlm.nih.gov/23160366/)
56. Elks C.E., et al., Adult obesity susceptibility variants are associated with greater childhood weight gain and a faster tempo of growth: the 1946 British Birth Cohort Study. *Am J Clin Nutr*, 2012. 95(5): p. 1150–6. doi: [10.3945/ajcn.111.027870](https://doi.org/10.3945/ajcn.111.027870) PMID: [22456663](https://pubmed.ncbi.nlm.nih.gov/22456663/)
57. Sovio U., et al., Association between common variation at the FTO locus and changes in body mass index from infancy to late childhood: the complex nature of genetic association through growth and development. *PLoS Genet*, 2011. 7(2): p. e1001307. doi: [10.1371/journal.pgen.1001307](https://doi.org/10.1371/journal.pgen.1001307) PMID: [21379325](https://pubmed.ncbi.nlm.nih.gov/21379325/)
58. Graff M., et al., Genome-wide analysis of BMI in adolescents and young adults reveals additional insight into the effects of genetic loci over the life course. *Hum Mol Genet*, 2013. 22(17): p. 3597–607. doi: [10.1093/hmg/ddt205](https://doi.org/10.1093/hmg/ddt205) PMID: [23669352](https://pubmed.ncbi.nlm.nih.gov/23669352/)
59. Do R., et al., Common variants associated with plasma triglycerides and risk for coronary artery disease. *Nat Genet*, 2013. 45(11): p. 1345–52. doi: [10.1038/ng.2795](https://doi.org/10.1038/ng.2795) PMID: [24097064](https://pubmed.ncbi.nlm.nih.gov/24097064/)
60. Fromm-Dornieden C., et al., Extrinsic and intrinsic regulation of DOR/TP53INP2 expression in mice: effects of dietary fat content, tissue type and sex in adipose and muscle tissues. *Nutr Metab (Lond)*, 2012. 9(1): p. 86.
61. McCarthy M.I., Genomics, type 2 diabetes, and obesity. *N Engl J Med*, 2010. 363(24): p. 2339–50. doi: [10.1056/NEJMra0906948](https://doi.org/10.1056/NEJMra0906948) PMID: [21142536](https://pubmed.ncbi.nlm.nih.gov/21142536/)
62. Travers M.E. and McCarthy M.I., Type 2 diabetes and obesity: genomics and the clinic. *Hum Genet*, 2011. 130(1): p. 41–58. doi: [10.1007/s00439-011-1023-8](https://doi.org/10.1007/s00439-011-1023-8) PMID: [21647602](https://pubmed.ncbi.nlm.nih.gov/21647602/)
63. Li Y., et al., Genotype Imputation. *Annual Review of Genomics and Human Genetics*, 2009. 10(1): p. 387–406.
64. Marchini J., et al., A new multipoint method for genome-wide association studies by imputation of genotypes. *Nat Genet*, 2007. 39(7): p. 906–913. PMID: [17572673](https://pubmed.ncbi.nlm.nih.gov/17572673/)
65. Guan Y. and Stephens M., Practical Issues in Imputation-Based Association Mapping. *PLoS Genet*, 2008. 4(12): p. e1000279. doi: [10.1371/journal.pgen.1000279](https://doi.org/10.1371/journal.pgen.1000279) PMID: [19057666](https://pubmed.ncbi.nlm.nih.gov/19057666/)
66. Voight B.F., et al., The metabochip, a custom genotyping array for genetic studies of metabolic, cardiovascular, and anthropometric traits. *PLoS Genet*, 2012. 8(8): p. e1002793. doi: [10.1371/journal.pgen.1002793](https://doi.org/10.1371/journal.pgen.1002793) PMID: [22876189](https://pubmed.ncbi.nlm.nih.gov/22876189/)
67. Li Y., et al., MaCH: using sequence and genotype data to estimate haplotypes and unobserved genotypes. *Genetic Epidemiology*, 2010. 34(8): p. 816–34. doi: [10.1002/gepi.20533](https://doi.org/10.1002/gepi.20533) PMID: [21058334](https://pubmed.ncbi.nlm.nih.gov/21058334/)
68. Aulchenko Y.S., Struchalin M.V., and van Duijn C.M., ProbABEL package for genome-wide association analysis of imputed data. *BMC Bioinformatics*, 2010. 11: p. 134. doi: [10.1186/1471-2105-11-134](https://doi.org/10.1186/1471-2105-11-134) PMID: [20233392](https://pubmed.ncbi.nlm.nih.gov/20233392/)
69. Aulchenko Y.S., et al., GenABEL: an R library for genome-wide association analysis. *Bioinformatics*, 2007. 23(10): p. 1294–6. PMID: [17384015](https://pubmed.ncbi.nlm.nih.gov/17384015/)
70. Abecasis G.R. and Wigginton J.E., Handling marker-marker linkage disequilibrium: pedigree analysis with clustered markers. *Am J Hum Genet*, 2005. 77(5): p. 754–67. PMID: [16252236](https://pubmed.ncbi.nlm.nih.gov/16252236/)
71. Purcell S., et al., PLINK: a tool set for whole-genome association and population-based linkage analyses. *The American Journal of Human Genetics*, 2007. 81: p. 559–575. PMID: [17701901](https://pubmed.ncbi.nlm.nih.gov/17701901/)
72. Kutalik Z., et al., Methods for testing association between uncertain genotypes and quantitative traits. *Biostatistics*, 2011. 12(1): p. 1–17. doi: [10.1093/biostatistics/kxq039](https://doi.org/10.1093/biostatistics/kxq039) PMID: [20543033](https://pubmed.ncbi.nlm.nih.gov/20543033/)
73. Winkler T.W., et al., Quality control and conduct of genome-wide association meta-analyses. *Nat Protoc*, 2014. 9(5): p. 1192–212. doi: [10.1038/nprot.2014.071](https://doi.org/10.1038/nprot.2014.071) PMID: [24762786](https://pubmed.ncbi.nlm.nih.gov/24762786/)
74. Devlin B. and Roeder K., Genomic control for association studies. *Biometrics*, 1999. 55(4): p. 997–1004. PMID: [11315092](https://pubmed.ncbi.nlm.nih.gov/11315092/)
75. Willer C.J., Li Y., and Abecasis G.R., METAL: fast and efficient meta-analysis of genomewide association scans. *Bioinformatics*, 2010. 26(17): p. 2190–1. doi: [10.1093/bioinformatics/btq340](https://doi.org/10.1093/bioinformatics/btq340) PMID: [20616382](https://pubmed.ncbi.nlm.nih.gov/20616382/)
76. Benjamini Y. and Hochberg Y., Controlling the False Discovery Rate—a Practical and Powerful Approach to Multiple Testing. *Journal of the Royal Statistical Society Series B-Methodological*, 1995. 57(1): p. 289–300.

77. Rowland M.L., Self-reported weight and height. *Am J Clin Nutr*, 1990. 52(6): p. 1125–33. PMID: [2239790](#)
78. Elgar F.J., et al., Validity of self-reported height and weight and predictors of bias in adolescents. *J Adolesc Health*, 2005. 37(5): p. 371–5. PMID: [16227121](#)
79. Keith S.W., et al., Use of self-reported height and weight biases the body mass index-mortality association. *Int J Obes (Lond)*, 2011. 35(3): p. 401–8.
80. Purcell S., et al., PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet*, 2007. 81(3): p. 559–75. PMID: [17701901](#)
81. Genomes Project, C., et al., A map of human genome variation from population-scale sequencing. *Nature*, 2010. 467(7319): p. 1061–73. doi: [10.1038/nature09534](#) PMID: [20981092](#)
82. Hindorf, L.A., et al., *A Catalog of Published Genome-Wide Association Studies*. 2010.
83. Lango Allen, H., et al., Hundreds of variants clustered in genomic loci and biological pathways affect human height. *Nature*, 2010. 467(7317): p. 832–8. doi: [10.1038/nature09410](#) PMID: [20881960](#)
84. Kamatani Y., et al., Genome-wide association study of hematological and biochemical traits in a Japanese population. *Nature Genetics*, 2010. 42(3): p. 210–5. doi: [10.1038/ng.531](#) PMID: [20139978](#)
85. Franke A., et al., Genome-wide meta-analysis increases to 71 the number of confirmed Crohn's disease susceptibility loci. *Nature Genetics*, 2010. 42(12): p. 1118–25. doi: [10.1038/ng.717](#) PMID: [21102463](#)
86. Sawcer S., et al., Genetic risk and a primary role for cell-mediated immune mechanisms in multiple sclerosis. *Nature*, 2011. 476(7359): p. 214–9. doi: [10.1038/nature10251](#) PMID: [21833088](#)
87. Wang K.S., Liu X.F., and Aragam N., A genome-wide meta-analysis identifies novel loci associated with schizophrenia and bipolar disorder. *Schizophr Res*, 2010. 124(1–3): p. 192–9. doi: [10.1016/j.schres.2010.09.002](#) PMID: [20889312](#)
88. Cirulli E.T., et al., Common genetic variation and performance on standardized cognitive tests. *Eur J Hum Genet*, 2010. 18(7): p. 815–20. doi: [10.1038/ejhg.2010.2](#) PMID: [20125193](#)
89. Estrada K., et al., Genome-wide meta-analysis identifies 56 bone mineral density loci and reveals 14 loci associated with risk of fracture. *Nature Genetics*, 2012. 44(5): p. 491–501. doi: [10.1038/ng.2249](#) PMID: [22504420](#)
90. Gieger C., et al., New gene functions in megakaryopoiesis and platelet formation. *Nature*, 2011. 480(7376): p. 201–8. doi: [10.1038/nature10659](#) PMID: [22139419](#)
91. Need A.C., et al., A genome-wide study of common SNPs and CNVs in cognitive performance in the CANTAB. *Hum Mol Genet*, 2009. 18(23): p. 4650–61. doi: [10.1093/hmg/ddp413](#) PMID: [19734545](#)
92. Calvano S.E., et al., A network-based analysis of systemic inflammation in humans. *Nature*, 2005. 437(7061): p. 1032–7. PMID: [16136080](#)
93. Yang J., et al., GCTA: a tool for genome-wide complex trait analysis. *Am J Hum Genet*, 2011. 88(1): p. 76–82. doi: [10.1016/j.ajhg.2010.11.011](#) PMID: [21167468](#)
94. Kutalik Z., et al., Novel method to estimate the phenotypic variation explained by genome-wide association studies reveals large fraction of the missing heritability. *Genet Epidemiol*, 2011. 35(5): p. 341–9. doi: [10.1002/gepi.20582](#) PMID: [21465548](#)
95. Heid I.M., et al., Meta-analysis identifies 13 new loci associated with waist-hip ratio and reveals sexual dimorphism in the genetic basis of fat distribution. *Nat Genet*, 2010. 42(11): p. 949–960. doi: [10.1038/ng.685](#) PMID: [20935629](#)
96. Johnson A.D., et al., SNAP: a web-based tool for identification and annotation of proxy SNPs using HapMap. *Bioinformatics*, 2008. 24(24): p. 2938–9. doi: [10.1093/bioinformatics/btn564](#) PMID: [18974171](#)
97. Kumar P., Henikoff S., and Ng P.C., Predicting the effects of coding non-synonymous variants on protein function using the SIFT algorithm. *Nat Protoc*, 2009. 4(7): p. 1073–81. doi: [10.1038/nprot.2009.86](#) PMID: [19561590](#)
98. Wang K., Li M., and Hakonarson H., ANNOVAR: functional annotation of genetic variants from high-throughput sequencing data. *Nucleic Acids Res*, 2010. 38(16): p. e164. doi: [10.1093/nar/gkq603](#) PMID: [20601685](#)
99. Boyle A.P., et al., Annotation of functional variation in personal genomes using RegulomeDB. *Genome Res*, 2012. 22(9): p. 1790–7. doi: [10.1101/gr.137323.112](#) PMID: [22955989](#)