

Ghrelin



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ABSTRACT

Background: The gastrointestinal peptide hormone ghrelin was discovered in 1999 as the endogenous ligand of the growth hormone secretagogue receptor. Increasing evidence supports more complicated and nuanced roles for the hormone, which go beyond the regulation of systemic energy metabolism.

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Scope of review: In this review, we discuss the diverse biological functions of ghrelin, the regulation of its secretion, and address questions that still remain 15 years after its discovery.

Major conclusions: In recent years, ghrelin has been found to have a plethora of central and peripheral actions in distinct areas including learning and memory, gut motility and gastric acid secretion, sleep/wake rhythm, reward seeking behavior, taste sensation and glucose metabolism.

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Keywords Ghrelin; Growth hormone secretagogue receptor

1. INTRODUCTION

In 1999, Masayasu Kojima, Kenji Kangawa, and colleagues discovered the gastrointestinal peptide hormone ghrelin as the endogenous ligand for the growth hormone secretagogue receptor (GHSR)1a, capable of stimulating growth hormone (GH) release from the anterior pituitary gland [1]. In 2000, Mark Heiman and Matthias Tschöp discovered that ghrelin acts in the brain to regulate food intake, body weight, adiposity, and glucose metabolism [2]. Ghrelin was found to modulate systemic metabolism via activation of orexigenic neural circuits [3,4]. Subsequently, numerous central and peripheral actions of ghrelin were described, including stimulation of gut motility and gastric acid secretion [5,6], modulation of sleep [7–9], taste sensation and reward seeking behavior [10–16], regulation of glucose metabolism [17–20], suppression of brown fat thermogenesis [21–25], modulation of stress and anxiety [26–28], protection against muscle atrophy [29,30], and improvement of cardiovascular functions such as vasodilatation and cardiac contractility [31–34] (Figure 1).

In the early stages of ghrelin research, a model emerged suggesting that ghrelin acts as a “meal initiation” or “hunger” hormone, signaling

gastrointestinal (GI) fuel status to the central nervous system (CNS) in order to adjust food intake and energy expenditure [3,35–38]. Consistent with this role, ghrelin is produced in the oxyntic glands of the gastric fundus [1], its blood levels rise with increased hunger sensations [36,39], and its receptor is located in the hypothalamic neurons that regulate food intake and satiety [40–42]. Recently, however, this traditional and narrowly defined view of ghrelin as a “hunger hormone” has been challenged. Increasing evidence supports a more complex role for ghrelin in the regulation of hunger and metabolism. The aim of this review is to examine the variety of biological functions of ghrelin in order to emphasize its multifaceted nature and to answer some questions that persist after 15 years of ghrelin research.

2. DISCOVERY OF GHRELIN AS THE ENDOGENOUS LIGAND OF THE GROWTH HORMONE SECRETAGOGUE RECEPTOR 1A (GHSR1A)

In the late 1970s, the work of Cyril Bowers and Frank Momany led to the generation of a group of synthetic opioid peptide derivatives that promoted the release of GH from the anterior pituitary [43,44]. The

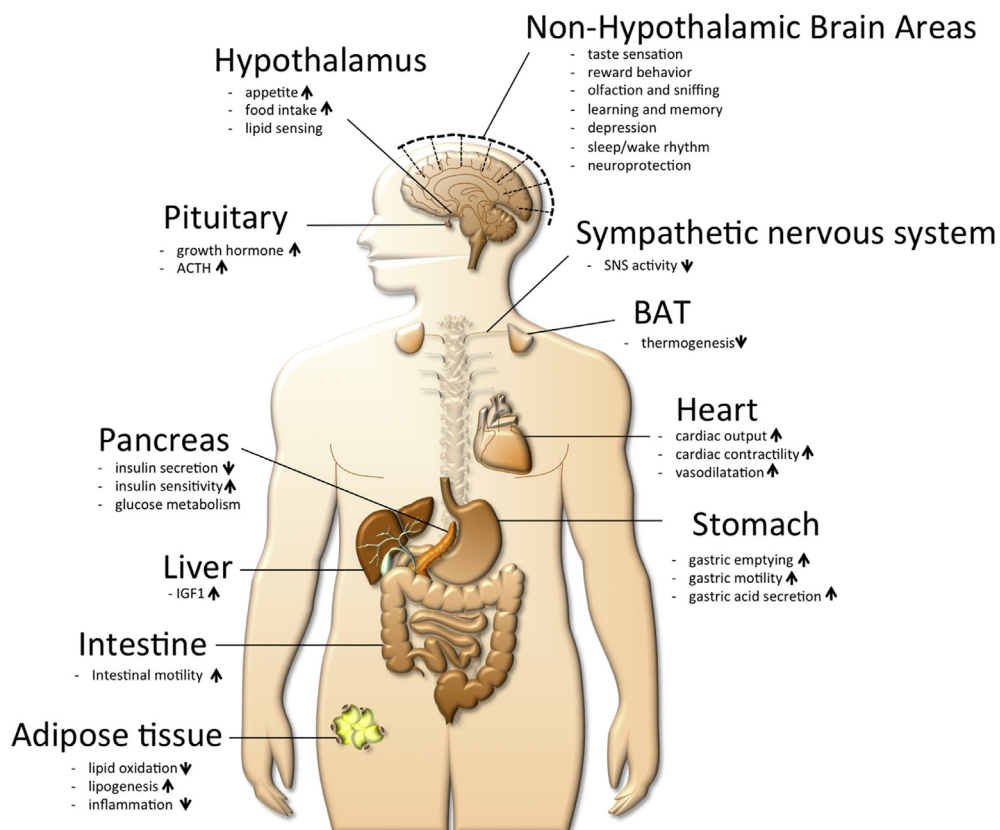


Figure 1: Schematic on ghrelin's physiological effects.

molecules, which Bowers and Momany referred to as GH releasing peptides (GHRPs), were generated by the chemical modification of met-enkephalin and included growth hormone releasing peptide (GHRP)-6, GHRP-2, and hexarelin [45]. Initially, it was thought that these GHRPs acted only on the pituitary, but soon it became clear that they also acted on the hypothalamic arcuate nucleus (ARC) [46], specifically on GH-releasing hormone (GHRH) neurons [41]. The mechanism by which these molecules promoted the release of GH was unknown, but it was distinct from that of the GHRH/somatostatin pathway [46–49]. In 1996, the GHS clinical candidate MK0677 was employed by Roy Smith and Lex van der Ploeg to clone the GH secretagogue receptor (GHSR1a) [50], at which GHSs and GHRPs were shown to be agonists.

In humans, the *GHSR1* gene codes for the full-length G-protein coupled seven transmembrane protein GHSR1a, but a truncated isoform (GHSR1b), which has a wide tissue distribution, is also transcribed [51]. GHSR1a has been shown to homodimerize, but the possibility has been raised that GHSR1a and GHSR1b also heterodimerize [52,53] and that the heterodimer inhibits the activation of GHSR1a [53].

GHSR1a is expressed predominantly in the anterior pituitary gland, pancreatic islets, adrenal gland, thyroid, myocardium, ARC, hippocampus,

the substantia nigra pars compacta (SNpc), ventral tegmental area (VTA), and raphe nuclei [40,51]. In the ARC, in addition to being expressed in GHRH neurons, *GHSR1a* is colocalized in neurons that express neuro-peptide Y (*Npy*) and Agouti related peptide (*AgRP*), which regulate food intake and satiety [42]. Along with the observation that GHRP-6 induced activation of GHSR1 and increased c-Fos expression in NPY neurons [41], these data suggested the presence of an unknown but endogenous ligand for GHSR1, one that might regulate systemic metabolism.

In the years that followed, extensive research efforts were aimed at identifying the endogenous ligand for GHSR1. The ligand remained elusive until 1999 when Kojima and colleagues identified the cognate agonist for GHSR1. Purified from rat stomach extracts, the 28 amino acid peptide was named ‘ghrelin’, a name originating from ‘ghre’, the Proto-Indo-European root of the word ‘grow’ [1].

3. REGULATION OF GHRELIN ACYLATION

3.1. Acylation of ghrelin by the ghrelin O-acyl-transferase (GOAT)

Ghrelin is encoded by the preproghrelin gene (Figure 2), which, in addition to ghrelin, also encodes for a small signal peptide and the 23 amino acid peptide obestatin. Originally, it was thought that obestatin

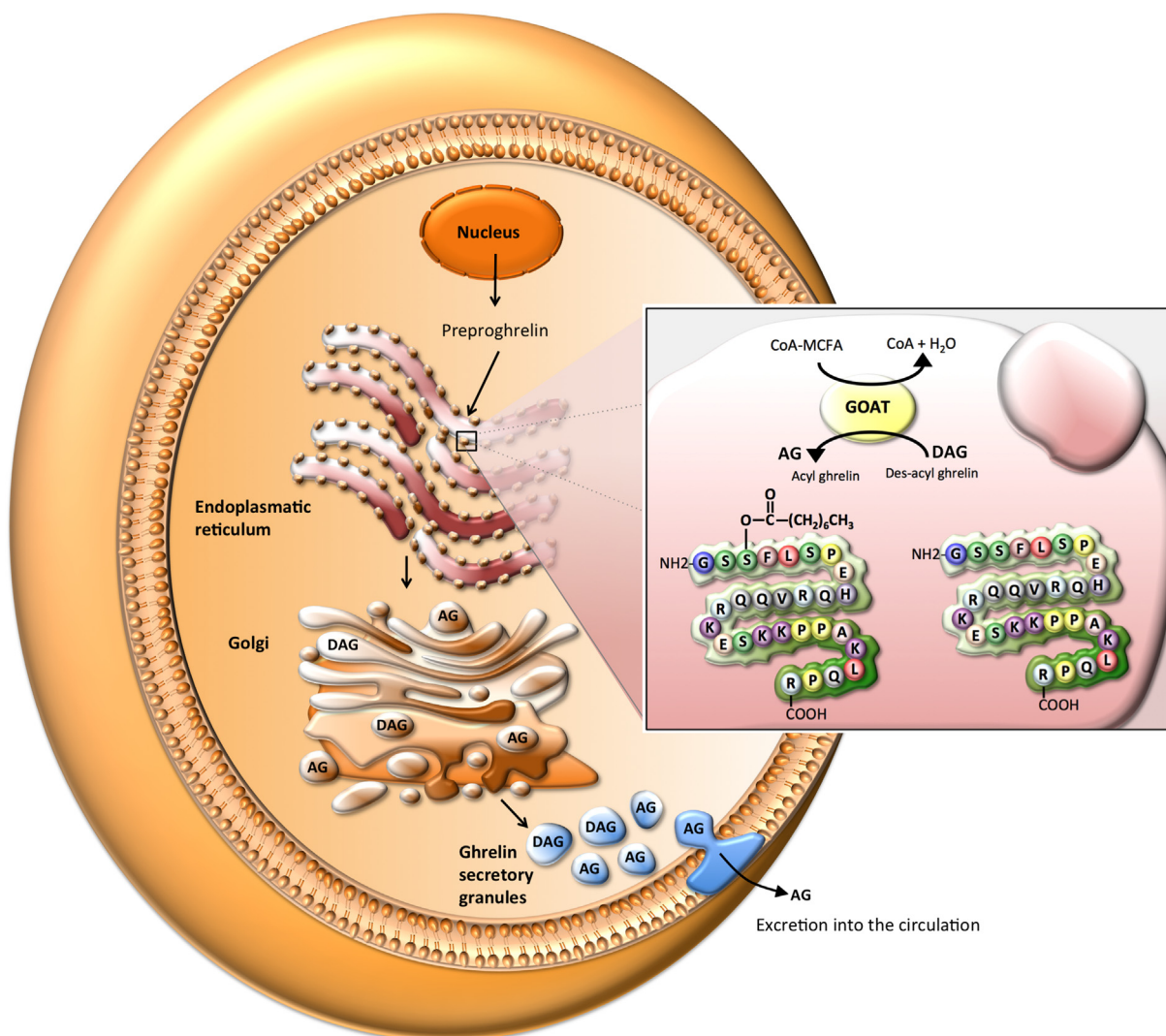


Figure 2: Schematic on the post-translational processing and acylation of ghrelin.

was the endogenous ligand for GPR39 and could inhibit food intake and gastric motility, functions that could counteract the effect of ghrelin [54]. However, several independent groups could not confirm these findings and identified Zn^{2+} as a physiological agonist of GPR39 [55]. To activate its only known receptor, ghrelin requires the attachment of a fatty acid side-chain (preferably C8 or C10) to its serine 3 residue, a rare post-translational modification (acylation) that is achieved by the ghrelin *O*-acyl-transferase (GOAT), a member of the membrane-bound *O*-acyltransferase (MBOAT) family [56,57] (Figure 2).

The discovery of GOAT as the enzyme responsible for ghrelin acylation [56,57] has been a major breakthrough for understanding the role that acyl-modification plays in ghrelin's physiology (Figure 3). This modification, mainly octanoylation and, to a lesser extent, decanoylation, is required for ghrelin's effects on systemic metabolism. The data demonstrating GOAT's essential role in the activation of ghrelin are clear. First, GOAT and des-acyl ghrelin are sufficient to recapitulate the production of acyl-modified ghrelin in cells that normally do not express either of these gene products [56,57]. Second, ghrelin and GOAT share a similar tissue expression profiles in both humans and mice with highest GOAT expression in pancreas and stomach in humans and the stomach and intestine in mice [56,58,59]. Third, GOAT, like ghrelin, is highly conserved across vertebrates. Humans, rats, mice, and zebrafish all exhibit functional GOAT activity, and sequences with amino acid similarities to GOAT are present in other vertebrates, consistent with the presence of octanoylated forms of ghrelin across vertebrates [56]. Finally, the most convincing data for GOAT as ghrelin's acyl transferase are from *GOAT*-deficient mice, which completely lack octanoyl and decanoyl modified forms of ghrelin [20,56,60–62].

3.2. Substrates for GOAT-mediated ghrelin acylation

Intriguingly, the lipids used for ghrelin activation are, at least in part, directly recruited from the pool of ingested dietary lipids [61,63] in a process that may take advantage of the fact that ghrelin-producing X/

A-like cells are located within gastric oxyntic glands. A significant number of these cells are apposed to the stomach lumen, allowing for direct access to a supply of dietary lipids [64]. Furthermore, the preferred fatty acid substrates for GOAT are derived from medium-chain-triglycerides, which can be directly absorbed into the circulation without being broken down by lipases and bile acids [65]. Despite this evidence, the relative contribution of *de novo* synthesized fatty acids in comparison to those directly derived from the diet as substrate of GOAT for ghrelin acylation remains unknown. Mutation studies in the region of the acylated serine 3 have revealed that glycine 1, serine 3, and phenylalanine 4 are critical components of the recognition sequence for GOAT, whereas serine 2, leucine 5, serine 6 and proline 7 seem to be less important [62].

Biochemically, GOAT appears to have two critical substrates, des-acyl ghrelin and short-to mid-chain fatty acids thioesterified with Coenzyme A. Cells expressing both ghrelin and GOAT synthesize serine 3 acyl-ghrelin, with the acyl moiety precursors derived from fatty acids ranging from acetate (C2) to tetradecanoic acid (C14) [56]. The length of the fatty acid used for ghrelin acylation seems to be of importance for ghrelin's metabolic effects, as alterations in the fatty acid length result in differential activation of GHSR1a *in vitro* and alter ghrelin's effect on food intake and adiposity *in vivo* [66]. Thus, modulation of the acyl side-chain may also represent an interesting therapeutic control point for future interventions.

Octanoyl- and decanoyl-modified ghrelin forms are the optimal ligands for activation of the GHSR1a [57,62]. *In vitro* studies recreating the acyl-modification of ghrelin with des-acyl ghrelin peptides, fatty acid CoA esters, and GOAT containing microsomes define the substrate specificity for GOAT. These studies support the idea that GOAT requires fatty acid substrates as high energy fatty acid CoA thioesters and that the amino acid sequence GXSF, where G, X, S, and F correspond to unblocked amino terminal glycine (G), any amino acid (X), serine (S) and phenylalanine (F), respectively, is sufficient as a substrate for

Milestones in ghrelin research until December 2014

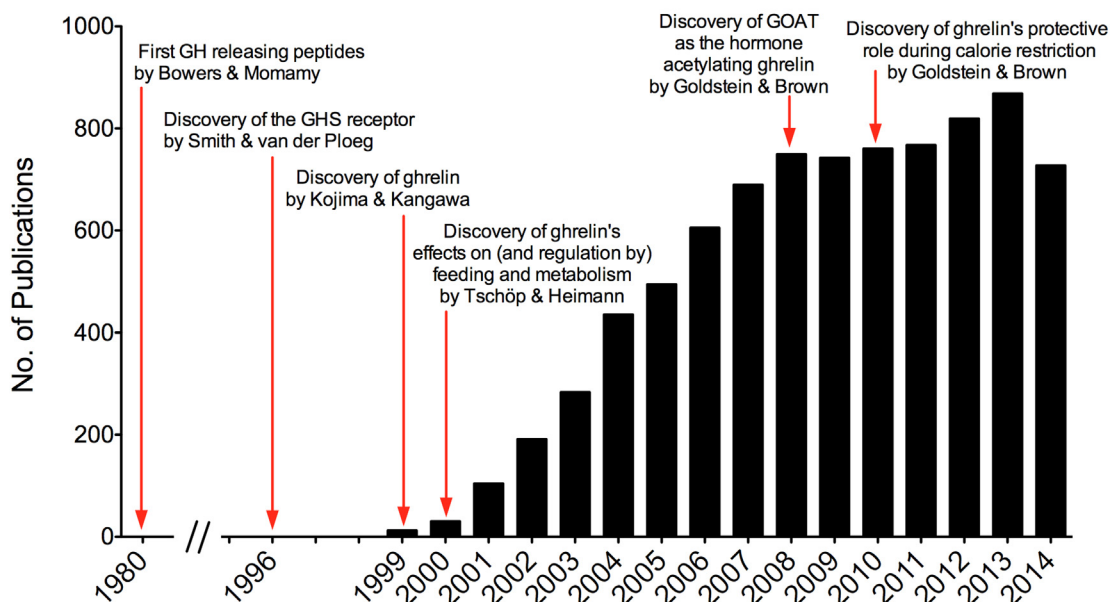


Figure 3: Milestones in ghrelin research. Bar graph represents the number of publications listed in the US National Library of Medicine National Institute of Health (PubMed) and that contain the word 'ghrelin' in either the title or the abstract until December 2014.

GOAT acylation [67]. The structural constraints defined by this amino acid motif appear specific only for ghrelin and suggest that ghrelin may be the principal peptide substrate for GOAT. Most recent studies comparing the *in vitro* selectivity of hexanoyl- and octanoyl-CoA substrates suggest that GOAT may actually prefer hexanoyl CoA over octanoyl CoA substrates, highlighting the importance of the specific fatty acid metabolism in acyl ghrelin producing cells, responsible for producing circulating levels of octanoyl and decanoyl ghrelin [67].

3.3. Evidence suggesting a role of the GOAT-ghrelin system as a nutrient sensor

Most recent studies with genetically modified mice, which are either lacking GOAT or overexpressing both ghrelin and GOAT, establish that the GOAT-ghrelin system acts as a nutrient sensor informing the body of the presence of nutrients, rather than the absence, as commonly proposed [61]. Several observations support this statement. First, prolonged fasting of mice led to well-established, increased levels of total ghrelin, which were caused by increased des-acyl ghrelin rather than acyl ghrelin. This increase in des-acyl ghrelin occurred as GOAT transcript levels decreased in response to the prolonged fasting treatments [61]. Consistent with these observations, GOAT-null mice showed significantly increased total ghrelin levels, being driven only by des-acyl ghrelin as these mice are unable to produce acyl modified ghrelin [56,61]. Second, several studies showed that dietary medium-chain fatty acids (MCFAs) can be a direct source of substrates for ghrelin acylation in rodents and that sensing of MCFAs involves the gustatory G-protein, α -gustducin [61,63,68]. Third, studies also show that mice lacking *GOAT* have lower body weight and fat mass on a MCFA-containing diet compared to wt mice, whereas transgenic mice overexpressing ghrelin and *GOAT* show higher body weight and fat mass and decreased energy expenditure than wt littermates, demonstrating a role for the endogenous acyl-ghrelin in the control of energy balance and adiposity. In addition, data show that a sufficient dietary supply of medium chain triglycerides is crucial for ghrelin acylation, since ghrelin and *GOAT* overexpressing mice are unable to produce large amounts of octanoylated ghrelin when fed a low fat carbohydrate-rich chow diet. Interestingly, transgenic mice fed a regular chow diet show substantial amounts of inactive C2-acetyl-modified ghrelin in the absence of octanoylated ghrelin, suggesting that, at least under these experimental conditions, the GOAT fatty acid substrate for acylation, acetyl-CoA, is sufficiently available for ghrelin acylation. Dietary supplementation of octanoyl triglycerides increases octanoyl-modified ghrelin in these ghrelin and GOAT transgenic animals [61].

Based on these data, it is likely that the GOAT-ghrelin system acts as a nutrient sensor by using readily absorbable MCFAs to signal to the brain that high caloric food is available, leading to optimization of nutrient partitioning and growth signals [61,63]. These recent observations, while informative on the regulation of ghrelin's function by GOAT, highlight key questions that need to be resolved. First, are the observations on the role of GOAT and ghrelin in nutrient sensing and the endocrine control of energy expenditure translatable to humans? Second, what is the physiological role and what are key players for this proposed acyl-ghrelin feedback mechanism observed in the GOAT-deficient animals? Finally, what is (are) the specific biochemical pathway(s) in ghrelin and GOAT expressing cells that produce the necessary levels of C8-CoAs critical for the synthesis of physiologically relevant octanoylated ghrelin? Understanding these fundamental aspects for ghrelin and GOAT will provide critical new insights on the physiological function of this pathway on human physiology.

4. BIOLOGICAL FUNCTIONS OF GHRELIN

4.1. Clinical pharmacology studies on ghrelin's effect on energy metabolism

Numerous human studies have evaluated the effect of ghrelin and its analogs on GH secretion [69–72], food intake [11,34,73–79], body weight [34,78,80] energy expenditure [81–83], glucose homeostasis [84–89], and gastrointestinal motility [90–93]. Peripheral ghrelin or GHRP-2 administration reliably induces the sensation of hunger and increases food intake in lean, obese, healthy and malnourished individuals [94]. Interestingly, iv administration of ghrelin in healthy volunteers increases neural activity in specific brain regions in response to pictures of food. Endogenous fasting ghrelin is positively related to hunger-modulated activity in the hypothalamus, amygdala, and prefrontal cortex in response to palatable food stimuli [95,96]. Activation of these reward centers by ghrelin suggests enhancement of food consumption is a more complex mechanism than the physical sensations of hunger or satiety [97,98]. Interestingly, these ghrelin-brain activity relationships are absent in women with anorexia nervosa, suggesting the possibility of CNS-regulated ghrelin resistance in these individuals [95].

4.2. Clinical pharmacology studies on ghrelin's effect on the GH-Axis

In humans, a single intravenous (iv) injection or a continuous 24 h ghrelin infusion induces acute GH release [99,100] and increases 24 h pulsatile GH secretion [101]. The importance of ghrelin in GH regulation is supported by the observation that abnormalities of GHSR function may be associated with familial short stature [102]. The GHSR locus is one of the top sites suggested to contribute to the genetic variation of height [103]. Several studies have assessed the association of ghrelin and GHSR single nucleotide polymorphisms (SNPs) with height under conditions of obesity and diabetes [reviewed in [104]]. At high doses, ghrelin also increases levels of adrenocorticotrophic hormone (ACTH), prolactin, and cortisol levels [70], while it inhibits levels of luteinizing hormone (LH) [105]. These effects desensitize and normalize with prolonged ghrelin (or ghrelin mimetic) treatment [45,106]. Furthermore, ghrelin mimetics have been investigated as diagnostic agents to establish growth hormone deficiency [107] as well as a therapeutic option for age-dependent GH decline and have yielded some potentially beneficial effects [80] (Table 1). Notably, whereas several clinical

Table 1 – Summary of ghrelin mimetics tested in clinical trials.

Compound	Company	Active/inactive	Indication
Ghrelin mimetic			
Pralmorelin	Kaken Pharma	Approved	Diagnostic for GH deficiency
	Sella Pharma	Approved	
Macimorelin	Aeterna Zentaris	Phase III	Diagnostic for GH deficiency
Anamorelin	Helsinn	Phase III	
Relamorelin	Rhythm	Phase IIb	Diabetic gastroparesis
Uimorelin	Tranzyme	Inactive	Opioid induced constipation/GI functions
Ipamorelin	Helsinn	Inactive	GI functional disorders
Carpromorelin	Pfizer	Inactive	Frailty in elderly
CP 464709	Pfizer	Inactive	Frailty in elderly
Tabimorelin	Novo Nordisk	Inactive	GH deficiency
Ibutamoren	Merck	Inactive	Frailty in elderly
Examorelin/Hexarelin	Diverse Academic sponsored studies	Inactive	GH release
SM 130686	Sumitomo	Inactive	Growth hormone deficiency
LY 426410	Eli Lilly	Inactive	GH release
LY 444711			

studies support a role of ghrelin in regulation growth and height, mice lacking GHSR, ghrelin or GOAT show no growth abnormalities. Whether these discrepant results from mice and humans speaks for distinct GH-release pathways as of today elusive.

4.3. Clinical pharmacology studies on ghrelin's effect on glucose metabolism

Both ghrelin and its receptor are widely expressed in multiple regions of the brain [3,108,109] and in peripheral tissues, such as the intestine [110], pituitary [111,112], kidney [108,113], lung [108,114], heart [110,115,116], ovaries [108], and pancreatic islets [17,40]. Expression in pancreatic islets is consistent with a series of human studies showing increased plasma levels of glucose and decreased plasma levels of insulin following ghrelin administration [84–86,88,89]. GHSRs are also expressed by α -cells of the pancreatic islet and likely contribute to the ability of ghrelin to directly stimulate glucagon secretion [117].

Ghrelin inhibits insulin secretion in most animal studies [118–121], and blockade of pancreatic-derived ghrelin enhances insulin secretion and ameliorates the development of diet-induced glucose intolerance [122]. Supporting these data is the finding that plasma ghrelin and insulin levels seem to be negatively correlated, as the two hormones exhibit reciprocal changes during the day and during a hyperinsulinemic, euglycemic clamp [37,123]. Furthermore, continuous ghrelin infusion for 65 min suppresses glucose-stimulated insulin secretion and impairs glucose tolerance in healthy individuals [124]. In line with this, pharmacological inhibition of GOAT improves glucose disposal by stimulating the release of insulin [60]. The reciprocal relationship of ghrelin and insulin is supported by epidemiologic studies showing an inverse relationship between circulating ghrelin levels and indexes of insulin resistance [39]. A single iv dose of ghrelin significantly increases plasma glucose levels followed by a reduction in fasting insulin levels in lean [84] and obese subjects with or without polycystic ovarian syndrome [87], suggesting inhibition of insulin secretion. Intriguingly, ghrelin's suppression of insulin secretion in pancreatic β -cells is mediated by a non-canonical GHSR1a signaling pathway in which $G\alpha_i$ rather than $G\alpha_q$ is coupled to the receptor [125]. This modified signaling is dependent upon agonist mediated molecular interactions between GHSR1a and somatostatin receptor subtype-5 (SST5) and the formation of GHSR1a:SST5 heterodimers [126]. Conversely, some studies suggest that ghrelin has positive trophic activity, protecting from β -cell damage in experimental models of type 1 diabetes [127,128]. However, the onset of type 1 diabetes is associated with decreased circulating ghrelin levels [129,130].

4.4. Clinical pharmacology studies on ghrelin's effect on GI-Motility

Very shortly after the discovery of ghrelin by Kojima and Kangawa, ghrelin was also described by another group, who named it motilin-related peptide because of its homology with motilin, a gut hormone involved in the regulation of the migrating motor complex (MMC) with effects on gastric emptying [131,132]. However, this group did not describe the octanoylation of ghrelin. It was soon hypothesized that ghrelin may mimic the effect of motilin on gastrointestinal motility. When ghrelin is administered iv into healthy individuals, it induces the MMC in the fasted state, inhibits gastric accommodation, and accelerates gastric emptying in the postprandial state [6,92,133,134]. Several clinical trials are currently investigating the potential of ghrelin mimetics in the treatment of hypomotility disorders (diabetic gastroparesis, postoperative ileus), but none of these has been marketed so far [135] (Table 1).

4.5. Rodent pharmacology studies on ghrelin's effect on food intake

Ghrelin is the only circulating hormone that, upon systemic and central administration, potently increases adiposity and food intake [2]. Similar to other GH secretagogues [136] the effect of ghrelin on adiposity is GH-independent and involves neural circuits that control food intake, energy expenditure, nutrient partitioning, and reward [3]. In the ARC, a key hypothalamic center regulating food intake and satiety [42], ghrelin increases the activity of *Npy* and *Agrp* expressing neurons while inhibiting the activity of proopiomelanocortin (*Pomc*) neurons [3]. NPY and AgRP are crucial for ghrelin's effect on feeding behavior as ghrelin fails to increase food intake in mice lacking both *Npy* and *Agrp* [137]. In line with this notion, AgRP neuron-selective GHSR re-expression in otherwise GHSR deficient mice partially restores the orexigenic response to administered ghrelin and fully restored the lowered blood glucose levels observed upon caloric restriction [138]. Of appreciable note, hyperphagia observed under pathophysiological conditions, such as streptozotocin-induced diabetes, is mediated by increased ghrelin release, which targets the ghrelin receptor on NPY and AgRP neurons [139]. A crucial regulator of *Npy* and *Agrp* expression is the hypothalamic homeobox domain transcription factor *Bsx* [140], which is also essential for ghrelin's ability to stimulate food intake [140].

4.6. Rodent pharmacology studies on ghrelin's effect on adiposity

The increased adiposity induced by the central administration of ghrelin involves the stimulation of key enzymes promoting fatty acid storage, while genes controlling the rate-limiting step in fat oxidation are decreased [141]. These actions of the brain ghrelin system on adipose tissue are mediated by the sympathetic nervous system independent of food intake or energy expenditure [141]. In addition to its actions on lipid metabolism in adipose tissue, chronic central infusion of ghrelin also increases plasma cholesterol levels, and more specifically HDL, an effect that is consistent with the fact that mice lacking both ghrelin and GHSR1a show lower plasma cholesterol levels than wild type mice [142]. Ghrelin's effects on adiposity, therefore, are achieved through centrally and peripherally mediated signaling mechanisms, including modulation of the hypothalamic melanocortinergic system and the food intake independent modulation of peripheral genetic programs regulating lipogenesis [141,143].

4.7. Rodent pharmacology studies using genetically engineered mouse models

4.7.1. Studies in mice with adult-onset ablation of ghrelin-producing cells

The group of Joseph Goldstein and Michael Brown recently generated transgenic mice in which the diphtheria toxin receptor is expressed in ghrelin-secreting cells. Adult-onset ablation of ghrelin-producing cells in these mice, following administration of diphtheria toxin, had no effect on food intake and body weight, indicating that while ghrelin has potent pharmacological effects on food intake, energy metabolism, and body weight, it is not an essential endogenous regulator of those endpoints [144].

Despite this finding, diet-induced obesity renders NPY/AgRP neurons unresponsive to the stimulatory actions of ghrelin on food intake [143,145], an effect that is reversed with diet-restricted weight loss [146]. However, mice continue to gain adiposity on a high-fat diet (HFD) [143]; thus, reinstatement of ghrelin sensitivity with diet-induced weight loss may provide a physiological means to protect a higher body weight set point established after prolonged HFD exposure. Moreover, selective reduction of the expression of GHSR1a in the paraventricular

nucleus of the hypothalamus (PVH) reduces body weight without affecting food intake [147], which supports the idea of two parallel ghrelin-responsive hypothalamic circuits that regulate food intake and adiposity independently. As yet, the differences in these neural circuits remain unknown.

4.7.2. Studies in ghrelin deficient mice

In recent years, studies using genetically engineered mouse models in which the function of the endogenous ghrelin system is altered either by loss [19,148–153] or gain of function [61,154] have contributed significantly to our knowledge about the multiple facets of ghrelin action. Results from these studies must be interpreted with caution, however, as most have utilized mice of mixed background (mainly S129/C57BL6J) and S129 favors a lean phenotype [155]. In one study, ghrelin deficient mice of mixed background were reported to have lower body weight and fat mass, which might be attributed to an observed increase in energy expenditure and locomotor activity [19]. Upon HFD exposure, these mice also show a lower respiratory quotient, indicating a shift in the metabolic fuel preference toward higher lipid utilization. While these data indicate that ghrelin promotes energy conservation by increasing carbohydrate metabolism while promoting fat storage in adipose tissue, other studies using mice of mixed background show no overt changes in body weight, adiposity, or food intake between ghrelin deficient mice and wt controls under chow-fed conditions [149,150]. However, when chronically exposed to a HFD, especially beginning at an early age, these same mice show a clear metabolic benefit from ghrelin-deficiency. It should be noted that this benefit is not evident in mature, congenic knock out (ko) mice on a pure C57BL/6J background [155]. The phenotype of loss-of-function models for ghrelin depends on environmental conditions. Under sub-thermoneutral conditions accompanied by fasting, ghrelin ko mice become compromised and are unable to integrate sleep and thermoregulatory responses to metabolic challenges [156]. When chronically challenged with a HFD, ghrelin deficient mice show improved glucose disposal and insulin sensitivity compared to wt controls [19]. When crossing ghrelin deficient mice with leptin deficient ob/ob mice [157], double mutants retain the marked body adiposity phenotype of ob/ob mice. However, the double mutants show a significant decrease in basal glucose and an increase in basal insulin levels, as well as improved glucose tolerance and insulin sensitivity when stimulated with a glucose or insulin challenge, compared to native ob/ob mice. In addition, fasting glucose levels are normalized in the double mutants, compared to ob/ob mice [157].

The lack of significant changes in food intake in ghrelin deficient mice does not support a role for ghrelin as an essential ‘meal initiation’ or ‘hunger’ hormone. Nevertheless, HFD exposure of ghrelin deficient mice reveals physiological roles for ghrelin in regulating body weight and adiposity, potentially through altering fat deposition and metabolism by decreasing fuel efficiency and increasing fat oxidation.

Such improvements in body weight homeostasis might lead indirectly to improvements in glucose homeostasis; in addition, ghrelin deficiency might directly improve glucose sensitivity and pancreatic β cell function. The effects of ghrelin on insulin sensitivity are at least partly mediated by the central nervous system, and more specifically by AgRP neurons within the ARC. Using a tamoxifen-inducible AgRP-CreER(T2) transgenic mouse model that allows spatiotemporally-controlled re-expression of physiological levels of ghrelin receptors (GHSRs) in AgRP neurons of adult GHSR-null mice that otherwise lack GHSR expression, it was found that AgRP neuron-selective GHSR re-expression fully restored the lowered blood glucose levels observed upon caloric restriction [138]. The restoration of glucose levels was

associated to glucagon rises and hepatic gluconeogenesis induction [138]

4.7.3. Studies in GHSR deficient mice

The orexigenic effect of ghrelin is specifically modulated through GHSR1a, as exogenous ghrelin fails to promote food intake in mice lacking this receptor [158] and in rats treated centrally with GHSR1a antagonists [147]. Despite the well-described actions of ghrelin on NPY and AgRP neurons, very little is known about the function of other hypothalamic neuronal populations expressing the GHSR. These include the ventromedial nucleus of the hypothalamus (VMH), dorso-medial nucleus of the hypothalamus and medial preoptic area [159,160]. Elucidating the function of these populations will highlight the role of ghrelin as being more than simply a “hunger hormone.”

As in mice lacking ghrelin, mice deficient for GHSR are protected from diet-induced obesity (DIO) when fed a HFD. This might be explained, in part, by a mild hypophagia and preferential utilization of fat as an energy substrate in these mice [151,152]. Expression of *GHSR* antisense RNA under the TH promoter in the ARC of rats results in hypophagia and decreased body weight and body fat [161]. Compared to wt littermate controls, *GHSR* deficient mice also show improved glucose disposal and insulin sensitivity upon HFD exposure. However, body weight and fat mass are not affected when male *GHSR* deficient mice are maintained on a standard chow diet [151,158]. Consistently, ablation of ghrelin receptor reduces adiposity and improves insulin sensitivity during aging by regulating fat metabolism in white and brown adipose tissues [25]. The effects of ghrelin on glucose metabolism during aging might be associated to GH levels, as it is known that circulating GH levels, which cause insulin resistance [162], are decreased in later stages.

Interestingly, simultaneous deletion of both ghrelin and *GHSR* results in lower body weight and fat mass even when the double mutant mice are fed a standard chow diet [153]. One possible explanation for this finding is the potential existence of additional ligands for the GHSR and/or additional receptors for ghrelin, which may exacerbate the metabolic phenotype of double mutant mice where ghrelin and GHSR are inactivated [153].

Another possibility is that GHSR may affect food intake independently of ghrelin signaling, e.g. by heterodimerization with other receptors such as the dopamine receptor [163] or GPR83 [164]. Of note, the impact of GHSR signaling on food intake, body weight, or energy and glucose homeostasis might be influenced by the receptor’s intrinsic constitutive activity [165], thus complicating the direct comparison of metabolic phenotypes of ghrelin and GHSR deficient mouse models. However, given the level of GHSR1a expression found in native tissues, it is doubtful that basal activity is a contributing factor. Furthermore, this interpretation might be confounded by the observation that GHSR1a, but not ghrelin, is essential for appetite regulation by dopamine receptor subtype-2 [166].

4.7.4. Studies in GOAT deficient mice

Generation of mice with the genetic inactivation of GOAT also has proven to be a useful tool to assess the role of des-acyl ghrelin without *in vivo* octanoylation [20,61,167]. Work by the laboratories of Joseph Goldstein and Michael Brown in young fat-depleted *GOAT* ko mice showed that acyl-ghrelin is of crucial importance for preventing life-threatening events of hypoglycemia under conditions of acute caloric restriction, an effect attributable to ghrelin’s ability to promote the release of GH from the anterior pituitary [20,168,169]. Accordingly, when body fat is reduced by caloric restriction, ghrelin stimulates GH secretion, which allows maintenance of glucose production, even

when food intake is eliminated. In line with this role of ghrelin to prevent hypoglycemia, adult-onset ablation of ghrelin producing cells induces profound hypoglycemia during prolonged caloric restriction [144]. Moreover, severe caloric restriction substantially increases plasma GH levels and promotes hepatic autophagy in wt mice, allowing the mice to maintain viable levels of blood glucose while lethal hypoglycemia and a blunted GH increase is observed in mice deficient for GOAT [170]. Hypoglycemia is also observed in GHSR-null mice following the same prolonged caloric restriction protocol [138] and upon initiation of acute caloric restriction of both ghrelin ko and *GHSR* ko mice, although both genotypes adapted after 14 days [155]. Also pharmacological inhibition of GOAT has been shown to improve glucose disposal and to enhance insulin secretion, an effect notably not seen in *GHSR* ko mice [60].

4.8. FTO and ghrelin

Several SNPs within the first intron of the fat mass and obesity-associated gene (*FTO*) are robustly associated with increased BMI and adiposity across different ages and populations [171–176]. Subjects homozygous for the obesity-risk (A) allele of SNP rs9939609 have a 1.7-fold increased risk for obesity and exhibit overall increased *ad libitum* food-intake [177–179], particularly fat consumption [177,179–181], and impaired satiety [182,183] compared to subjects homozygous for the low-risk (T) allele. Recently, a series of studies implicated ghrelin in mediating this altered feeding behavior. In two independent cohorts of normal-weight, adiposity-matched individuals with either *FTO* rs9939609 TT or the obesity risk AA genotype [184], AA subjects exhibited attenuated post-meal suppression of both hunger and circulating acyl-ghrelin levels. Using fMRI, these studies demonstrated that *FTO* rs9939609 genotype modulated the neural responses to food images in homeostatic and reward brain regions. Furthermore, AA and TT subjects exhibited divergent neural responsiveness to circulating acyl-ghrelin within brain regions that regulate appetite, reward-processing and incentive motivation. At the molecular level, *FTO* directly demethylates *N*⁶-methyladenosine (*m*⁶A), a naturally occurring adenosine modification in RNA and *ghrelin* mRNA has been identified as an *FTO* target [184,185]. *FTO* over-expression in MGN3-1 cells, a validated ghrelin cell line, reduced ghrelin mRNA *m*⁶A methylation, increased *ghrelin* mRNA abundance and the synthesis and secretion of acyl-ghrelin. Furthermore, subjects with the A allele of rs9939609 exhibit increased *FTO* expression [184,186] and decreased *ghrelin* *m*⁶A methylation coupled with increased *ghrelin* expression [184]. Interestingly, *FTO* also regulates the *m*⁶A methylation and expression of key molecular components of the mid-brain dopaminergic system, which is known to play a key role in mediating the rewarding effects of ghrelin [187]. Altered dopaminergic signaling may account for the altered neural ghrelin sensitivity reported in rs9939609 *FTO* AA subjects [184]. This suggests that the known actions of acyl-ghrelin, increased food intake, increased adiposity, preference for high-fat food, enhanced operant responding for food rewards, induced conditioned place preference for food rewards and a role in cue-potentiated feeding are strikingly similar to the feeding phenotype of rs9939609 AA subjects. However, while these findings of altered ghrelin function in *FTO* rs9939609 AA subjects provide a parsimonious explanation for the obesity risk phenotype seen in these subjects, given the pleiotropic effects of *FTO* a number of other mechanisms could also be implicated.

4.9. Additional functions of ghrelin

In addition to ghrelin's role in glucose and energy homeostasis, research over the last 15 years has revealed a surprising variety of

additional physiological functions of ghrelin in areas as distinct as learning and memory [188–190], psychological stress, mood and anxiety [191,192], depression [26,193,194], thymopoiesis [195], sleep/wake rhythm [7–9,196,197], and aging [198,199]. Recent pharmacological intervention trials also point to a neuroprotective role of ghrelin in neurodegenerative diseases (e.g., Parkinson's disease) [198,199]. The ghrelin system's neuroprotective effects are apparent in mouse models of chronic psychosocial stress, wherein stress-induced decreases in adult hippocampal neurogenesis become exaggerated in mice lacking GHSRs [200]. Additional studies examining the genetic and pharmacological modulation of the ghrelin system will help elucidate these novel roles of ghrelin.

5. GHRELIN ACTION IN THE BRAIN

5.1. Hypothalamic effects of ghrelin on energy metabolism

Although an important site of action of ghrelin on the control of food intake is the ARC, ghrelin administration into other hypothalamic sites, including the PVH [201,202] and the lateral hypothalamus [203] also promote a positive energy balance. In the hypothalamus, ghrelin triggers endocannabinoid release [204], leading to activation of the calcium/calmodulin-dependent protein kinase 2 (CaMKK2) and increased phosphorylation of the energy sensor AMP-activated protein kinase (AMPK) [205–207]. Ghrelin mediated activation of GHSR1a also triggers hypothalamic sirtuin 1 (Sirt1) [208,209], which deacetylates p53, leading to increased phosphorylated levels of AMP-activated protein kinase (AMPK) [206] and to the inactivation of enzymatic steps of *de novo* fatty acid biosynthetic pathway in the VMH [207]. These molecular events induce changes in uncoupling protein 2 (UCP2) [210] and the upregulation of the transcription factors Bsx [140], forkhead box O1 (FoxO1), and cAMP response-element binding protein (pCREB) [211] followed by subsequent activation of downstream signaling pathways. Within the hypothalamus, ghrelin increases expression of the prolyl carboxypeptidase (PRCP) a negative regulator of the melanocortin 4 receptor agonist α -melanocyte stimulating hormone (α -MSH) [212] and the mechanistic target of rapamycin (mTOR) in the ARC. In fact, central inhibition of mTOR signaling with rapamycin decreases ghrelin's orexigenic action [213].

5.2. Non-hypothalamic effects of ghrelin on energy metabolism

Ghrelin also promotes a positive energy balance when administered to non-hypothalamic sites such as the hindbrain [214–216] and limbic/paralimbic regions including the amygdala [202,217,218]. A recent study employed a genetically-engineered mouse model with *Ghsr* expression limited to the hindbrain to determine if such site-selective, hindbrain GHSR expression is sufficient to mediate ghrelin's actions on food intake and blood glucose [219]. When these animals were provided food *ad libitum*, hindbrain-specific GHSR expression was not sufficient to permit the characteristic orexigenic response to subcutaneous ghrelin administration that is observed in wt animals. With respect to the modulation of glucose homeostasis, hindbrain GHSR expression was sufficient to defend against the exacerbated fasting-induced fall in blood glucose that is otherwise observed in mice with global GHSR deficiency. These data help clarify the relevant sites of ghrelin receptor action in the brain in the modulation of food intake and blood glucose and complement a prior study investigating the effects of tyrosine hydroxylase-Cre-driven GHSR expression, in which GHSR expression occurs selectively in catecholaminergic (predominantly dopaminergic) neurons, such as those in the VTA [28]. Notably, and unlike with paired mesoderm homeobox 2B (Phox2b) cre-driven hindbrain *GHSR* expression, catecholaminergic *GHSR* expression is

sufficient to partially rescue ghrelin-stimulated acute food intake, while fully restoring the ability of administered ghrelin and chronic stress to modulate food reward [28]. Also, unlike with the hindbrain-selective *GHSR* expression, fasting blood glucose levels are not rescued by selective *GHSR* expression in catecholaminergic cells [28].

Several lines of evidence indicate that the brainstem contributes to ghrelin's orexigenic action, as peripheral administration of GHSs and intracerebroventricular (icv) ghrelin administration increase c-Fos expression in the nucleus tractus solitarius and the area postrema [220,221]. The role of the vagus nerve in the regulation of ghrelin-induced food intake is more controversial, however, as one study shows that blockade of gastric vagal afferents diminishes ghrelin's effect on food intake and decreases ghrelin induced c-Fos expression in the ARC [222], while another study reports that gut vagal afferents are not necessary for the hyperphagic action of ghrelin [223]. As gastrectomy is accompanied by vagotomy, the fact that ghrelin analogs are anabolic when given after gastrectomy suggests that the vagus is not essential for ghrelin's orexigenic effects [224,225].

5.3. Ghrelin and the reward system

Ghrelin engages reward neurocircuits that are activated by drugs of abuse [15,226–232]. In particular, the central ghrelin signaling system seems to be important for the rewarding properties of alcohol [228], nicotine [233,234] and cocaine [235]. One of the neurocircuits involved in these effects is the mesolimbic, dopaminergic pathway that projects from the VTA to the nucleus accumbens (NAc) [229,236], a pathway with a key role in reward-seeking behavior. Acting on this pathway, ghrelin affects the motivation and drive to eat. Ghrelin administration to the VTA and the NAc increases both food intake [232,237] and extracellular dopamine [13,238]. Underscoring the importance of dopamine for ghrelin's orexigenic effects is the finding that intra-VTA delivery of a ghrelin antagonist blocks the ability of parenteral ghrelin to increase feeding [239]. Dopamine modulates the incentive salience of food [240] and the animal's willingness to work for food [241]. In short, it increases feeding by increasing the drive, arousal, foraging, and motor hyperactivity that occur during food anticipation. For example, ghrelin ko mice do not show the normal anticipatory locomotion to scheduled meals [242,243]. Ghrelin also has direct effects on two other regions implicated in the control of feeding: the hippocampus and amygdala, where it facilitates learning and memory [188] and emotional arousal [244] and cue-potentiated feeding [189].

GHSR1a and dopamine receptor-2 (D2R) are present as GHSR1a:D2R heterodimers in native hypothalamic neurons and the inhibitory effects of D2R signaling on food intake is dependent on the presence of GHSR1a. *GHSR* ko mice and wt mice treated with a selective GHSR1a antagonist are resistant to the anorexigenic effects of a DRD2 agonist. Remarkably, ghrelin ko mice are fully sensitive to DRD2 agonist suppression of food intake, demonstrating a dependence on GHSR1a but not on ghrelin [163]. At the level of the VTA, but not the NAc, ghrelin increases motivation for food, reflected by an increased lever pressing for sucrose pellets in a progressive ratio task [231]. Interestingly, the VTA-driven effects of ghrelin on food motivation involve different neurocircuits than those involved in food intake. NAc delivery of dopamine receptor (D1R and D2R) antagonists blocked the effects of intra-VTA infused ghrelin on food motivation/reward behavior but not food intake, suggesting that the VTA-NAc dopamine reward pathway is important for food motivation but not food intake [245]. Given that central blockade or stimulation of the dopamine receptors 1, 2, and 3 suppress the effects of icv delivered ghrelin on food intake [246], it can be inferred that dopamine has a role outside of this classic reward

pathway to regulate ghrelin's orexigenic effects. Consistent with this, GHSR1a and D2R have been shown to interact within hypothalamic neurons blunting the anorexigenic actions of D2R agonism [163]. Divergence in the mesolimbic circuitry mediating ghrelin's orexigenic versus and food reward effects also occur at the VTA level and can be parsed using opioid and NPY Y1 receptor antagonists [232,236]. Interactions between ghrelin and the opioid system occur not only in the mesolimbic dopamine system but also in the hypothalamus. More precisely, GHSR1a and kappa opioid receptor colocalize in hypothalamic areas and the blockade of the kappa opioid receptor in the ARC is sufficient to blunt ghrelin-induced food intake [247]. Collectively, studies linking ghrelin to the mesolimbic reward circuitry suggest that ghrelin's role in hunger and meal initiation may extend to reward-driven behaviors, including food motivation.

5.4. The role of ghrelin learning and memory performance

Ghrelin exhibits dense receptor expression in the hippocampus [188], where it has been found to forms of learning and memory performance in rodents. For example, ghrelin administration has been shown to promote long term potentiation in the hippocampus, increase spine density of neurons in the hippocampal CA1 region, and enhance performance in several types of hippocampal-dependent learning and memory tasks [188,248]. Additionally, ghrelin has been shown to increase survival and reduce cell death of hippocampal neurons following ischemia/reperfusion injury [249]. Finally, it was recently shown that ghrelin cells receive direct synaptic input from the suprachiasmatic nucleus and the lateral geniculate nucleus, suggesting that ghrelin is implicated in mediating circadian and visual cues for the hypothalamic arousal system [250]

6. REGULATION OF GHRELIN SECRETION

Despite a growing body of literature characterizing ghrelin action and the distribution of ghrelin cells, relatively little is known about the exact molecular pathways responsible for the biosynthesis and release of ghrelin. Instead, most of what is known regarding the control of circulating ghrelin is on a broader, systemic level.

6.1. Ghrelin secretion in response to fasting and feeding

It has been known for several years that ghrelin levels rise preprandially and decrease to baseline levels within the first hour after a meal [37], a pattern that can be entrained by artificial meal schedules [251]. The magnitude of ghrelin reduction is proportional to the caloric load and macronutrient content, and ingested lipids are the least effective suppressor of plasma ghrelin [252]. Also, it is well established that plasma levels of both acyl and des-acyl ghrelin rise with prolonged food deprivation, increases that can be blocked by reserpine, which depletes adrenergic neurotransmitters from sympathetic neurons [253]. Sham feeding also suppresses ghrelin levels [254]. Furthermore, the recovery of ghrelin levels does not seem to be an important determinant of intermeal intervals [255], and mice that lack ghrelin have normal meal intervals.

6.2. Ghrelin levels in pathological conditions

It is well established that ghrelin plays a role in long-term energy balance regulation, defending against prolonged energy deficiency. Accordingly, in humans, circulating ghrelin levels are generally inversely associated with weight gain, adiposity, and insulin resistance [256] and positively correlated with weight loss induced by exercise, low-calorie diet, mixed life-style modification, anorexia nervosa and cachexia due to chronic obstructive lung disease (COPD) or chronic

heart failure (CHF) [71,257,258]. Ghrelin levels are low in obesity [259] and even lower in obese binge eaters [260], suggesting that, in these instances, ghrelin is a consequence rather than a cause of overeating. In line with the observation that ghrelin levels increase by fasting, plasma levels of ghrelin are high in patients with cachexia or in patients with eating disorders such as anorexia nervosa and bulimia nervosa [as reviewed in [166,258,261]]. Interestingly, extreme fasting reduces ghrelin levels in healthy subjects [262–264]. These effects are prevented by subdiaphragmatic vagotomy and, separately, by administration of the anticholinergic agent, atropine [265]. Conversely, obese patients with Prader-Willi syndrome are hyperphagic and have very high circulating ghrelin levels [266,267]. Elevated levels of ghrelin are further reported from patients with Hashimoto's Thyroiditis [268] but not from overweight/obese patients with Bardet-Biedl syndrome [269], Cushing's Disease [270,271], or HIV-Lipodystrophy [272]. Taken together, these data suggest that changes in circulating plasma levels of ghrelin may be relevant for the increase in adiposity in humans, although the degree of its contribution remains to be determined.

6.3. Ghrelin levels after bariatric surgery

Recent data demonstrate variable effects of various bariatric surgery procedures (i.e. Roux-en-Y gastric bypass, vertical sleeve gastrectomy, laparoscopic adjustable banding) on ghrelin levels (generally demonstrating decreases post-surgery) [273–279], shedding light on how ghrelin exerts its mechanistic effects in the gastrointestinal tract (reviewed in [280]). Low ghrelin levels have been reported for individuals after weight loss induced by Roux-en-Y gastric bypass, initially believed to play a key role in the decreased appetite observed after this surgery [257]. Subsequent data, however, show that ghrelin levels rise within the first year after this surgery in humans [281] and within 6 weeks after surgery in mice [282]. Additionally, compared to wt control mice, in ghrelin ko mice, vertical sleeve gastrectomy is equally efficient in lowering body weight [283], indicating a ghrelin independent effect in this type of bariatric surgery.

6.4. Ghrelin secretion in response to external food cues

Ghrelin release during fasting is mediated via activation of the autonomic nervous system. There is evidence that both cholinergic and adrenergic neurotransmission are involved in the release of ghrelin. Stimulation of ghrelin release in response to cholinergic activation by pharmacological substances or sham-feeding has been reported in humans and rodents [284–286]. In addition, food deprivation-induced elevation of plasma ghrelin levels is driven by an increased vagal efferent tone [284,285]. However, these results are not confirmed by *in vitro* studies either in a ghrelinoma cell line [287] or primary cell culture from rat stomach [288].

External food cues such as sight, smell, and taste trigger the cephalic phase of ingestive behavior, which consists of increased gut motility, gut hormone secretion, and autonomic arousal [289]. This response, in turn, triggers central arousal and incentive mechanisms that promote food consumption. The cephalic response includes ghrelin release, which increases after exposure to food cues in humans [290]. Conversely, recent evidence suggests that anticipation of the caloric content of an investigator-supplied milkshake modulates the postprandial reduction in ghrelin levels [291]. When subjects believed they were consuming a high calorie rather than a “healthy” milkshake, their ghrelin levels were much more reduced. In sum, ghrelin secretion is part of a CNS-gut control loop for feeding; food cues promote ghrelin release from the stomach, which feeds back to the CNS to activate hypothalamic and dopaminergic feeding centers. This feedback allows other factors such as chronic stress, negative energy balance, leptin

and insulin to affect motivation to feed by enhancing or reducing the cephalic release of ghrelin [290,292].

6.5. Suppression of ghrelin secretion

There has been significant interest in unveiling the mechanisms involved in postprandial suppression of circulating ghrelin levels. The placement of a pyloric cuff in rats to block normal flow of gastric contents into the duodenum prevents drops in circulating ghrelin usually observed following intragastric infusion of glucose [293]. Furthermore, stomach distention by infusion of water into animals whose gastric outflow was occluded at the level of the pylorus also was ineffective in changing ghrelin levels [294]. Thus, it appears that neither nutrient detection by the stomach nor gastric distention is sufficient for eliciting the usual postprandial fall in circulating ghrelin. On the other hand, both intraduodenal and intrajejunal administration of nutrients via intestinal cannulas lower circulating ghrelin levels. Within the stomach, considered the predominant source of circulating ghrelin, ghrelin cells tend to cluster towards the base of the gastric mucosal glands and are of the round, closed-type variety that do not have direct contact with gastric luminal contents (reviewed in [295]). There is also credible evidence that ghrelin cells exist, although in fewer numbers, throughout the entirety of the gastrointestinal tract, including the duodenum, where more elongated, opened-type ghrelin cells, which have direct contact with the intestinal lumen and may be regulated differently than their gastric counterparts.

6.6. Ghrelin secretion in response to dietary macronutrients

A recent randomized, within-subjects crossover human trial helped characterize the manner in which different types of nutrients influence the pattern of postprandial fluctuations in plasma ghrelin levels [296]. For this study, isocaloric, isovolemic beverages, composed primarily of carbohydrates, proteins, or lipids, were administered to volunteers whose plasma levels of acyl and total ghrelin were measured multiple times over the next 6 h. The lipid drink was the least effective, and the protein drink was most effective in lowering ghrelin levels. Although the carbohydrate drink resulted in the largest drop in ghrelin initially, it was the only drink to induce a subsequent rebound to above pre-prandial levels. Interestingly, ghrelin levels are also suppressed by sham feeding in humans [254], underscoring the role of the cephalic phase in the modification of ghrelin levels prior to and in response to a meal. Animal data suggest that these nutrient-associated decreases in circulating plasma ghrelin levels do not appear to involve the vagus nerve [223], which relays interoceptive sensory information from the viscera to the CNS and helps control visceral function. In fact, following intragastric gavage of a liquid diet to animals that had undergone subdiaphragmatic vagotomy, decreased ghrelin levels were observed [265]. However, the vagus nerve does seem to play a role in the rise of plasma ghrelin associated with a negative energy balance, as 48 h food deprivation-associated elevations in circulating ghrelin are prevented by subdiaphragmatic vagotomy or administration of atropine [265].

6.7. Hormones and neurotransmitter regulating ghrelin secretion

Recent studies have demonstrated that increases in ghrelin levels also occur upon acute or chronic stress that is not necessarily related to negative energy balance [reviewed in [26,28,297,298]]. Indeed, sympathetic activation increases ghrelin secretion [192]. Both *in vitro* and *in vivo* studies have demonstrated release of ghrelin in response to sympathetic stimulation mediated via β 1-adrenergic receptors present on the ghrelin cell [253,284,287,288,299].

To more directly study the determinants of ghrelin secretion, a recent study performed local infusion of candidate compounds into the gastric

submucosa followed by measurement of ghrelin mobilization via implanted microdialysis probes [300]. Using this method, epinephrine, norepinephrine, endothelin, and secretin were found to stimulate ghrelin release. In contrast to the stimulation of ghrelin release by activation of the sympathetic nervous system (SNS), the inhibition of ghrelin release seems primarily mediated by gastrointestinal hormones released during nutrient digestion, such as somatostatin and gastrin releasing peptide/bombesin. Numerous other hormones as well as many neurotransmitters, neuropeptides, glucose, and amino acids had no effect. Although this microdialysis technique and others, such as an *ex vivo* stomach explant culture system [294,301], help focus on locally acting compounds that influence ghrelin release, the techniques do not discriminate between compounds that act directly on ghrelin cells versus those that act indirectly via effects on neighboring cells. In summary, the regulation of ghrelin release is a complex process that is tightly controlled by both, the SNS and the gastrointestinal tract and which involves hormonal stimuli not necessarily involved in energy balance regulation. The observation that also factors not involved in systems metabolism regulate ghrelin secretion speaks for a broader physiological role of ghrelin and is in line with a multitude of ghrelin effects beyond the regulation of hunger and satiety. A potential beneficial effect of this complexity is the possibility to target the ghrelin system for the treatment of pathological conditions not necessarily related to a negative energy balance, such as e.g. gastroparesis. Recent studies showed that the ghrelin cell is chemosensory and contains taste receptors similar to those located in the tongue. Indeed, the ghrelin cell is co-localized with the gustatory G-proteins, α -gustducin and α -transducin. Studies in α -gustducin ko mice show that α -gustducin partially mediates the effect of bitter tastants on ghrelin release [302]. Similarly, the taste 1 receptor subtype, TAS1R3, involved in sensing both sugars and amino acids, is co-localized with ghrelin cells in the antrum [303]. The closed-type ghrelin cells in the stomach may receive chemosensory input from the bloodstream while the opened-type cells in the duodenum may respond to luminal stimuli. The long-chain fatty acid sensing receptor GPR120 is co-localized with ghrelin containing cells in the duodenum but not in the stomach and has been shown to play a role in the lipid-sensing cascade of the

ghrelin cell [68]. Free fatty acid receptor 1 (FFAR1), involved in sensing of long/medium chain fatty acids, is expressed only in the des-acyl (non-active)-containing ghrelin cell population in the stomach, and its function is unclear [68]. More studies are warranted to elucidate the role of taste receptors in the effects of nutrients on ghrelin secretion. It is likely that recent findings and new tools will provide greater insight into the regulation of ghrelin secretion (as reviewed in [304]). For instance, the development of novel, high-throughput sandwich enzyme linked immunosorbent assays or radioimmunoassays for the specific and sensitive detection of acyl-ghrelin as well as new mass spectrometry methods will permit accurate means to detect the different forms of circulating ghrelin and determine how various manipulations influence the levels of these different forms [305]. Another key development is the identification of GOAT, the enzyme responsible for catalyzing ghrelin's unique post-translational modification [56,57]. Recent work involving genetic manipulations of *GOAT* expression and the aforementioned mass spectrometry methods are challenging some of the accepted dogma about ghrelin secretion and regulation and suggest that ghrelin acylation and the secretion of acylated ghrelin represent two independent processes [61].

6.8. Ghrelin secretion regulated by G-protein coupled receptors

Over the last few years, several new ghrelin secretion models have been developed. Quantitative PCR analysis of FACS-purified gastric ghrelin cells identified a series of G-protein coupled receptors (GPCRs) that regulate ghrelin secretion. The GPCRs stimulating ghrelin secretion were mainly Gs-coupled and include the β 1-adrenergic receptor, the GIP receptor, the secretin receptor (SCTR) and, interestingly, the sensory neuropeptide receptor CGRP, and the melanocortin 4 receptor (MC4R) [306]. GPCRs inhibiting ghrelin release were Gq and/or Gi coupled and included the somatostatin receptors (SSTRs), the lactate receptor (GPR81) and receptors for short chain fatty acids (FFAR2) and long chain fatty acids (FFAR4) (Figure 4) [306].

6.9. Models to assess ghrelin secretion using transgenic mice

Other models developed to assess ghrelin secretion include genetically-engineered mouse models in which green fluorescence

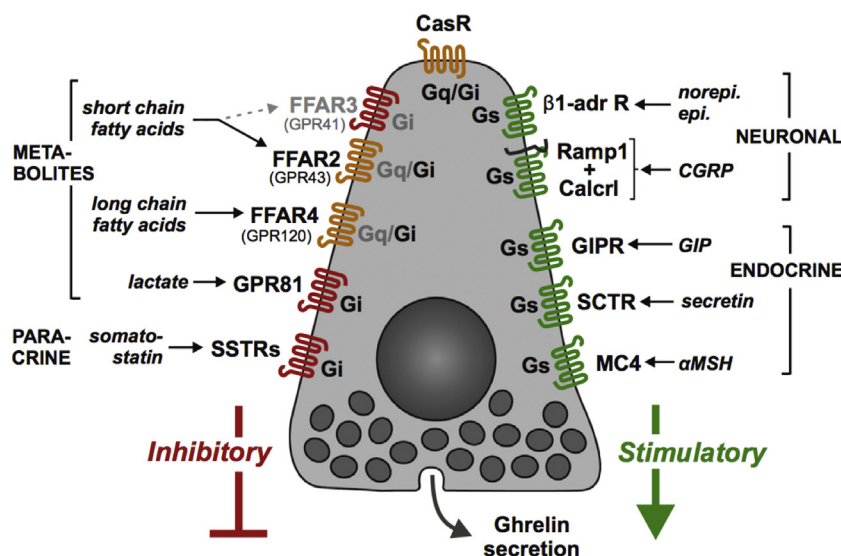


Figure 4: Schematic overview of the 7TM receptors judged to be either stimulating (in green to the right) or inhibiting (red or orange to the left and top) ghrelin secretion directly on the ghrelin cell. The main signaling pathway (*G α s* or *G α i*) employed by each of the receptors in the ghrelin cell is indicated inside the receptor in black. Figure taken from Engelstoft et al., *Mol Metab.* 2013 [303].

protein reports on the location of ghrelin-expressing cells. This method enables direct visualization of ghrelin cells, and fluorescence activated cell sorting-mediated isolation of ghrelin cells for expression analyses and cell culture [295,307,308]. Primary cell cultures of dispersed gastric mucosal cells from adult mice and 8-day-old rat pups also have been developed to investigate ghrelin secretion [288,307,309]. Ghrelin-secreting immortalized cell lines developed from ghrelinomas in the stomachs (SG-1, MGN3-1) and pancreatic islets (PG-1) of transgenic mice expressing SV40 large T-antigen under the control of preproghrelin promoter are now available [253,310]. These ghrelinoma cell lines retain many of the key, phenotypic features of ghrelin cells and respond to many of the same regulators of ghrelin secretion that have been described *in vivo* and in primary culture systems [253,288,307,310].

Using these models, the modulation of ghrelin release by peptide hormones, monoaminergic neurotransmitters, glucose, fatty acids, second messengers, potential downstream effector enzymes, and channels has now been investigated. Insulin, glucagon, oxytocin, somatostatin, dopamine, glucose, and long-chain fatty acids all have been shown to regulate ghrelin secretion through their direct interaction with ghrelin cells [253,288,307–310]. In addition, all of these models, as well as related *in vivo* studies, have been used to confirm that the catecholamines norepinephrine and epinephrine act as direct ghrelin secretagogues [253,287,288,299,307]. These data are supported by high levels of β 1-adrenergic receptor expression in ghrelin cells enriched from the stomach of ghrelin-green fluorescent protein reporter mice as well as in the SG-1 and PG-1 ghrelin cell lines [253]. Forskolin, a potent activator of adenylyl cyclase, mimics the effect of norepinephrine [253], suggesting that activation of adenylyl cyclase and an ensuing elevation of cAMP occurs following engagement of β 1-adrenergic receptors, as has been shown in other cell systems [311,312]. Interestingly, neuronal and endocrine signals have stimulatory effects on ghrelin secretion whereas paracrine signals and macronutrient metabolites such as fatty acids inhibit ghrelin release [306].

Altogether, these findings, along with the aforementioned microdialysis experiments, link fasting-induced stimulation of the sympathetic nervous system and ensuing release of norepinephrine locally in the stomach wall to the release of ghrelin [168,299]. New transgenic and cell culture models should allow for many more discoveries in the regulation of ghrelin secretion and other aspects of ghrelin cell physiology. In summary, recent studies reveal a comprehensive picture of the receptor repertoire expressed on the ghrelin cells, which allows for deeper analyses of the physiological properties and pharmacological potential of the ghrelin cell.

7. OPEN QUESTIONS

7.1. What is the role of des-acyl ghrelin?

7.1.1. Enzymes regulating des-acylation of ghrelin

Depending on the species, the serum half-life of acyl-ghrelin varies between 240 min in humans and 30 min in rats [313]. The differences in ghrelin's half-life might be explained by the fact that the enzymes responsible for the des-acylation and cleavage of ghrelin differ remarkably across species. Butyrylcholinesterase is the predominant enzyme responsible for ghrelin inactivation in humans whereas carboxylesterases allow for an eight times faster ghrelin des-octanoylation in rodents [313]. In rodents, des-octanoyl ghrelin is localized in two ghrelin cell populations in the stomach: cells that contain only des-octanoyl ghrelin and cells that contain both des-octanoyl and

octanoyl ghrelin [302,314]. Most studies have analyzed both des-acyl and acyl ghrelin finding that the majority of ghrelin in circulation is des-acylated. No receptor for des-acyl ghrelin has been identified and a recent study in which acyl ghrelin was assessed in human plasma using mass spectrometry suggests that all ghrelin in circulation is acylated and that des-acyl ghrelin may be an artifact of sample handling [315].

7.1.2. Effects of des-acyl ghrelin

Nevertheless, several studies suggest that des-acyl ghrelin promotes differentiation and fusion in C2C12 skeletal muscle cells [29], prevents muscle atrophy [30], elicits GHSR1 independent effects on energy and glucose metabolism [316–318], and exerts a cardioprotective effect on endothelial cells and cardiomyocytes [319,320]. Central infusion of des-acyl ghrelin into the third ventricle of rats increases short-term food intake, whereas peripheral administration of des-acyl ghrelin seems to have no direct effect on food intake [317]. The effect of des-acyl ghrelin on food intake is independent of GHSR1a and Npy signaling and might be orexin mediated [317]. Other studies, however, suggest that des-acyl ghrelin decreases food intake in rats and disrupts stomach motor activity under conditions of fasting [321]. Transgenic mice overexpressing des-acyl ghrelin have reduced body fat mass when fed a regular chow diet and are protected from diet-induced obesity when challenged with a HFD [321]. Although there are speculations about a potential receptor for des-acyl ghrelin located in the cardiovascular system [320], to date no such receptor has been identified.

Growing evidence points to a GHSR1a-independent role of des-acyl ghrelin in glucose metabolism, possibly antagonizing the effect of acyl-ghrelin. The often reported increase of plasma glucose levels and decrease of plasma insulin levels upon ghrelin administration [84–86,88,89] seem to be antagonized by co-administration of des-acyl ghrelin [322]. Several human studies report a positive relationship between des-acyl ghrelin and insulin sensitivity [323,324], although other studies do not support this finding [325]. The effect of des-acyl ghrelin on glucose metabolism might be triggered indirectly via modulation of lipid metabolism, as transgenic mice overexpressing des-acyl ghrelin have lower body fat mass, lower body weight gain, and improved insulin sensitivity compared to wt controls [318,326]. Furthermore, administration of des-acyl ghrelin decreases activation of gene programs regulating lipogenesis [327]. A more recent study in mice shows that chronic, subcutaneous administration of des-acyl ghrelin prevents the typical metabolic alterations caused by chronic HFD exposure, such as increased expression of pro-inflammatory cytokines and the development of HFD-induced glucose intolerance and insulin resistance [328]. Conversely, mice overexpressing ghrelin driven by the neuron-specific enolase (NSE) promoter develop age-related glucose intolerance despite having lower body weight [18]. Recent observations suggest a direct role on glucose metabolism based on observations indicating that des-acyl ghrelin promotes survival of pancreatic β -cells and as des-acyl ghrelin prevents the diabetogenic effect of streptozotocin [329–332]. Other data suggest that des-acyl ghrelin administered centrally to mice at high pharmacological doses acts to increase adiposity and glucose-stimulated plasma insulin through a GHSR-dependent mechanism [333].

7.2. Is ghrelin a 'hunger' hormone?

Ghrelin levels rise pre-prandially, and administered ghrelin reliably increases food intake in humans and rodents [2,94], supporting a role for ghrelin in hunger, meal initiation, and feeding behavior in normal physiology. The acute, orexigenic effects of ghrelin, however, are most

profound when ghrelin is delivered centrally, perhaps reflecting more widespread and simultaneous activation of diverse CNS sites. Indeed, orexigenic effects are observed not only after parenchymal delivery to the hypothalamic, brainstem and mesolimbic reward areas but also when administered to brain areas with a less well-established role in feeding control such as the amygdala [217], hippocampus [189] and dorsal raphe nucleus [334]. Recruitment of diverse feeding pathways by endogenous ghrelin may be under physiological control, perhaps reflecting food availability and/or nutritional status. Consistent with this, the ARC appears to show increased responsiveness to ghrelin in fasted rats relative to fed rats [335], and more ghrelin appears to gain CNS access in the fasted state [336].

It is not at all clear that ghrelin's acute orexigenic and chronic pro-obesity effects are coupled. While ghrelin may provide an acute hunger signal in the pre-prandial period, there is little evidence to suggest that sustaining high ghrelin levels induces hyperphagia in the long term. Despite numerous studies showing increased food intake upon acute or chronic systemic ghrelin application, mice deficient for ghrelin, *GHSR* or *GOAT*, nor transgenic mice overexpressing ghrelin and/or *GOAT* show alterations in food intake compared to wt controls [61,149,153]. Also, ghrelin antagonists, which were developed as anti-obesity drugs, do not appear to have chronic anorexigenic properties *per se* [337]. It may be helpful, therefore, to separate the suggested role for ghrelin as a hunger-promoting hormone in normal physiology from its therapeutically relevant, long-term obesogenic effects, which may be less linked to feeding control.

The surge in ghrelin before a meal could be linked to another role for ghrelin — to prepare the organism for incoming food in order to metabolize and store energy efficiently [61]. In line with this, ghrelin activation is influenced greatly by dietary lipids [61,63], and ghrelin might signal to the brain that abundant calories are available to acutely fill the organism's fuel stores.

One might argue that the typical increase of plasma ghrelin levels during prolonged food deprivation and the increase of ghrelin before a meal followed by the subsequent decrease afterwards clearly point to its role as a 'hunger' hormone. However, as discussed, the surges in ghrelin before a meal could also be explained by the theory that ghrelin prepares the organism for incoming food in order to metabolize and store energy efficiently [61]. In line with this, ghrelin activation is highly influenced by dietary lipids [61,63], and, therefore, ghrelin might signal to the brain that abundant calories are available to acutely fill the organism's fuel stores. The observation that ghrelin, independent of its effect on food intake, stimulates genetic programs regulating lipogenesis [141,143] is in line with this proposed role as a lipid sensor (as discussed in section 3.3).

7.3. Is the ghrelin receptor still a druggable target?

Ghrelin and its agonists appeal to those who desire to exploit the potent anabolic biology of the hormone. Such attention is directed at cachexic and frail states. Ghrelin has a very short half-life but the peptide can be engineered for a more sustained delivery and better pharmacokinetic properties. Treatment with ghrelin by infusion may be indicated in very acute circumstances when a short-term anabolic state is desired, such as prior to an elective surgery. The non-peptide GHSR1a agonists developed prior to the discovery of ghrelin are orally bioavailable; most importantly, they produce significant exposure levels for up to 24 h. The extended half-life of these compounds is of metabolic and functional importance, because, in contrast to ghrelin, chronic administration of non-peptide agonists results in sustained but modest increases in GH and IGF-1 superimposed on endogenous GH/IGF-1 without an increase in cortisol [338]. Interestingly, in obese subjects the anabolic effects of MK0677 produce an increase in the ratio of

lean/fat mass [339]. Chronic therapy with orally active stable GHSR1a agonists may have utility in frail, elderly subjects because MK0677 rejuvenates the GH/IGF1 axis by enhancing pulse amplitude of episodic GH release to match the physiological profile of young adults [338]. This effect is consistent with rescue of the epigenetic-mediated age-dependent decline in *GHSR* expression that reduces ghrelin sensitivity [340]. Indeed, encouraging results were observed in elderly patients recovering from hip fracture [341], with beneficial effects on skeletal muscle and bone density [80].

Other positive effects of ghrelin are reported in patients with cachexia, sarcopenia (muscle wasting due to aging [342]), myopenia (muscle wasting due to chronic illness [343]), and frailty states [343,344]. Among the first applications of ghrelin in human chronic illness were studies in CHF [34] and COPD [78,345]. A pilot study in 10 CHF patients reported decreased levels of norepinephrine, improved cardiac function and exercise capacity [34]. In animal models using a placebo control the cardiac effects could not be confirmed [346], but these models validated weight gain and the skeletal muscle anabolic effects of ghrelin and ghrelin analogs [110,347,348]. Of interest, in CHF, ghrelin secretion is modulated by application of brain natriuretic peptide, which is produced in the heart and generally increased in heart failure [349]. In advanced CHF, ghrelin resistance has been observed [350]. The first studies of ghrelin in COPD, focused on cachexia [78]. Preliminary studies suggested that ghrelin increases skeletal muscle mass and improves exercise capacity. In a recent double-blind controlled trial, ghrelin improved symptom scores and increased respiratory muscle strength [345]. An additional indication for ghrelin treatment may be replacement therapy to patients who have undergone total gastrectomy due to gastric cancer. Theoretically, this is based on the fact that most ghrelin is produced in the stomach and that total gastrectomy results in loss of appetite, body fat and also lean body mass. Indeed, proof of concept studies in mice and humans have yielded positive results [224,225].

Inhibition of GOAT-mediated ghrelin acylation is considered an interesting opportunity to tackle obesity. Several GOAT inhibitors have been developed [60,62], and their intraperitoneal administration promotes weight loss while improving glucose tolerance in wildtype (wt) mice but not in mice deficient for ghrelin [60]. Ghrelin receptor agonists such as BIM-28131 (a.k.a. RM-131) have sustained effects in increasing body weight at long-term [351]. Chronic therapy with ghrelin agonists, however, is associated with weight gain, fat attrition, and insulin resistance. Such observations have led to drug discovery efforts designed to block ghrelin action, inducing a negative energy balance with a goal of treating obesity and insulin resistance. Although ghrelin levels are lower in obesity, that circulating ghrelin levels increase during a negative energy state may suggest that a method inhibiting ghrelin activity may be useful for preventing weight regain after diet and exercise (or another weight loss treatment) rather than as a weight loss therapeutic. The greatest utility of des-acyl ghrelin, in fact, appears to be for the treatment of insulin resistance, but only when injected in combination with ghrelin.

Several synthetic ghrelin mimetics are being pursued in clinical trials for diverse indications (Table 1). Three compounds are currently in development. Macimorelin is in clinical trials for the diagnosis of GH deficiency, relying on the stimulation of the hypothalamic pituitary axis, as described earlier in this review [352]. A second compound, Anamorelin, is in clinical trials for the treatment of cancer cachexia in the treatment of non-small cell lung cancer. Anamorelin mechanistically relies on the anabolic effects noted with ghrelin and ghrelin mimetics [353]. Ghrelin and synthetic ghrelin mimetics can also stimulate gastric emptying and can function as gastrointestinal prokinetics [354–357].

While our understanding of the mechanism by which ghrelin elicits these effects is still limited, a direct effect of the ghrelin agonist Relamorelin has been shown on human and mouse fundus and jejunal smooth muscle cells and human and rodent colonic circular smooth muscle [357], where expression of the GHSR has been described [358]. The ghrelin mimetic Relamorelin (also known as RM-131) is being developed for the treatment of diabetic gastroparesis [357], for which the compound is well tolerated, and has direct beneficial effects on gastric emptying, while reducing the incidence of nausea and vomiting. Beneficial effects have also been noted for colonic motility disorders where prokinetic effects have been demonstrated in humans in the colon [354]. Relamorelin is currently in Phase II clinical trials for treatment of diabetic gastroparesis and other gastrointestinal (GI) disorders [93]. In a rodent model of irritable bowel syndrome, ghrelin [359] and synthetic ghrelin mimetics [93] show improvements in tissue damage while modulating inflammatory responses. Synthetic ghrelin mimetics, therefore, may find beneficial applications in diverse functional gastrointestinal disorders.

8. CONCLUSIONS

Since ghrelin was discovered in 1999, we have come a long way in understanding ghrelin's multifaceted nature. Numerous studies on ghrelin's physiological effects (Figure 3) have led us to new aspects of human and animal physiology and revealed a complex system for acylating hormones, which was previously unknown. The next era should exploit this unique biology for diagnostic and therapeutic benefit.

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CONFLICT OF INTEREST

The authors wish to declare that Dr. L Van der Ploeg is an employee of Rhythm Pharmaceuticals, a privately held Biotechnology company, which is developing RM-131 for the treatment of diverse functional gastrointestinal disorders.

REFERENCES

- [1] Kojima, M., Hosoda, H., Date, Y., Nakazato, M., Matsuo, H., Kangawa, K., 1999. Ghrelin is a growth-hormone-releasing acylated peptide from stomach. *Nature* 402:656–660.
- [2] Tschop, M., Smiley, D.L., Heiman, M.L., 2000. Ghrelin induces adiposity in rodents. *Nature* 407:908–913.
- [3] Cowley, M.A., Smith, R.G., Diano, S., Tschop, M., Pronchuk, N., Grove, K.L., et al., 2003. The distribution and mechanism of action of ghrelin in the CNS demonstrates a novel hypothalamic circuit regulating energy homeostasis. *Neuron* 37:649–661.
- [4] Nakazato, M., Murakami, N., Date, Y., Kojima, M., Matsuo, H., Kangawa, K., et al., 2001. A role for ghrelin in the central regulation of feeding. *Nature* 409:194–198.

- [5] Masuda, Y., Tanaka, T., Inomata, N., Ohnuma, N., Tanaka, S., Itoh, Z., et al., 2000. Ghrelin stimulates gastric acid secretion and motility in rats. *Biochemical and Biophysical Research Communications* 276:905–908.
- [6] Asakawa, A., Inui, A., Kaga, T., Yuzuriha, H., Nagata, T., Ueno, N., et al., 2001. Ghrelin is an appetite-stimulatory signal from stomach with structural resemblance to motilin. *Gastroenterology* 120:337–345.
- [7] Tolle, V., Bassant, M.H., Zizzari, P., Poindessous-Jazat, F., Tomasetto, C., Epelbaum, J., et al., 2002. Ultradian rhythmicity of ghrelin secretion in relation with GH, feeding behavior, and sleep-wake patterns in rats. *Endocrinology* 143:1353–1361.
- [8] Weikel, J.C., Wichniak, A., Ising, M., Brunner, H., Friess, E., Held, K., et al., 2003. Ghrelin promotes slow-wave sleep in humans. *American Journal of Physiology Endocrinology and Metabolism* 284:E407–E415.
- [9] Szentirmai, E., Hajdu, I., Obal Jr., F., Krueger, J.M., 2006. Ghrelin-induced sleep responses in ad libitum fed and food-restricted rats. *Brain Research* 1088:131–140.
- [10] Cai, H., Cong, W.N., Daimon, C.M., Wang, R., Tschop, M.H., Sevigny, J., et al., 2013. Altered lipid and salt taste responsivity in ghrelin and GOAT null mice. *PLoS One* 8:e76553.
- [11] Druce, M.R., Wren, A.M., Park, A.J., Milton, J.E., Patterson, M., Frost, G., et al., 2005. Ghrelin increases food intake in obese as well as lean subjects. *International Journal of Obesity (London)* 29:1130–1136.
- [12] Jerlhag, E., Egecioglu, E., Dickson, S.L., Andersson, M., Svensson, L., Engel, J.A., 2006. Ghrelin stimulates locomotor activity and accumbal dopamine-overflow via central cholinergic systems in mice: implications for its involvement in brain reward. *Addiction Biology* 11:45–54.
- [13] Jerlhag, E., Egecioglu, E., Dickson, S.L., Douhan, A., Svensson, L., Engel, J.A., 2007. Ghrelin administration into tegmental areas stimulates locomotor activity and increases extracellular concentration of dopamine in the nucleus accumbens. *Addiction Biology* 12:6–16.
- [14] Overduin, J., Figlewicz, D.P., Bennett-Jay, J., Kittleson, S., Cummings, D.E., 2012. Ghrelin increases the motivation to eat, but does not alter food palatability. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology* 303:R259–R269.
- [15] Skibicka, K.P., Hansson, C., Egecioglu, E., Dickson, S.L., 2012. Role of ghrelin in food reward: impact of ghrelin on sucrose self-administration and mesolimbic dopamine and acetylcholine receptor gene expression. *Addiction Biology* 17:95–107.
- [16] Tong, J., Mannea, E., Aime, P., Pfluger, P.T., Yi, C.X., Castaneda, T.R., et al., 2011. Ghrelin enhances olfactory sensitivity and exploratory sniffing in rodents and humans. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience* 31:5841–5846.
- [17] Date, Y., Nakazato, M., Hashiguchi, S., Dezaki, K., Mondal, M.S., Hosoda, H., et al., 2002. Ghrelin is present in pancreatic alpha-cells of humans and rats and stimulates insulin secretion. *Diabetes* 51:124–129.
- [18] Reed, J.A., Benoit, S.C., Pfluger, P.T., Tschop, M.H., D'Alessio, D.A., Seeley, R.J., 2008. Mice with chronically increased circulating ghrelin develop age-related glucose intolerance. *American Journal of Physiology Endocrinology and Metabolism* 294:E752–E760.
- [19] Wortley, K.E., del Rincon, J.P., Murray, J.D., Garcia, K., Iida, K., Thorne, M.O., et al., 2005. Absence of ghrelin protects against early-onset obesity. *The Journal of Clinical Investigation* 115:3573–3578.
- [20] Zhao, T.J., Liang, G., Li, R.L., Xie, X., Sleeman, M.W., Murphy, A.J., et al., 2010. Ghrelin O-acyltransferase (GOAT) is essential for growth hormone-mediated survival of calorie-restricted mice. *Proceedings of the National Academy of Sciences of the United States of America* 107:7467–7472.
- [21] Yasuda, T., Masaki, T., Kakuma, T., Yoshimatsu, H., 2003. Centrally administered ghrelin suppresses sympathetic nerve activity in brown adipose tissue of rats. *Neuroscience Letters* 349:75–78.
- [22] Tsubone, T., Masaki, T., Katsuragi, I., Tanaka, K., Kakuma, T., Yoshimatsu, H., 2005. Ghrelin regulates adiposity in white adipose tissue and

- UCP1 mRNA expression in brown adipose tissue in mice. *Regulatory Peptides* 130:97–103.
- [23] Mano-Otagiri, A., Ohata, H., Iwasaki-Sekino, A., Nemoto, T., Shibasaki, T., 2009. Ghrelin suppresses noradrenaline release in the brown adipose tissue of rats. *The Journal of Endocrinology* 201:341–349.
- [24] Mano-Otagiri, A., Iwasaki-Sekino, A., Nemoto, T., Ohata, H., Shuto, Y., Nakabayashi, H., et al., 2010. Genetic suppression of ghrelin receptors activates brown adipocyte function and decreases fat storage in rats. *Regulatory Peptides* 160:81–90.
- [25] Lin, L., Saha, P.K., Ma, X., Henshaw, I.O., Shao, L., Chang, B.H., et al., 2011. Ablation of ghrelin receptor reduces adiposity and improves insulin sensitivity during aging by regulating fat metabolism in white and brown adipose tissues. *Aging Cell* 10:996–1010.
- [26] Lutter, M., Sakata, I., Osborne-Lawrence, S., Rovinsky, S.A., Anderson, J.G., Jung, S., et al., 2008. The orexigenic hormone ghrelin defends against depressive symptoms of chronic stress. *Nature Neuroscience* 11:752–753.
- [27] Spencer, S.J., Xu, L., Clarke, M.A., Lemus, M., Reichenbach, A., Geenen, B., et al., 2012. Ghrelin regulates the hypothalamic-pituitary-adrenal axis and restricts anxiety after acute stress. *Biological Psychiatry* 72:457–465.
- [28] Chuang, J.C., Perello, M., Sakata, I., Osborne-Lawrence, S., Savitt, J.M., Lutter, M., et al., 2011. Ghrelin mediates stress-induced food-reward behavior in mice. *The Journal of Clinical Investigation* 121:2684–2692.
- [29] Filigheddu, N., Gnocchi, V.F., Coscia, M., Cappelli, M., Porporato, P.E., Taulli, R., et al., 2007. Ghrelin and des-acyl ghrelin promote differentiation and fusion of C2C12 skeletal muscle cells. *Molecular Biology of the Cell* 18:986–994.
- [30] Porporato, P.E., Filigheddu, N., Reano, S., Ferrara, M., Angelino, E., Gnocchi, V.F., et al., 2013. Acylated and unacylated ghrelin impair skeletal muscle atrophy in mice. *The Journal of Clinical Investigation* 123:611–622.
- [31] Rizzo, M., Rizvi, A.A., Sudar, E., Soskic, S., Obradovic, M., Montalto, G., et al., 2013. A review of the cardiovascular and anti-atherogenic effects of Ghrelin. *Current Pharmaceutical Design* 19:4953–4963.
- [32] Nagaya, N., Uematsu, M., Kojima, M., Ikeda, Y., Yoshihara, F., Shimizu, W., et al., 2001. Chronic administration of ghrelin improves left ventricular dysfunction and attenuates development of cardiac cachexia in rats with heart failure. *Circulation* 104:1430–1435.
- [33] Okumura, H., Nagaya, N., Enomoto, M., Nakagawa, E., Oya, H., Kangawa, K., 2002. Vasodilatory effect of ghrelin, an endogenous peptide from the stomach. *Journal of Cardiovascular Pharmacology* 39:779–783.
- [34] Nagaya, N., Moriya, J., Yasumura, Y., Uematsu, M., Ono, F., Shimizu, W., et al., 2004. Effects of ghrelin administration on left ventricular function, exercise capacity, and muscle wasting in patients with chronic heart failure. *Circulation* 110:3674–3679.
- [35] Cummings, D.E., 2006. Ghrelin and the short- and long-term regulation of appetite and body weight. *Physiology & Behavior* 89:71–84.
- [36] Cummings, D.E., Frayo, R.S., Marmonier, C., Aubert, R., Chapelot, D., 2004. Plasma ghrelin levels and hunger scores in humans initiating meals voluntarily without time- and food-related cues. *American Journal of Physiology Endocrinology and Metabolism* 287:E297–E304.
- [37] Cummings, D.E., Purnell, J.Q., Frayo, R.S., Schmidova, K., Wisse, B.E., Weigle, D.S., 2001. A preprandial rise in plasma ghrelin levels suggests a role in meal initiation in humans. *Diabetes* 50:1714–1719.
- [38] Kojima, M., Kangawa, K., 2005. Ghrelin: structure and function. *Physiological Reviews* 85:495–522.
- [39] Tschoop, M., Wawarta, R., Riepl, R.L., Friedrich, S., Bidlingmaier, M., Landgraf, R., et al., 2001. Post-prandial decrease of circulating human ghrelin levels. *Journal of Endocrinological Investigation* 24:RC19–21.
- [40] Guan, X.M., Yu, H., Palyha, O.C., McKee, K.K., Feighner, S.D., Sirinathsinghji, D.J., et al., 1997. Distribution of mRNA encoding the growth hormone secretagogue receptor in brain and peripheral tissues. *Brain Research Molecular Brain Research* 48:23–29.
- [41] Dickson, S.L., Luckman, S.M., 1997. Induction of c-fos messenger ribonucleic acid in neuropeptide Y and growth hormone (GH)-releasing factor neurons in the rat arcuate nucleus following systemic injection of the GH secretagogue, GH-releasing peptide-6. *Endocrinology* 138:771–777.
- [42] Willesen, M.G., Kristensen, P., Romer, J., 1999. Co-localization of growth hormone secretagogue receptor and NPY mRNA in the arcuate nucleus of the rat. *Neuroendocrinology* 70:306–316.
- [43] Bowers, C.Y., Momany, F., Reynolds, G.A., Chang, D., Hong, A., Chang, K., 1980. Structure-activity relationships of a synthetic pentapeptide that specifically releases growth hormone in vitro. *Endocrinology* 106:663–667.
- [44] Momany, F.A., Bowers, C.Y., Reynolds, G.A., Chang, D., Hong, A., Newlander, K., 1981. Design, synthesis, and biological activity of peptides which release growth hormone in vitro. *Endocrinology* 108:31–39.
- [45] Smith, R.G., Van der Ploeg, L.H., Howard, A.D., Feighner, S.D., Cheng, K., Hickey, G.J., et al., 1997. Peptidomimetic regulation of growth hormone secretion. *Endocrine Reviews* 18:621–645.
- [46] Dickson, S.L., Leng, G., Robinson, I.C., 1993. Systemic administration of growth hormone-releasing peptide activates hypothalamic arcuate neurons. *Neuroscience* 53:303–306.
- [47] Blake, A.D., Smith, R.G., 1991. Desensitization studies using perfused rat pituitary cells show that growth hormone-releasing hormone and His-D-Trp-Ala-Trp-D-Phe-Lys-NH₂ stimulate growth hormone release through distinct receptor sites. *The Journal of Endocrinology* 129:11–19.
- [48] Bowers, C.Y., Reynolds, G.A., Durham, D., Barrera, C.M., Pezzoli, S.S., Thorner, M.O., 1990. Growth hormone (GH)-releasing peptide stimulates GH release in normal men and acts synergistically with GH-releasing hormone. *The Journal of Clinical Endocrinology and Metabolism* 70:975–982.
- [49] Cheng, K., Chan, W.W., Butler, B., Barreto Jr., A., Smith, R.G., 1991. Evidence for a role of protein kinase-C in His-D-Trp-Ala-Trp-D-Phe-Lys-NH₂-induced growth hormone release from rat primary pituitary cells. *Endocrinology* 129:3337–3342.
- [50] Howard, A.D., Feighner, S.D., Cully, D.F., Arena, J.P., Liberators, P.A., Rosenblum, C.I., et al., 1996. A receptor in pituitary and hypothalamus that functions in growth hormone release. *Science* 273:974–977.
- [51] Gnanapavan, S., Kola, B., Bustin, S.A., Morris, D.G., McGee, P., Fairclough, P., et al., 2002. The tissue distribution of the mRNA of ghrelin and subtypes of its receptor, GHS-R, in humans. *The Journal of Clinical Endocrinology and Metabolism* 87:2988.
- [52] Jiang, H., Betancourt, L., Smith, R.G., 2006. Ghrelin amplifies dopamine signaling by cross talk involving formation of growth hormone secretagogue receptor/dopamine receptor subtype 1 heterodimers. *Molecular Endocrinology* 20:1772–1785.
- [53] Leung, P.K., Chow, K.B., Lau, P.N., Chu, K.M., Chan, C.B., Cheng, C.H., et al., 2007. The truncated ghrelin receptor polypeptide (GHS-R1b) acts as a dominant-negative mutant of the ghrelin receptor. *Cellular Signalling* 19:1011–1022.
- [54] Zhang, J.V., Ren, P.G., Avsian-Kretschmer, O., Luo, C.W., Rauch, R., Klein, C., et al., 2005. Obestatin, a peptide encoded by the ghrelin gene, opposes ghrelin's effects on food intake. *Science* 310:996–999.
- [55] Depoortere, I., 2012. GI functions of GPR39: novel biology. *Current Opinion in Pharmacology* 12:647–652.
- [56] Gutierrez, J.A., Solenberg, P.J., Perkins, D.R., Willency, J.A., Knierman, M.D., Jin, Z., et al., 2008. Ghrelin octanoylation mediated by an orphan lipid transferase. *Proceedings of the National Academy of Sciences of the United States of America* 105:6320–6325.
- [57] Yang, J., Brown, M.S., Liang, G., Grishin, N.V., Goldstein, J.L., 2008. Identification of the acyltransferase that octanoylates ghrelin, an appetite-stimulating peptide hormone. *Cell* 132:387–396.
- [58] Sakata, I., Yang, J., Lee, C.E., Osborne-Lawrence, S., Rovinsky, S.A., Elmquist, J.K., et al., 2009. Colocalization of ghrelin O-acyltransferase and ghrelin in gastric mucosal cells. *American Journal of Physiology Endocrinology and Metabolism* 297:E134–E141.

- [59] Lim, C.T., Kola, B., Grossman, A., Korbonits, M., 2011. The expression of ghrelin O-acyltransferase (GOAT) in human tissues. *Endocrine Journal* 58:707–710.
- [60] Barnett, B.P., Hwang, Y., Taylor, M.S., Kirchner, H., Pfluger, P.T., Bernard, V., et al., 2010. Glucose and weight control in mice with a designed ghrelin O-acyltransferase inhibitor. *Science* 330:1689–1692.
- [61] Kirchner, H., Gutierrez, J.A., Solenberg, P.J., Pfluger, P.T., Czyzyk, T.A., Willency, J.A., et al., 2009. GOAT links dietary lipids with the endocrine control of energy balance. *Nature Medicine* 15:741–745.
- [62] Yang, J., Zhao, T.J., Goldstein, J.L., Brown, M.S., 2008. Inhibition of ghrelin O-acyltransferase (GOAT) by octanoylated pentapeptides. *Proceedings of the National Academy of Sciences of the United States of America* 105:10750–10755.
- [63] Nishi, Y., Hiejima, H., Hosoda, H., Kaiya, H., Mori, K., Fukue, Y., et al., 2005. Ingested medium-chain fatty acids are directly utilized for the acyl modification of ghrelin. *Endocrinology* 146:2255–2264.
- [64] Sakata, I., Nakamura, K., Yamazaki, M., Matsubara, M., Hayashi, Y., Kangawa, K., et al., 2002. Ghrelin-producing cells exist as two types of cells, closed- and opened-type cells, in the rat gastrointestinal tract. *Peptides* 23:531–536.
- [65] Armand, M., Hamosh, M., Mehta, N.R., Angelus, P.A., Philpott, J.R., Henderson, T.R., et al., 1996. Effect of human milk or formula on gastric function and fat digestion in the premature infant. *Pediatric Research* 40:429–437.
- [66] Heppner, K.M., Chaudhary, N., Muller, T.D., Kirchner, H., Habegger, K.M., Ottaway, N., et al., 2012. Acylation type determines ghrelin's effects on energy homeostasis in rodents. *Endocrinology* 153:4687–4695.
- [67] Ohgusu, H., Shirouzu, K., Nakamura, Y., Nakashima, Y., Ida, T., Sato, T., et al., 2009. Ghrelin O-acyltransferase (GOAT) has a preference for n-hexanoyl-CoA over n-octanoyl-CoA as an acyl donor. *Biochemical and Biophysical Research Communications* 386:153–158.
- [68] Janssen, S., Laermans, J., Iwakura, H., Tack, J., Depoortere, I., 2012. Sensing of fatty acids for octanoylation of ghrelin involves a gustatory G-protein. *PLoS One* 7:e40168.
- [69] Takaya, K., Ariyasu, H., Kanamoto, N., Iwakura, H., Yoshimoto, A., Harada, M., et al., 2000. Ghrelin strongly stimulates growth hormone release in humans. *The Journal of Clinical Endocrinology and Metabolism* 85:4908–4911.
- [70] Arvat, E., Maccario, M., Di Vito, L., Broglio, F., Benso, A., Gottero, C., et al., 2001. Endocrine activities of ghrelin, a natural growth hormone secretagogue (GHS), in humans: comparison and interactions with hexarelin, a nonnatural peptidyl GHS, and GH-releasing hormone. *The Journal of Clinical Endocrinology and Metabolism* 86:1169–1174.
- [71] Nagaya, N., Kojima, M., Uematsu, M., Yamagishi, M., Hosoda, H., Oya, H., et al., 2001. Hemodynamic and hormonal effects of human ghrelin in healthy volunteers. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology* 280:R1483–R1487.
- [72] Hataya, Y., Akamizu, T., Takaya, K., Kanamoto, N., Ariyasu, H., Saijo, M., et al., 2001. A low dose of ghrelin stimulates growth hormone (GH) release synergistically with GH-releasing hormone in humans. *The Journal of Clinical Endocrinology and Metabolism* 86:4552.
- [73] Mericq, V., Cassorla, F., Bowers, C.Y., Avila, A., Gonen, B., Merriam, G.R., 2003. Changes in appetite and body weight in response to long-term oral administration of the ghrelin agonist GHRP-2 in growth hormone deficient children. *Journal of Pediatric Endocrinology & Metabolism: JPEM* 16:981–985.
- [74] Schmid, D.A., Held, K., Ising, M., Uhr, M., Weikel, J.C., Steiger, A., 2005. Ghrelin stimulates appetite, imagination of food, GH, ACTH, and cortisol, but does not affect leptin in normal controls. *Neuropsychopharmacology: Official Publication of the American College of Neuropsychopharmacology* 30:1187–1192.
- [75] Hotta, M., Ohwada, R., Akamizu, T., Shibasaki, T., Takano, K., Kangawa, K., 2009. Ghrelin increases hunger and food intake in patients with restricting-type anorexia nervosa: a pilot study. *Endocrine Journal* 56:1119–1128.
- [76] Wynne, K., Giannitsopoulou, K., Small, C.J., Patterson, M., Frost, G., Ghatei, M.A., et al., 2005. Subcutaneous ghrelin enhances acute food intake in malnourished patients who receive maintenance peritoneal dialysis: a randomized, placebo-controlled trial. *Journal of the American Society of Nephrology: JASN* 16:2111–2118.
- [77] Ashby, D.R., Ford, H.E., Wynne, K.J., Wren, A.M., Murphy, K.G., Busbridge, M., et al., 2009. Sustained appetite improvement in malnourished dialysis patients by daily ghrelin treatment. *Kidney International* 76:199–206.
- [78] Nagaya, N., Itoh, T., Murakami, S., Oya, H., Uematsu, M., Miyatake, K., et al., 2005. Treatment of cachexia with ghrelin in patients with COPD. *Chest* 128:1187–1193.
- [79] Neary, N.M., Small, C.J., Wren, A.M., Lee, J.L., Druce, M.R., Palmieri, C., et al., 2004. Ghrelin increases energy intake in cancer patients with impaired appetite: acute, randomized, placebo-controlled trial. *The Journal of Clinical Endocrinology and Metabolism* 89:2832–2836.
- [80] Nass, R., Pezzoli, S.S., Oliveri, M.C., Patrie, J.T., Harrell Jr., F.E., Clasey, J.L., et al., 2008. Effects of an oral ghrelin mimetic on body composition and clinical outcomes in healthy older adults: a randomized trial. *Annals of Internal Medicine* 149:601–611.
- [81] Ravussin, E., Tschop, M., Morales, S., Bouchard, C., Heiman, M.L., 2001. Plasma ghrelin concentration and energy balance: overfeeding and negative energy balance studies in twins. *The Journal of Clinical Endocrinology and Metabolism* 86:4547–4551.
- [82] Lejeune, M.P., Westerterp, K.R., Adam, T.C., Luscombe-Marsh, N.D., Westerterp-Plantenga, M.S., 2006. Ghrelin and glucagon-like peptide 1 concentrations, 24-h satiety, and energy and substrate metabolism during a high-protein diet and measured in a respiration chamber. *The American Journal of Clinical Nutrition* 83:89–94.
- [83] Huda, M.S., Dovey, T., Wong, S.P., English, P.J., Halford, J., McCulloch, P., et al., 2009. Ghrelin restores 'lean-type' hunger and energy expenditure profiles in morbidly obese subjects but has no effect on postgastrectomy subjects. *International Journal of Obesity (London)* 33:317–325.
- [84] Broglio, F., Arvat, E., Benso, A., Gottero, C., Muccioli, G., Papotti, M., et al., 2001. Ghrelin, a natural GH secretagogue produced by the stomach, induces hyperglycemia and reduces insulin secretion in humans. *The Journal of Clinical Endocrinology and Metabolism* 86:5083–5086.
- [85] Broglio, F., Gottero, C., Benso, A., Prodam, F., Destefanis, S., Gauna, C., et al., 2003. Effects of ghrelin on the insulin and glycemic responses to glucose, arginine, or free fatty acids load in humans. *The Journal of Clinical Endocrinology and Metabolism* 88:4268–4272.
- [86] Egido, E.M., Rodriguez-Gallardo, J., Silvestre, R.A., Marco, J., 2002. Inhibitory effect of ghrelin on insulin and pancreatic somatostatin secretion. *European Journal of Endocrinology/European Federation of Endocrine Societies* 146:241–244.
- [87] Fusco, A., Bianchi, A., Mancini, A., Milardi, D., Giampietro, A., Cimino, V., et al., 2007. Effects of ghrelin administration on endocrine and metabolic parameters in obese women with polycystic ovary syndrome. *Journal of Endocrinological Investigation* 30:948–956.
- [88] Guido, M., Romualdi, D., De Marinis, L., Porcellini, T., Giuliani, M., Costantini, B., et al., 2007. Administration of exogenous ghrelin in obese patients with polycystic ovary syndrome: effects on plasma levels of growth hormone, glucose, and insulin. *Fertility and Sterility* 88:125–130.
- [89] Tassone, F., Broglio, F., Destefanis, S., Rovere, S., Benso, A., Gottero, C., et al., 2003. Neuroendocrine and metabolic effects of acute ghrelin administration in human obesity. *The Journal of Clinical Endocrinology and Metabolism* 88:5478–5483.
- [90] Falken, Y., Hellstrom, P.M., Sanger, G.J., Dewit, O., Dukes, G., Gryback, P., et al., 2010. Actions of prolonged ghrelin infusion on gastrointestinal transit and glucose homeostasis in humans. *Neurogastroenterology and Motility: the Official Journal of the European Gastrointestinal Motility Society* 22:e192–200.

- [91] Murray, C.D., Martin, N.M., Patterson, M., Taylor, S.A., Ghatei, M.A., Kamm, M.A., et al., 2005. Ghrelin enhances gastric emptying in diabetic gastroparesis: a double blind, placebo controlled, crossover study. *Gut* 54:1693–1698.
- [92] Tack, J., Depoortere, I., Bisschops, R., Delpoortere, C., Coulie, B., Meulemans, A., et al., 2006. Influence of ghrelin on interdigestive gastrointestinal motility in humans. *Gut* 55:327–333.
- [93] Van der Ploeg, L., Laken, H., Sharma, S., Datta, R., Halem, H., Dong, J., et al., 2014. Preclinical gastrointestinal prokinetic efficacy and endocrine effects of the ghrelin mimetic RM-131. *Life Sciences* 109:20–29.
- [94] Wren, A.M., Seal, L.J., Cohen, M.A., Brynes, A.E., Frost, G.S., Murphy, K.G., et al., 2001. Ghrelin enhances appetite and increases food intake in humans. *The Journal of Clinical Endocrinology and Metabolism* 86:5992.
- [95] Holsen, L.M., Lawson, E.A., Christensen, K., Klibanski, A., Goldstein, J.M., 2014. Abnormal relationships between the neural response to high- and low-calorie foods and endogenous acylated ghrelin in women with active and weight-recovered anorexia nervosa. *Psychiatry Research* 223:94–103.
- [96] Kroemer, N.B., Krebs, L., Kobiella, A., Grimm, O., Pilhatsch, M., Bidlingmaier, M., et al., 2012. Fasting levels of ghrelin covary with the brain response to food pictures. *Addiction Biology* 8:855–862.
- [97] Malik, S., McGlone, F., Bedrossian, D., Dagher, A., 2008. Ghrelin modulates brain activity in areas that control appetitive behavior. *Cell metabolism* 7: 400–409.
- [98] Goldstone, A.P., Prechtel, C.G., Scholtz, S., Miras, A.D., Chhina, N., Durighel, G., et al., 2014. Ghrelin mimics fasting to enhance human hedonic, orbitofrontal cortex, and hippocampal responses to food. *The American Journal of Clinical Nutrition* 99:1319–1330.
- [99] Arvat, E., Di Vito, L., Broglio, F., Papotti, M., Muccioli, G., Dieguez, C., et al., 2000. Preliminary evidence that Ghrelin, the natural GH secretagogue (GHS)-receptor ligand, strongly stimulates GH secretion in humans. *Journal of Endocrinological Investigation* 23:493–495.
- [100] Peino, R., Baldelli, R., Rodriguez-Garcia, J., Rodriguez-Segade, S., Kojima, M., Kangawa, K., et al., 2000. Ghrelin-induced growth hormone secretion in humans. *European Journal of Endocrinology/European Federation of Endocrine Societies* 143:R11–R14.
- [101] Veldhuis, J.D., Reynolds, G.A., Iranmanesh, A., Bowers, C.Y., 2008. Twenty-four hour continuous ghrelin infusion augments physiologically pulsatile, nycthemeral, and entropic (feedback-regulated) modes of growth hormone secretion. *The Journal of Clinical Endocrinology and Metabolism* 93:3597–3603.
- [102] Pantel, J., Legendre, M., Cabrol, S., Hilal, L., Hajaji, Y., Morisset, S., et al., 2006. Loss of constitutive activity of the growth hormone secretagogue receptor in familial short stature. *The Journal of Clinical Investigation* 116: 760–768.
- [103] Lango Allen, H., Estrada, K., Lettre, G., Berndt, S.I., Weedon, M.N., Rivadeneira, F., et al., 2010. Hundreds of variants clustered in genomic loci and biological pathways affect human height. *Nature* 467:832–838.
- [104] Liu, B., Garcia, E.A., Korbonits, M., 2011. Genetic studies on the ghrelin, growth hormone secretagogue receptor (GHSR) and ghrelin O-acyl transferase (GOAT) genes. *Peptides* 32:2191–2207.
- [105] Lanfranco, F., Bonelli, L., Baldi, M., Me, E., Broglio, F., Ghigo, E., 2008. Acylated ghrelin inhibits spontaneous luteinizing hormone pulsatility and responsiveness to naloxone but not that to gonadotropin-releasing hormone in young men: evidence for a central inhibitory action of ghrelin on the gonadal axis. *The Journal of Clinical Endocrinology and Metabolism* 93: 3633–3639.
- [106] Patchett, A.A., Nargund, R.P., Tata, J.R., Chen, M.H., Barakat, K.J., Johnston, D.B., et al., 1995. Design and biological activities of L-163,191 (MK-0677): a potent, orally active growth hormone secretagogue. *Proceedings of the National Academy of Sciences of the United States of America* 92:7001–7005.
- [107] Popovic, V., Leal, A., Micic, D., Koppeschaar, H.P., Torres, E., Paramo, C., et al., 2000. GH-releasing hormone and GH-releasing peptide-6 for diagnostic testing in GH-deficient adults. *Lancet* 356:1137–1142.
- [108] Ueberberg, B., Unger, N., Saeger, W., Mann, K., Petersenn, S., 2009. Expression of ghrelin and its receptor in human tissues. *Hormone and Metabolic Research = Hormon- und Stoffwechselforschung = Hormones et metabolisme* 41:814–821.
- [109] Hou, Z., Miao, Y., Gao, L., Pan, H., Zhu, S., 2006. Ghrelin-containing neuron in cerebral cortex and hypothalamus linked with the DVC of brainstem in rat. *Regulatory Peptides* 134:126–131.
- [110] Palus, S., Schur, R., Akashi, Y.J., Bockmeyer, B., Datta, R., Halem, H., et al., 2011. Ghrelin and its analogues, BIM-28131 and BIM-28125, improve body weight and regulate the expression of MuRF-1 and MAFbx in a rat heart failure model. *PLoS One* 6:e26865.
- [111] Korbonits, M., Kojima, M., Kangawa, K., Grossman, A.B., 2001. Presence of ghrelin in normal and adenomatous human pituitary. *Endocrine* 14:101–104.
- [112] Korbonits, M., Bustin, S.A., Kojima, M., Jordan, S., Adams, E.F., Lowe, D.G., et al., 2001. The expression of the growth hormone secretagogue receptor ligand ghrelin in normal and abnormal human pituitary and other neuroendocrine tumors. *The Journal of Clinical Endocrinology and Metabolism* 86: 881–887.
- [113] Volante, M., Allia, E., Fulcheri, E., Cassoni, P., Ghigo, E., Muccioli, G., et al., 2003. Ghrelin in fetal thyroid and follicular tumors and cell lines: expression and effects on tumor growth. *The American Journal of Pathology* 162:645–654.
- [114] Volante, M., Fulcheri, E., Allia, E., Cerrato, M., Pucci, A., Papotti, M., 2002. Ghrelin expression in fetal, infant, and adult human lung. *The Journal of Histochemistry and Cytochemistry: Official Journal of the Histochemistry Society* 50:1013–1021.
- [115] Granata, R., Isgaard, J., Alloati, G., Ghigo, E., 2011. Cardiovascular actions of the ghrelin gene-derived peptides and growth hormone-releasing hormone. *Experimental Biology and Medicine (Maywood)* 236:505–514.
- [116] Beiras-Fernandez, A., Kreth, S., Weis, F., Ledderose, C., Pottinger, T., Dieguez, C., et al., 2010. Altered myocardial expression of ghrelin and its receptor (GHSR-1a) in patients with severe heart failure. *Peptides* 31:2222–2228.
- [117] Chuang, J.C., Sakata, I., Kohno, D., Perello, M., Osborne-Lawrence, S., Repa, J.J., et al., 2011. Ghrelin directly stimulates glucagon secretion from pancreatic alpha-cells. *Molecular Endocrinology* 25:1600–1611.
- [118] Reimer, M.K., Pacini, G., Ahren, B., 2003. Dose-dependent inhibition by ghrelin of insulin secretion in the mouse. *Endocrinology* 144:916–921.
- [119] Dezaki, K., Hosoda, H., Kakei, M., Hashiguchi, S., Watanabe, M., Kangawa, K., et al., 2004. Endogenous ghrelin in pancreatic islets restricts insulin release by attenuating Ca²⁺ signaling in beta-cells: implication in the glycemic control in rodents. *Diabetes* 53:3142–3151.
- [120] Salehi, A., Dornonville de la Cour, C., Hakanson, R., Lundquist, I., 2004. Effects of ghrelin on insulin and glucagon secretion: a study of isolated pancreatic islets and intact mice. *Regulatory Peptides* 118:143–150.
- [121] Qader, S.S., Lundquist, I., Ekelund, M., Hakanson, R., Salehi, A., 2005. Ghrelin activates neuronal constitutive nitric oxide synthase in pancreatic islet cells while inhibiting insulin release and stimulating glucagon release. *Regulatory Peptides* 128:51–56.
- [122] Dezaki, K., Sone, H., Koizumi, M., Nakata, M., Kakei, M., Nagai, H., et al., 2006. Blockade of pancreatic islet-derived ghrelin enhances insulin secretion to prevent high-fat diet-induced glucose intolerance. *Diabetes* 55:3486–3493.
- [123] Flanagan, D.E., Evans, M.L., Monsod, T.P., Rife, F., Heptulla, R.A., Tamborlane, W.V., et al., 2003. The influence of insulin on circulating ghrelin. *American Journal of Physiology Endocrinology and Metabolism* 284:E313–E316.

- [124] Tong, J., Prigeon, R.L., Davis, H.W., Bidlingmaier, M., Kahn, S.E., Cummings, D.E., et al., 2010. Ghrelin suppresses glucose-stimulated insulin secretion and deteriorates glucose tolerance in healthy humans. *Diabetes* 59: 2145–2151.
- [125] Dezaki, K., Kakei, M., Yada, T., 2007. Ghrelin uses Galphai2 and activates voltage-dependent K⁺ channels to attenuate glucose-induced Ca²⁺ signaling and insulin release in islet beta-cells: novel signal transduction of ghrelin. *Diabetes* 56:2319–2327.
- [126] Park, S., Jiang, H., Zhang, H., Smith, R.G., 2012. Modification of ghrelin receptor signaling by somatostatin receptor-5 regulates insulin release. *Proceedings of the National Academy of Sciences of the United States of America* 109:19003–19008.
- [127] Adeghate, E., Ponery, A.S., 2002. Ghrelin stimulates insulin secretion from the pancreas of normal and diabetic rats. *Journal of Neuroendocrinology* 14: 555–560.
- [128] Irako, T., Akamizu, T., Hosoda, H., Iwakura, H., Ariyasu, H., Tojo, K., et al., 2006. Ghrelin prevents development of diabetes at adult age in streptozotocin-treated newborn rats. *Diabetologia* 49:1264–1273.
- [129] Soriano-Guillen, L., Barrios, V., Martos, G., Chowen, J.A., Campos-Barros, A., Argente, J., 2004. Effect of oral glucose administration on ghrelin levels in obese children. *European Journal of Endocrinology/European Federation of Endocrine Societies* 151:119–121.
- [130] Martos-Moreno, G.A., Barrios, V., Soriano-Guillen, L., Argente, J., 2006. Relationship between adiponectin levels, acylated ghrelin levels, and short-term body mass index changes in children with diabetes mellitus type 1 at diagnosis and after insulin therapy. *European Journal of Endocrinology/European Federation of Endocrine Societies* 155:757–761.
- [131] De Smet, B., Mitselos, A., Depoortere, I., 2009. Motilin and ghrelin as prokinetic drug targets. *Pharmacology & Therapeutics* 123:207–223.
- [132] Tomasetto, C., Karam, S.M., Ribieras, S., Masson, R., Lefebvre, O., Staub, A., et al., 2000. Identification and characterization of a novel gastric peptide hormone: the motilin-related peptide. *Gastroenterology* 119:395–405.
- [133] Ang, D., Nicolai, H., Vos, R., Mimidis, K., Akyuz, F., Kindt, S., et al., 2009. Influence of ghrelin on the gastric accommodation reflex and on meal-induced satiety in man. *Neurogastroenterology and Motility: The Official Journal of the European Gastrointestinal Motility Society* 21:528–533 e528–529.
- [134] Levin, F., Edholm, T., Schmidt, P.T., Gryback, P., Jacobsson, H., Degerblad, M., et al., 2006. Ghrelin stimulates gastric emptying and hunger in normal-weight humans. *The Journal of Clinical Endocrinology and Metabolism* 91:3296–3302.
- [135] Avau, B., Carbone, F., Tack, J., Depoortere, I., 2013. Ghrelin signaling in the gut, its physiological properties, and therapeutic potential. *Neurogastroenterology and Motility: The Official Journal of the European Gastrointestinal Motility Society* 25:720–732.
- [136] Lall, S., Tung, L.Y., Ohlsson, C., Jansson, J.O., Dickson, S.L., 2001. Growth hormone (GH)-independent stimulation of adiposity by GH secretagogues. *Biochemical and Biophysical Research Communications* 280:132–138.
- [137] Chen, H.Y., Trumbauer, M.E., Chen, A.S., Weingarth, D.T., Adams, J.R., Frazier, E.G., et al., 2004. Orexigenic action of peripheral ghrelin is mediated by neuropeptide Y and agouti-related protein. *Endocrinology* 145:2607–2612.
- [138] Wang, Q., Liu, C., Uchida, A., Chuang, J.C., Walker, A., Liu, T., et al., 2014. Arcuate AgRP neurons mediate orexigenic and glucoregulatory actions of ghrelin. *Molecular Metabolism* 3:64–72.
- [139] Verhulst, P.J., De Smet, B., Saels, I., Thijs, T., Ver Donck, L., Moechars, D., et al., 2008. Role of ghrelin in the relationship between hyperphagia and accelerated gastric emptying in diabetic mice. *Gastroenterology* 135:1267–1276.
- [140] Sakkou, M., Wiedmer, P., Anlag, K., Hamm, A., Seuntjens, E., Ettwiller, L., et al., 2007. A role for brain-specific homeobox factor Bsx in the control of hyperphagia and locomotory behavior. *Cell Metabolism* 5:450–463.
- [141] Theander-Carrillo, C., Wiedmer, P., Cettour-Rose, P., Nogueiras, R., Perez-Tilve, D., Pfluger, P., et al., 2006. Ghrelin action in the brain controls adipocyte metabolism. *The Journal of Clinical Investigation* 116:1983–1993.
- [142] Perez-Tilve, D., Hofmann, S.M., Basford, J., Nogueiras, R., Pfluger, P.T., Patterson, J.T., et al., 2010. Melanocortin signaling in the CNS directly regulates circulating cholesterol. *Nature Neuroscience* 13:877–882.
- [143] Perez-Tilve, D., Heppner, K., Kirchner, H., Lockie, S.H., Woods, S.C., Smiley, D.L., et al., 2011. Ghrelin-induced adiposity is independent of orexigenic effects. *FASEB Journal: Official publication of the Federation of American Societies for Experimental Biology* 25:2814–2822.
- [144] McFarlane, M.R., Brown, M.S., Goldstein, J.L., Zhao, T.J., 2014. Induced ablation of ghrelin cells in adult mice does not decrease food intake, body weight, or response to high-fat diet. *Cell Metabolism* 20:54–60.
- [145] Briggs, D.I., Enriori, P.J., Lemus, M.B., Cowley, M.A., Andrews, Z.B., 2010. Diet-induced obesity causes ghrelin resistance in arcuate NPY/AgRP neurons. *Endocrinology* 151:4745–4755.
- [146] Briggs, D.I., Lockie, S.H., Wu, Q., Lemus, M.B., Stark, R., Andrews, Z.B., 2013. Calorie-restricted weight loss reverses high-fat diet-induced ghrelin resistance, which contributes to rebound weight gain in a ghrelin-dependent manner. *Endocrinology* 154:709–717.
- [147] Shrestha, Y.B., Wickwire, K., Giraudo, S., 2009. Effect of reducing hypothalamic ghrelin receptor gene expression on energy balance. *Peptides* 30: 1336–1341.
- [148] De Smet, B., Depoortere, I., Moechars, D., Swennen, Q., Moreaux, B., Cryns, K., et al., 2006. Energy homeostasis and gastric emptying in ghrelin knockout mice. *The Journal of Pharmacology and Experimental Therapeutics* 316:431–439.
- [149] Sun, Y., Ahmed, S., Smith, R.G., 2003. Deletion of ghrelin impairs neither growth nor appetite. *Molecular and Cellular Biology* 23:7973–7981.
- [150] Wortley, K.E., Anderson, K.D., Garcia, K., Murray, J.D., Malinova, L., Liu, R., et al., 2004. Genetic deletion of ghrelin does not decrease food intake but influences metabolic fuel preference. *Proceedings of the National Academy of Sciences of the United States of America* 101:8227–8232.
- [151] Zigman, J.M., Nakano, Y., Coppari, R., Balthasar, N., Marcus, J.N., Lee, C.E., et al., 2005. Mice lacking ghrelin receptors resist the development of diet-induced obesity. *The Journal of Clinical Investigation* 115:3564–3572.
- [152] Longo, K.A., Charoenthongtrakul, S., Giuliana, D.J., Govek, E.K., McDonagh, T., Qi, Y., et al., 2008. Improved insulin sensitivity and metabolic flexibility in ghrelin receptor knockout mice. *Regulatory Peptides* 150:55–61.
- [153] Pfluger, P.T., Kirchner, H., Gunnell, S., Schrott, B., Perez-Tilve, D., Fu, S., et al., 2008. Simultaneous deletion of ghrelin and its receptor increases motor activity and energy expenditure. *American Journal of Physiology Gastrointestinal and Liver Physiology* 294:G610–G618.
- [154] Yamada, G., Ariyasu, H., Iwakura, H., Hosoda, H., Akamizu, T., Nakao, K., et al., 2010. Generation of transgenic mice overexpressing a ghrelin analog. *Endocrinology* 151:5935–5940.
- [155] Sun, Y., Butte, N.F., Garcia, J.M., Smith, R.G., 2008. Characterization of adult ghrelin and ghrelin receptor knockout mice under positive and negative energy balance. *Endocrinology* 149:843–850.
- [156] Szentirmai, E., Kapas, L., Sun, Y., Smith, R.G., Krueger, J.M., 2009. The preproghrelin gene is required for the normal integration of thermoregulation and sleep in mice. *Proceedings of the National Academy of Sciences of the United States of America* 106:14069–14074.
- [157] Sun, Y., Asnicar, M., Saha, P.K., Chan, L., Smith, R.G., 2006. Ablation of ghrelin improves the diabetic but not obese phenotype of ob/ob mice. *Cell Metabolism* 3:379–386.
- [158] Sun, Y., Wang, P., Zheng, H., Smith, R.G., 2004. Ghrelin stimulation of growth hormone release and appetite is mediated through the growth hormone secretagogue receptor. *Proceedings of the National Academy of Sciences of the United States of America* 101:4679–4684.
- [159] Mani, B.K., Walker, A.K., Lopez Soto, E.J., Raingo, J., Lee, C.E., Perello, M., et al., 2014. Neuroanatomical characterization of a growth hormone

- secretagogue receptor-green fluorescent protein reporter mouse. *The Journal of Comparative Neurology* 522:3644–3666.
- [160] Zigman, J.M., Jones, J.E., Lee, C.E., Saper, C.B., Elmquist, J.K., 2006. Expression of ghrelin receptor mRNA in the rat and the mouse brain. *The Journal of Comparative Neurology* 494:528–548.
- [161] Shuto, Y., Shibasaki, T., Otagiri, A., Kuriyama, H., Ohata, H., Tamura, H., et al., 2002. Hypothalamic growth hormone secretagogue receptor regulates growth hormone secretion, feeding, and adiposity. *The Journal of Clinical Investigation* 109:1429–1436.
- [162] Rizza, R.A., Mandarino, L.J., Gerich, J.E., 1982. Effects of growth hormone on insulin action in man. Mechanisms of insulin resistance, impaired suppression of glucose production, and impaired stimulation of glucose utilization. *Diabetes* 31:663–669.
- [163] Kern, A., Albarran-Zeckler, R., Walsh, H.E., Smith, R.G., 2012. Apo-ghrelin receptor forms heteromers with DRD2 in hypothalamic neurons and is essential for anorexigenic effects of DRD2 agonism. *Neuron* 73:317–332.
- [164] Muller, T.D., Muller, A., Yi, C.X., Habegger, K.M., Meyer, C.W., Gaylann, B.D., et al., 2013. The orphan receptor Gpr83 regulates systemic energy metabolism via ghrelin-dependent and ghrelin-independent mechanisms. *Nature Communications* 4:1968.
- [165] Damian, M., Marie, J., Leyris, J.P., Fehrentz, J.A., Verdier, P., Martinez, J., et al., 2012. High constitutive activity is an intrinsic feature of ghrelin receptor protein: a study with a functional monomeric GHS-R1a receptor reconstituted in lipid discs. *The Journal of Biological Chemistry* 287:3630–3641.
- [166] Atalayer, D., Gibson, C., Konopacka, A., Geliebter, A., 2013. Ghrelin and eating disorders. *Progress in Neuro-Psychopharmacology & Biological Psychiatry* 40:70–82.
- [167] Heppner, K.M., Muller, T.D., Kirchner, H., Perez-Tilve, D., Pfluger, P.T., Tschop, M.H., et al., 2013. The role of ghrelin-octanoyl-acyl-transferase in thermoregulation. *Journal of Endocrinological Investigation* 36:180–184.
- [168] Goldstein, J.L., Zhao, T.J., Li, R.L., Sherbet, D.P., Liang, G., Brown, M.S., 2011. Surviving starvation: essential role of the ghrelin-growth hormone axis. *Cold Spring Harbor Symposia on Quantitative Biology* 76:121–127.
- [169] Li, R.L., Sherbet, D.P., Elsbernd, B.L., Goldstein, J.L., Brown, M.S., Zhao, T.J., 2012. Profound hypoglycemia in starved, ghrelin-deficient mice is caused by decreased gluconeogenesis and reversed by lactate or fatty acids. *The Journal of Biological Chemistry* 287:17942–17950.
- [170] Zhang, Y., Fang, F., Goldstein, J.L., Brown, M.S., Zhao, T.J., 2015. Reduced autophagy in livers of fasted, fat-depleted, ghrelin-deficient mice: Reversal by growth hormone. *Proceedings of the National Academy of Sciences of the United States of America* 112:1226–1231.
- [171] Frayling, T.M., Timpson, N.J., Weedon, M.N., Zeggini, E., Freathy, R.M., Lindgren, C.M., et al., 2007. A common variant in the FTO gene is associated with body mass index and predisposes to childhood and adult obesity. *Science* 316:889–894.
- [172] Dina, C., Meyre, D., Gallina, S., Durand, E., Korner, A., Jacobson, P., et al., 2007. Variation in FTO contributes to childhood obesity and severe adult obesity. *Nature Genetics* 39:724–726.
- [173] Hardy, R., Wills, A.K., Wong, A., Elks, C.E., Wareham, N.J., Loos, R.J., et al., 2010. Life course variations in the associations between FTO and MC4R gene variants and body size. *Human Molecular Genetics* 19:545–552.
- [174] Li, H., Kilpelainen, T.O., Liu, C., Zhu, J., Liu, Y., Hu, C., et al., 2012. Association of genetic variation in FTO with risk of obesity and type 2 diabetes with data from 96,551 East and South Asians. *Diabetologia* 55:981–995.
- [175] Lauria, F., Siani, A., Bammann, K., Foraita, R., Huybrechts, I., Iacoviello, L., et al., 2012. Prospective analysis of the association of a common variant of FTO (rs9939609) with adiposity in children: results of the IDEFICS study. *PLoS One* 7:e48876.
- [176] Hinney, A., Nguyen, T.T., Scherag, A., Friedel, S., Bronner, G., Muller, T.D., et al., 2007. Genome wide association (GWA) study for early onset extreme obesity supports the role of fat mass and obesity associated gene (FTO) variants. *PLoS One* 2:e1361.
- [177] Cecil, J.E., Tavendale, R., Watt, P., Hetherington, M.M., Palmer, C.N., 2008. An obesity-associated FTO gene variant and increased energy intake in children. *The New England Journal of Medicine* 359:2558–2566.
- [178] Speakman, J.R., Rance, K.A., Johnstone, A.M., 2008. Polymorphisms of the FTO gene are associated with variation in energy intake, but not energy expenditure. *Obesity (Silver Spring)* 16:1961–1965.
- [179] Wardle, J., Llewellyn, C., Sanderson, S., Plomin, R., 2009. The FTO gene and measured food intake in children. *International Journal of Obesity (London)* 33:42–45.
- [180] Timpson, N.J., Emmett, P.M., Frayling, T.M., Rogers, I., Hattersley, A.T., McCarthy, M.I., et al., 2008. The fat mass- and obesity-associated locus and dietary intake in children. *The American Journal of Clinical Nutrition* 88:971–978.
- [181] Tanofsky-Kraff, M., Han, J.C., Anandalingam, K., Shomaker, L.B., Columbo, K.M., Wolkoff, L.E., et al., 2009. The FTO gene rs9939609 obesity-risk allele and loss of control over eating. *The American Journal of Clinical Nutrition* 90:1483–1488.
- [182] Wardle, J., Carnell, S., Haworth, C.M., Farooqi, I.S., O’Rahilly, S., Plomin, R., 2008. Obesity associated genetic variation in FTO is associated with diminished satiety. *The Journal of Clinical Endocrinology and Metabolism* 93:3640–3643.
- [183] den Hoed, M., Westerterp-Plantenga, M.S., Bouwman, F.G., Mariman, E.C., Westerterp, K.R., 2009. Postprandial responses in hunger and satiety are associated with the rs9939609 single nucleotide polymorphism in FTO. *The American Journal of Clinical Nutrition* 90:1426–1432.
- [184] Karra, E., O’Daly, O.G., Choudhury, A.I., Youssef, A., Millership, S., Neary, M.T., et al., 2013. A link between FTO, ghrelin, and impaired brain food-cue responsivity. *The Journal of Clinical Investigation* 123:3539–3551.
- [185] Meyer, K.D., Saletore, Y., Zumbo, P., Elemento, O., Mason, C.E., Jaffrey, S.R., 2012. Comprehensive analysis of mRNA methylation reveals enrichment in 3’ UTRs and near stop codons. *Cell* 149:1635–1646.
- [186] Berulava, T., Horsthemke, B., 2010. The obesity-associated SNPs in intron 1 of the FTO gene affect primary transcript levels. *European Journal of Human Genetics* 18:1054–1056.
- [187] Hess, M.E., Hess, S., Meyer, K.D., Verhagen, L.A., Koch, L., Bronneke, H.S., et al., 2013. The fat mass and obesity associated gene (Fto) regulates activity of the dopaminergic midbrain circuitry. *Nature Neuroscience* 16:1042–1048.
- [188] Diano, S., Farr, S.A., Benoit, S.C., McNay, E.C., da Silva, I., Horvath, B., et al., 2006. Ghrelin controls hippocampal spine synapse density and memory performance. *Nature Neuroscience* 9:381–388.
- [189] Kanoski, S.E., Fortin, S.M., Ricks, K.M., Grill, H.J., 2013. Ghrelin signaling in the ventral hippocampus stimulates learned and motivational aspects of feeding via PI3K-Akt signaling. *Biological Psychiatry* 73:915–923.
- [190] Kanoski, S.E., Hayes, M.R., Greenwald, H.S., Fortin, S.M., Gianessi, C.A., Gilbert, J.R., et al., 2011. Hippocampal leptin signaling reduces food intake and modulates food-related memory processing. *Neuropsychopharmacology: Official Publication of the American College of Neuropsychopharmacology* 36:1859–1870.
- [191] Chuang, J.C., Zigman, J.M., 2010. Ghrelin’s roles in stress, mood, and anxiety regulation. *International Journal of Peptides* 2010 pii: 460549.
- [192] Spencer, S.J., Emmerzaal, T.L., Kozicz, T., Andrews, Z.B., 2014. Ghrelin’s role in the hypothalamic-pituitary-adrenal Axis stress Response: Implications for mood disorders. *Biological Psychiatry* pii: S0006-3223(14)00808-7.
- [193] Barim, A.O., Aydin, S., Colak, R., Dag, E., Deniz, O., Sahin, I., 2009. Ghrelin, paraoxonase and arylesterase levels in depressive patients before and after citalopram treatment. *Clinical Biochemistry* 42:1076–1081.
- [194] Kurt, E., Guler, O., Serteser, M., Cansel, N., Ozbulut, O., Altinbas, K., et al., 2007. The effects of electroconvulsive therapy on ghrelin, leptin and

- cholesterol levels in patients with mood disorders. *Neuroscience Letters* 426:49–53.
- [195] Dixit, V.D., Yang, H., Sun, Y., Weeraratna, A.T., Youm, Y.H., Smith, R.G., et al., 2007. Ghrelin promotes thymopoiesis during aging. *The Journal of Clinical Investigation* 117:2778–2790.
- [196] Szentirmai, E., Kapas, L., Krueger, J.M., 2007. Ghrelin microinjection into forebrain sites induces wakefulness and feeding in rats. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology* 292:R575–R585.
- [197] Szentirmai, E., Kapas, L., Sun, Y., Smith, R.G., Krueger, J.M., 2007. Spontaneous sleep and homeostatic sleep regulation in ghrelin knockout mice. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology* 293:R510–R517.
- [198] Andrews, Z.B., Erion, D., Beiler, R., Liu, Z.W., Abizaid, A., Zigman, J., et al., 2009. Ghrelin promotes and protects nigrostriatal dopamine function via a UCP2-dependent mitochondrial mechanism. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience* 29:14057–14065.
- [199] Moon, M., Kim, H.G., Hwang, L., Seo, J.H., Kim, S., Hwang, S., et al., 2009. Neuroprotective effect of ghrelin in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mouse model of Parkinson's disease by blocking microglial activation. *Neurotoxicity Research* 15:332–347.
- [200] Walker, A.K., Rivera, P.D., Wang, Q., Chuang, J.C., Tran, S., Osborne-Lawrence, S., et al., 2014. The P7C3 class of neuroprotective compounds exerts antidepressant efficacy in mice by increasing hippocampal neurogenesis. *Molecular Psychiatry* 20:500–508.
- [201] Currie, P.J., Khelemsky, R., Rigsbee, E.M., Dono, L.M., Coiro, C.D., Chapman, C.D., et al., 2012. Ghrelin is an orexigenic peptide and elicits anxiety-like behaviors following administration into discrete regions of the hypothalamus. *Behavioural Brain Research* 226:96–105.
- [202] Olszewski, P.K., Grace, M.K., Billington, C.J., Levine, A.S., 2003. Hypothalamic paraventricular injections of ghrelin: effect on feeding and c-Fos immunoreactivity. *Peptides* 24:919–923.
- [203] van der Lely, A.J., Tschöp, M., Heiman, M.L., Ghigo, E., 2004. Biological, physiological, pathophysiological, and pharmacological aspects of ghrelin. *Endocrine Reviews* 25:426–457.
- [204] Kola, B., Farkas, I., Christ-Crain, M., Wittmann, G., Lolli, F., Amin, F., et al., 2008. The orexigenic effect of ghrelin is mediated through central activation of the endogenous cannabinoid system. *PLoS One* 3:e1797.
- [205] Anderson, K.A., Ribar, T.J., Lin, F., Noeldner, P.K., Green, M.F., Muehlbauer, M.J., et al., 2008. Hypothalamic CaMKK2 contributes to the regulation of energy balance. *Cell Metabolism* 7:377–388.
- [206] Kola, B., Hubina, E., Tucci, S.A., Kirkham, T.C., Garcia, E.A., Mitchell, S.E., et al., 2005. Cannabinoids and ghrelin have both central and peripheral metabolic and cardiac effects via AMP-activated protein kinase. *The Journal of Biological Chemistry* 280:25196–25201.
- [207] Lopez, M., Lage, R., Saha, A.K., Perez-Tilve, D., Vazquez, M.J., Varela, L., et al., 2008. Hypothalamic fatty acid metabolism mediates the orexigenic action of ghrelin. *Cell Metabolism* 7:389–399.
- [208] Dietrich, M.O., Antunes, C., Geliang, G., Liu, Z.W., Borok, E., Nie, Y., et al., 2010. AgRP neurons mediate Sirt1's action on the melanocortin system and energy balance: roles for Sirt1 in neuronal firing and synaptic plasticity. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience* 30:11815–11825.
- [209] Velasquez, D.A., Martinez, G., Romero, A., Vazquez, M.J., Boit, K.D., Dopeso-Reyes, I.G., et al., 2011. The central Sirtuin 1/p53 pathway is essential for the orexigenic action of ghrelin. *Diabetes* 60:1177–1185.
- [210] Andrews, Z.B., Liu, Z.W., Wallingford, N., Erion, D.M., Borok, E., Friedman, J.M., et al., 2008. UCP2 mediates ghrelin's action on NPY/AgRP neurons by lowering free radicals. *Nature* 454:846–851.
- [211] Lage, R., Vazquez, M.J., Varela, L., Saha, A.K., Vidal-Puig, A., Nogueiras, R., et al., 2010. Ghrelin effects on neuropeptides in the rat hypothalamus depend on fatty acid metabolism actions on BSX but not on gender. *FASEB Journal: Official Publication of the Federation of American Societies for Experimental Biology* 24:2670–2679.
- [212] Kwon Jeong, J., Dae Kim, J., Diano, S., 2013. Ghrelin regulates hypothalamic prolyl carboxypeptidase expression in mice. *Molecular Metabolism* 2:23–30.
- [213] Martins, L., Fernandez-Mallo, D., Novelle, M.G., Vazquez, M.J., Tena-Sempere, M., Nogueiras, R., et al., 2012. Hypothalamic mTOR signaling mediates the orexigenic action of ghrelin. *PLoS One* 7:e46923.
- [214] Faulconbridge, L.F., Cummings, D.E., Kaplan, J.M., Grill, H.J., 2003. Hyperphagic effects of brainstem ghrelin administration. *Diabetes* 52:2260–2265.
- [215] Faulconbridge, L.F., Grill, H.J., Kaplan, J.M., 2005. Distinct forebrain and caudal brainstem contributions to the neuropeptide Y mediation of ghrelin hyperphagia. *Diabetes* 54:1985–1993.
- [216] Faulconbridge, L.F., Grill, H.J., Kaplan, J.M., Daniels, D., 2008. Caudal brainstem delivery of ghrelin induces fos expression in the nucleus of the solitary tract, but not in the arcuate or paraventricular nuclei of the hypothalamus. *Brain Research* 1218:151–157.
- [217] Alvarez-Crespo, M., Skibicka, K.P., Farkas, I., Molnar, C.S., Egecioglu, E., Hrabovszky, E., et al., 2012. The amygdala as a neurobiological target for ghrelin in rats: neuroanatomical, electrophysiological and behavioral evidence. *PLoS One* 7:e46321.
- [218] Olszewski, P.K., Li, D., Grace, M.K., Billington, C.J., Kotz, C.M., Levine, A.S., 2003. Neural basis of orexigenic effects of ghrelin acting within lateral hypothalamus. *Peptides* 24:597–602.
- [219] Scott, M.M., Perello, M., Chuang, J.C., Sakata, I., Gautron, L., Lee, C.E., et al., 2012. Hindbrain ghrelin receptor signaling is sufficient to maintain fasting glucose. *PLoS One* 7:e44089.
- [220] Bailey, A.R., von Engelhardt, N., Leng, G., Smith, R.G., Dickson, S.L., 2000. Growth hormone secretagogue activation of the arcuate nucleus and brainstem occurs via a non-noradrenergic pathway. *Journal of Neuroendocrinology* 12:191–197.
- [221] Lawrence, C.B., Snape, A.C., Baudoin, F.M., Luckman, S.M., 2002. Acute central ghrelin and GH secretagogues induce feeding and activate brain appetite centers. *Endocrinology* 143:155–162.
- [222] Date, Y., Murakami, N., Toshinai, K., Matsukura, S., Nijijima, A., Matsuo, H., et al., 2002. The role of the gastric afferent vagal nerve in ghrelin-induced feeding and growth hormone secretion in rats. *Gastroenterology* 123:1120–1128.
- [223] Arnold, M., Mura, A., Langhans, W., Geary, N., 2006. Gut vagal afferents are not necessary for the eating-stimulatory effect of intraperitoneally injected ghrelin in the rat. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience* 26:11052–11060.
- [224] Dornonville de la Cour, C., Lindqvist, A., Egecioglu, E., Tung, Y.C., Surve, V., Ohlsson, C., et al., 2005. Ghrelin treatment reverses the reduction in weight gain and body fat in gastrectomized mice. *Gut* 54:907–913.
- [225] Adachi, S., Takiguchi, S., Okada, K., Yamamoto, K., Yamasaki, M., Miyata, H., et al., 2010. Effects of ghrelin administration after total gastrectomy: a prospective, randomized, placebo-controlled phase II study. *Gastroenterology* 138:1312–1320.
- [226] Dickson, S.L., Egecioglu, E., Landgren, S., Skibicka, K.P., Engel, J.A., Jerlhag, E., 2011. The role of the central ghrelin system in reward from food and chemical drugs. *Molecular and Cellular Endocrinology* 340:80–87.
- [227] Hansson, C., Shirazi, R.H., Naslund, J., Vogel, H., Neuber, C., Holm, G., et al., 2012. Ghrelin influences novelty seeking behavior in rodents and men. *PLoS One* 7:e50409.
- [228] Jerlhag, E., Egecioglu, E., Landgren, S., Salome, N., Heilig, M., Moechars, D., et al., 2009. Requirement of central ghrelin signaling for alcohol reward. *Proceedings of the National Academy of Sciences of the United States of America* 106:11318–11323.
- [229] Jerlhag, E., Janson, A.C., Waters, S., Engel, J.A., 2012. Concomitant release of ventral tegmental acetylcholine and accumbal dopamine by ghrelin in rats. *PLoS One* 7:e49557.

- [230] Perello, M., Sakata, I., Birnbaum, S., Chuang, J.C., Osborne-Lawrence, S., Rovinsky, S.A., et al., 2010. Ghrelin increases the rewarding value of high-fat diet in an orexin-dependent manner. *Biological Psychiatry* 67:880–886.
- [231] Skibicka, K.P., Hansson, C., Alvarez-Crespo, M., Friberg, P.A., Dickson, S.L., 2011. Ghrelin directly targets the ventral tegmental area to increase food motivation. *Neuroscience* 180:129–137.
- [232] Skibicka, K.P., Shirazi, R.H., Hansson, C., Dickson, S.L., 2012. Ghrelin interacts with neuropeptide Y Y1 and opioid receptors to increase food reward. *Endocrinology* 153:1194–1205.
- [233] Jerlhag, E., Engel, J.A., 2011. Ghrelin receptor antagonism attenuates nicotine-induced locomotor stimulation, accumbal dopamine release and conditioned place preference in mice. *Drug and Alcohol Dependence* 117: 126–131.
- [234] Wellman, P.J., Clifford, P.S., Rodriguez, J., Hughes, S., Eitan, S., Brunel, L., et al., 2011. Pharmacologic antagonism of ghrelin receptors attenuates development of nicotine induced locomotor sensitization in rats. *Regulatory Peptides* 172:77–80.
- [235] Jerlhag, E., Egecioglu, E., Dickson, S.L., Engel, J.A., 2010. Ghrelin receptor antagonism attenuates cocaine- and amphetamine-induced locomotor stimulation, accumbal dopamine release, and conditioned place preference. *Psychopharmacology* 211:415–422.
- [236] Kawahara, Y., Kaneko, F., Yamada, M., Kishikawa, Y., Kawahara, H., Nishi, A., 2013. Food reward-sensitive interaction of ghrelin and opioid receptor pathways in mesolimbic dopamine system. *Neuropharmacology* 67:395–402.
- [237] Naleid, A.M., Grace, M.K., Cummings, D.E., Levine, A.S., 2005. Ghrelin induces feeding in the mesolimbic reward pathway between the ventral tegmental area and the nucleus accumbens. *Peptides* 26:2274–2279.
- [238] Quarta, D., Di Francesco, C., Melotto, S., Mangiarini, L., Heidbreder, C., Hedou, G., 2009. Systemic administration of ghrelin increases extracellular dopamine in the shell but not the core subdivision of the nucleus accumbens. *Neurochemistry International* 54:89–94.
- [239] Abizaid, A., Liu, Z.W., Andrews, Z.B., Shanabrough, M., Borok, E., Elsworth, J.D., et al., 2006. Ghrelin modulates the activity and synaptic input organization of midbrain dopamine neurons while promoting appetite. *The Journal of Clinical Investigation* 116:3229–3239.
- [240] Berridge, K.C., Robinson, T.E., 1998. What is the role of dopamine in reward: hedonic impact, reward learning, or incentive salience? *Brain Research Brain Research Reviews* 28:309–369.
- [241] Salamone, J.D., Correa, M., Mingote, S., Weber, S.M., 2003. Nucleus accumbens dopamine and the regulation of effort in food-seeking behavior: implications for studies of natural motivation, psychiatry, and drug abuse. *The Journal of Pharmacology and Experimental Therapeutics* 305:1–8.
- [242] Blum, I.D., Patterson, Z., Khazall, R., Lamont, E.W., Sleeman, M.W., Horvath, T.L., et al., 2009. Reduced anticipatory locomotor responses to scheduled meals in ghrelin receptor deficient mice. *Neuroscience* 164:351–359.
- [243] Verhagen, L.A., Egecioglu, E., Luijendijk, M.C., Hillebrand, J.J., Adan, R.A., Dickson, S.L., 2011. Acute and chronic suppression of the central ghrelin signaling system reveals a role in food anticipatory activity. *European Neuropsychopharmacology: The Journal of the European College of Neuropsychopharmacology* 21:384–392.
- [244] Carlini, V.P., Monzon, M.E., Varas, M.M., Cragnolini, A.B., Schioth, H.B., Scimonelli, T.N., et al., 2002. Ghrelin increases anxiety-like behavior and memory retention in rats. *Biochemical and Biophysical Research Communications* 299:739–743.
- [245] Skibicka, K.P., Shirazi, R.H., Rabasa-Papio, C., Alvarez-Crespo, M., Neuber, C., Vogel, H., et al., 2013. Divergent circuitry underlying food reward and intake effects of ghrelin: dopaminergic VTA-accumbens projection mediates ghrelin's effect on food reward but not food intake. *Neuropharmacology* 73C:274–283.
- [246] Romero-Pico, A., Novelle, M.G., Folgueira, C., Lopez, M., Nogueiras, R., Dieguez, C., 2013. Central manipulation of dopamine receptors attenuates the orexigenic action of ghrelin. *Psychopharmacology* 229:275–283.
- [247] Romero-Pico, A., Vazquez, M.J., Gonzalez-Touceda, D., Folgueira, C., Skibicka, K.P., Alvarez-Crespo, M., et al., 2013. Hypothalamic kappa-opioid receptor modulates the orexigenic effect of ghrelin. *Neuropsychopharmacology: Official Publication of the American College of Neuropsychopharmacology* 38:1296–1307.
- [248] Ferrini, F., Salio, C., Lossi, L., Merighi, A., 2009. Ghrelin in central neurons. *Current Neuropharmacology* 7:37–49.
- [249] Liu, Y., Wang, P.S., Xie, D., Liu, K., Chen, L., 2006. Ghrelin reduces injury of hippocampal neurons in a rat model of cerebral ischemia/reperfusion. *The Chinese Journal of Physiology* 49:244–250.
- [250] Horvath, T.L., Abizaid, A., Dietrich, M.O., Li, Y., Takahashi, J.S., Bass, J., 2012. Ghrelin-immunopositive hypothalamic neurons tie the circadian clock and visual system to the lateral hypothalamic arousal center. *Molecular Metabolism* 1:79–85.
- [251] Frecka, J.M., Mattes, R.D., 2008. Possible entrainment of ghrelin to habitual meal patterns in humans. *American Journal of Physiology Gastrointestinal and Liver Physiology* 294:G699–G707.
- [252] Monteleone, P., Bencivenga, R., Longobardi, N., Serritella, C., Maj, M., 2003. Differential responses of circulating ghrelin to high-fat or high-carbohydrate meal in healthy women. *The Journal of Clinical Endocrinology and Metabolism* 88:5510–5514.
- [253] Zhao, T.J., Sakata, I., Li, R.L., Liang, G., Richardson, J.A., Brown, M.S., et al., 2010. Ghrelin secretion stimulated by {beta}1-adrenergic receptors in cultured ghrelinoma cells and in fasted mice. *Proceedings of the National Academy of Sciences of the United States of America* 107:15868–15873.
- [254] Arosio, M., Ronchi, C.L., Beck-Peccoz, P., Gebbia, C., Giavoli, C., Cappiello, V., et al., 2004. Effects of modified sham feeding on ghrelin levels in healthy human subjects. *The Journal of Clinical Endocrinology and Metabolism* 89:5101–5104.
- [255] Callahan, H.S., Cummings, D.E., Pepe, M.S., Breen, P.A., Matthys, C.C., Weigle, D.S., 2004. Postprandial suppression of plasma ghrelin level is proportional to ingested caloric load but does not predict intermeal interval in humans. *The Journal of Clinical Endocrinology and Metabolism* 89:1319–1324.
- [256] Tschöp, M., Weyer, C., Tataranni, P.A., Devanarayan, V., Ravussin, E., Heiman, M.L., 2001. Circulating ghrelin levels are decreased in human obesity. *Diabetes* 50:707–709.
- [257] Cummings, D.E., Weigle, D.S., Frayo, R.S., Breen, P.A., Ma, M.K., Dellinger, E.P., et al., 2002. Plasma ghrelin levels after diet-induced weight loss or gastric bypass surgery. *The New England Journal of Medicine* 346: 1623–1630.
- [258] Müller, T.D., Perez-Tilve, D., Tong, J., Pfluger, P.T., Tschöp, M.H., 2010. Ghrelin and its potential in the treatment of eating/wasting disorders and cachexia. *Journal of Cachexia, Sarcopenia and Muscle* 1:159–167.
- [259] Shiiya, T., Nakazato, M., Mizuta, M., Date, Y., Mondal, M.S., Tanaka, M., et al., 2002. Plasma ghrelin levels in lean and obese humans and the effect of glucose on ghrelin secretion. *The Journal of Clinical Endocrinology and Metabolism* 87:240–244.
- [260] Geliebter, A., Gluck, M.E., Hashim, S.A., 2005. Plasma ghrelin concentrations are lower in binge-eating disorder. *The Journal of Nutrition* 135:1326–1330.
- [261] Kamiji, M.M., Inui, A., 2008. The role of ghrelin and ghrelin analogues in wasting disease. *Current Opinion in Clinical Nutrition and Metabolic Care* 11: 443–451.
- [262] Korbonits, M., Blaine, D., Elia, M., Powell-Tuck, J., 2007. Metabolic and hormonal changes during the refeeding period of prolonged fasting. *European Journal of Endocrinology/European Federation of Endocrine Societies* 157: 157–166.

- [263] Korbonits, M., Blaine, D., Elia, M., Powell-Tuck, J., 2005. Refeeding David Blaine—studies after a 44-day fast. *The New England Journal of Medicine* 353:2306–2307.
- [264] Espelund, U., Hansen, T.K., Hojlund, K., Beck-Nielsen, H., Clausen, J.T., Hansen, B.S., et al., 2005. Fasting unmasks a strong inverse association between ghrelin and cortisol in serum: studies in obese and normal-weight subjects. *The Journal of Clinical Endocrinology and Metabolism* 90:741–746.
- [265] Williams, D.L., Grill, H.J., Cummings, D.E., Kaplan, J.M., 2003. Vagotomy dissociates short- and long-term controls of circulating ghrelin. *Endocrinology* 144:5184–5187.
- [266] Cummings, D.E., Clement, K., Purnell, J.Q., Vaisse, C., Foster, K.E., Frayo, R.S., et al., 2002. Elevated plasma ghrelin levels in Prader Willi syndrome. *Nature Medicine* 8:643–644.
- [267] DelParigi, A., Tschop, M., Heiman, M.L., Salbe, A.D., Vozarova, B., Sell, S.M., et al., 2002. High circulating ghrelin: a potential cause for hyperphagia and obesity in prader-willii syndrome. *The Journal of Clinical Endocrinology and Metabolism* 87:5461–5464.
- [268] Malandrino, N., Miceli, A., Leggio, L., Mingrone, G., Capristo, E., 2014. High ghrelin levels in post-treatment euthyroid patients with Hashimoto's thyroiditis: a case-control preliminary study. *Experimental and Clinical Endocrinology & Diabetes: Official Journal, German Society of Endocrinology [and] German Diabetes Association* 122:540–543.
- [269] Buscher, A.K., Cetiner, M., Buscher, R., Wingen, A.M., Hauffa, B.P., Hoyer, P.F., 2012. Obesity in patients with Bardet-Biedl syndrome: influence of appetite-regulating hormones. *Pediatric Nephrology* 27:2065–2071.
- [270] Giordano, R., Picu, A., Pagotto, U., De lasio, R., Bonelli, L., Prodam, F., et al., 2005. The negative association between total ghrelin levels, body mass and insulin secretion is lost in hypercortisolemic patients with Cushing's disease. *European Journal of Endocrinology/European Federation of Endocrine Societies* 153:535–543.
- [271] Libe, R., Morpurgo, P.S., Cappiello, V., Maffini, A., Bondioni, S., Locatelli, M., et al., 2005. Ghrelin and adiponectin in patients with Cushing's disease before and after successful transphenoidal surgery. *Clinical Endocrinology* 62:30–36.
- [272] Koutkia, P., Meininger, G., Canavan, B., Brey, J., Grinspoon, S., 2004. Metabolic regulation of growth hormone by free fatty acids, somatostatin, and ghrelin in HIV-lipodystrophy. *American Journal of Physiology Endocrinology and Metabolism* 286:E296–E303.
- [273] Bohdjalian, A., Langer, F.B., Shakeri-Leidenmuhler, S., Gfrerer, L., Ludvik, B., Zacherl, J., et al., 2010. Sleeve gastrectomy as sole and definitive bariatric procedure: 5-year results for weight loss and ghrelin. *Obesity Surgery* 20: 535–540.
- [274] Dimitriadis, E., Daskalakis, M., Kampa, M., Peppe, A., Papadakis, J.A., Melissas, J., 2013. Alterations in gut hormones after laparoscopic sleeve gastrectomy: a prospective clinical and laboratory investigational study. *Annals of Surgery* 257:647–654.
- [275] Hady, H.R., Dadan, J., Golaszewski, P., Safiejko, K., 2012. Impact of laparoscopic sleeve gastrectomy on body mass index, ghrelin, insulin and lipid levels in 100 obese patients. *Wideochirurgia i inne techniki malo inwazyjne = Videosurgery and other miniinvasive techniques/kwartalniki pod patronatem Sekcji Wideochirurgii TChP oraz Sekcji Chirurgii Bariatrycznej TChP* 7:251–259.
- [276] Kotidis, E.V., Koliakos, G.G., Baltzopoulos, V.G., Ioannidis, K.N., Yovos, J.G., Papavramidis, S.T., 2006. Serum ghrelin, leptin and adiponectin levels before and after weight loss: comparison of three methods of treatment—a prospective study. *Obesity Surgery* 16:1425–1432.
- [277] Langer, F.B., Reza Hoda, M.A., Bohdjalian, A., Felberbauer, F.X., Zacherl, J., Wenzl, E., et al., 2005. Sleeve gastrectomy and gastric banding: effects on plasma ghrelin levels. *Obesity Surgery* 15:1024–1029.
- [278] Lee, W.J., Chen, C.Y., Chong, K., Lee, Y.C., Chen, S.C., Lee, S.D., 2011. Changes in postprandial gut hormones after metabolic surgery: a comparison of gastric bypass and sleeve gastrectomy. *Surgery for Obesity and Related Diseases: Official Journal of the American Society for Bariatric Surgery* 7:683–690.
- [279] Peterli, R., Steinert, R.E., Woelnerhanssen, B., Peters, T., Christoffel-Courtin, C., Gass, M., et al., 2012. Metabolic and hormonal changes after laparoscopic Roux-en-Y gastric bypass and sleeve gastrectomy: a randomized, prospective trial. *Obesity Surgery* 22:740–748.
- [280] Stefater, M.A., Wilson-Perez, H.E., Chambers, A.P., Sandoval, D.A., Seeley, R.J., 2012. All bariatric surgeries are not created equal: insights from mechanistic comparisons. *Endocrine Reviews* 33:595–622.
- [281] Bose, M., Machineni, S., Olivan, B., Teixeira, J., McGinty, J.J., Bawa, B., et al., 2010. Superior appetite hormone profile after equivalent weight loss by gastric bypass compared to gastric banding. *Obesity (Silver Spring)* 18: 1085–1091.
- [282] Uchida, A., Zechner, J.F., Mani, B.K., Park, W.M., Aguirre, V., Zigman, J.M., 2014. Altered ghrelin secretion in mice in response to diet-induced obesity and Roux-en-Y gastric bypass. *Molecular Metabolism* 3:717–730.
- [283] Chambers, A.P., Kirchner, H., Wilson-Perez, H.E., Willency, J.A., Hale, J.E., Gaylann, B.D., et al., 2013. The effects of vertical sleeve gastrectomy in rodents are ghrelin independent. *Gastroenterology* 144:50–52 e55.
- [284] Hosoda, H., Kangawa, K., 2008. The autonomic nervous system regulates gastric ghrelin secretion in rats. *Regulatory Peptides* 146:12–18.
- [285] Broglio, F., Gottero, C., Van Koetsveld, P., Prodam, F., Destefanis, S., Benso, A., et al., 2004. Acetylcholine regulates ghrelin secretion in humans. *The Journal of Clinical Endocrinology and Metabolism* 89:2429–2433.
- [286] Simonian, H.P., Kresge, K.M., Boden, G.H., Parkman, H.P., 2005. Differential effects of sham feeding and meal ingestion on ghrelin and pancreatic polypeptide levels: evidence for vagal efferent stimulation mediating ghrelin release. *Neurogastroenterology and Motility: The Official Journal of the European Gastrointestinal Motility Society* 17:348–354.
- [287] Iwakura, H., Ariyasu, H., Hosoda, H., Yamada, G., Hosoda, K., Nakao, K., et al., 2011. Oxytocin and dopamine stimulate ghrelin secretion by the ghrelin-producing cell line, MGN3-1 in vitro. *Endocrinology* 152:2619–2625.
- [288] Gagnon, J., Anini, Y., 2012. Insulin and norepinephrine regulate ghrelin secretion from a rat primary stomach cell culture. *Endocrinology* 153:3646–3656.
- [289] Nederkoorn, C., Smulders, F.T., Jansen, A., 2000. Cephalic phase responses, craving and food intake in normal subjects. *Appetite* 35:45–55.
- [290] Monteleone, P., Serritella, C., Martiadis, V., Maj, M., 2008. Deranged secretion of ghrelin and obestatin in the cephalic phase of vagal stimulation in women with anorexia nervosa. *Biological Psychiatry* 64:1005–1008.
- [291] Crum, A.J., Corbin, W.R., Brownell, K.D., Salovey, P., 2011. Mind over milkshakes: mindsets, not just nutrients, determine ghrelin response. *Health Psychology: Official Journal of the Division of Health Psychology, American Psychological Association* 30:424–429 discussion 430–421.
- [292] Monteleone, P., Serritella, C., Scognamiglio, P., Maj, M., 2010. Enhanced ghrelin secretion in the cephalic phase of food ingestion in women with bulimia nervosa. *Psychoneuroendocrinology* 35:284–288.
- [293] Williams, D.L., Cummings, D.E., Grill, H.J., Kaplan, J.M., 2003. Meal-related ghrelin suppression requires postgastric feedback. *Endocrinology* 144: 2765–2767.
- [294] Seoane, L.M., Al-Massadi, O., Barreiro, F., Dieguez, C., Casanueva, F.F., 2007. Growth hormone and somatostatin directly inhibit gastric ghrelin secretion. An in vitro organ culture system. *Journal of Endocrinological Investigation* 30:RC22–25.
- [295] Sakata, I., Nakano, Y., Osborne-Lawrence, S., Rovinsky, S.A., Lee, C.E., Perello, M., et al., 2009. Characterization of a novel ghrelin cell reporter mouse. *Regulatory Peptides* 155:91–98.
- [296] Foster-Schubert, K.E., Overduin, J., Prudom, C.E., Liu, J., Callahan, H.S., Gaylann, B.D., et al., 2008. Acyl and total ghrelin are suppressed strongly by ingested proteins, weakly by lipids, and biphasically by carbohydrates. *The Journal of Clinical Endocrinology and Metabolism* 93:1971–1979.

- [297] Geliebter, A., Carnell, S., Gluck, M.E., 2013. Cortisol and ghrelin concentrations following a cold pressor stress test in overweight individuals with and without night eating. *International Journal of Obesity (London)* 37:1104–1108.
- [298] Schellekens, H., Finger, B.C., Dinan, T.G., Cryan, J.F., 2012. Ghrelin signalling and obesity: at the interface of stress, mood and food reward. *Pharmacology & Therapeutics* 135:316–326.
- [299] Munding, T.O., Cummings, D.E., Taborsky Jr., G.J., 2006. Direct stimulation of ghrelin secretion by sympathetic nerves. *Endocrinology* 147:2893–2901.
- [300] de la Cour, C.D., Norlen, P., Hakanson, R., 2007. Secretion of ghrelin from rat stomach ghrelin cells in response to local microinfusion of candidate messenger compounds: a microdialysis study. *Regulatory Peptides* 143:118–126.
- [301] Seoane, L.M., Al-Massadi, O., Caminos, J.E., Tovar, S.A., Dieguez, C., Casanueva, F.F., 2007. Sensory stimuli directly acting at the central nervous system regulate gastric ghrelin secretion. an ex vivo organ culture study. *Endocrinology* 148:3998–4006.
- [302] Janssen, S., Laermans, J., Verhulst, P.J., Thijs, T., Tack, J., Depoortere, I., 2011. Bitter taste receptors and alpha-gustducin regulate the secretion of ghrelin with functional effects on food intake and gastric emptying. *Proceedings of the National Academy of Sciences of the United States of America* 108:2094–2099.
- [303] Hass, N., Schwarzenbacher, K., Breer, H., 2010. T1R3 is expressed in brush cells and ghrelin-producing cells of murine stomach. *Cell and Tissue Research* 339:493–504.
- [304] Al Massadi, O., Lear, P.V., Muller, T.D., Lopez, M., Dieguez, C., Tschop, M.H., et al., 2014. Review of novel aspects of the regulation of ghrelin secretion. *Current Drug Metabolism* 15:398–413.
- [305] Prudom, C., Liu, J., Patrie, J., Gaylinn, B.D., Foster-Schubert, K.E., Cummings, D.E., et al., 2010. Comparison of competitive radioimmunoassays and two-site sandwich assays for the measurement and interpretation of plasma ghrelin levels. *The Journal of Clinical Endocrinology and Metabolism* 95:2351–2358.
- [306] Engelstoft, M.S., Park, W.M., Sakata, I., Kristensen, L.V., Husted, A.S., Osborne-Lawrence, S., et al., 2013. Seven transmembrane G protein-coupled receptor repertoire of gastric ghrelin cells. *Molecular Metabolism* 2:376–392.
- [307] Sakata, I., Park, W.M., Walker, A.K., Piper, P.K., Chuang, J.C., Osborne-Lawrence, S., et al., 2012. Glucose-mediated control of ghrelin release from primary cultures of gastric mucosal cells. *American Journal of Physiology Endocrinology and Metabolism* 302:E1300–E1310.
- [308] Lu, X., Zhao, X., Feng, J., Liou, A.P., Anthony, S., Pechhold, S., et al., 2012. Postprandial inhibition of gastric ghrelin secretion by long-chain fatty acid through GPR120 in isolated gastric ghrelin cells and mice. *American Journal of Physiology Gastrointestinal and Liver Physiology* 303:G367–G376.
- [309] Gagnon, J., Anini, Y., 2013. Glucagon stimulates ghrelin secretion through the activation of MAPK and EPAC and potentiates the effect of norepinephrine. *Endocrinology* 154:666–674.
- [310] Iwakura, H., Li, Y., Ariyasu, H., Hosoda, H., Kanamoto, N., Bando, M., et al., 2010. Establishment of a novel ghrelin-producing cell line. *Endocrinology* 151:2940–2945.
- [311] Lefkowitz, R.J., Stadel, J.M., Caron, M.G., 1983. Adenylate cyclase-coupled beta-adrenergic receptors: structure and mechanisms of activation and desensitization. *Annual Review of Biochemistry* 52:159–186.
- [312] Levitzki, A., 1986. Beta-adrenergic receptors and their mode of coupling to adenylate cyclase. *Physiological Reviews* 66:819–854.
- [313] De Vriese, C., Gregoire, F., Lema-Kisoka, R., Waelbroeck, M., Robberecht, P., Delporte, C., 2004. Ghrelin degradation by serum and tissue homogenates: identification of the cleavage sites. *Endocrinology* 145:4997–5005.
- [314] Mizutani, M., Atsuchi, K., Asakawa, A., Matsuda, N., Fujimura, M., Inui, A., et al., 2009. Localization of acyl ghrelin- and des-acyl ghrelin-immunoreactive cells in the rat stomach and their responses to intragastric pH. *American Journal of Physiology Gastrointestinal and Liver Physiology* 297:G974–G980.
- [315] Blatnik, M., Soderstrom, C.I., Dysinger, M., Fraser, S.A., 2012. Prandial ghrelin attenuation provides evidence that des-acyl ghrelin may be an artifact of sample handling in human plasma. *Bioanalysis* 4:2447–2455.
- [316] Thompson, N.M., Gill, D.A., Davies, R., Loveridge, N., Houston, P.A., Robinson, I.C., et al., 2004. Ghrelin and des-octanoyl ghrelin promote adipogenesis directly in vivo by a mechanism independent of the type 1a growth hormone secretagogue receptor. *Endocrinology* 145:234–242.
- [317] Toshinai, K., Yamaguchi, H., Sun, Y., Smith, R.G., Yamanaka, A., Sakurai, T., et al., 2006. Des-acyl ghrelin induces food intake by a mechanism independent of the growth hormone secretagogue receptor. *Endocrinology* 147:2306–2314.
- [318] Zhang, W., Chai, B., Li, J.Y., Wang, H., Mulholland, M.W., 2008. Effect of des-acyl ghrelin on adiposity and glucose metabolism. *Endocrinology* 149:4710–4716.
- [319] Li, L., Zhang, L.K., Pang, Y.Z., Pan, C.S., Qi, Y.F., Chen, L., et al., 2006. Cardioprotective effects of ghrelin and des-octanoyl ghrelin on myocardial injury induced by isoproterenol in rats. *Acta Pharmacologica Sinica* 27:527–535.
- [320] Baldanzi, G., Filigheddu, N., Cutrupi, S., Catapano, F., Bonissoni, S., Fubini, A., et al., 2002. Ghrelin and des-acyl ghrelin inhibit cell death in cardiomyocytes and endothelial cells through ERK1/2 and PI 3-kinase/AKT. *The Journal of Cell Biology* 159:1029–1037.
- [321] Chen, C.Y., Inui, A., Asakawa, A., Fujino, K., Kato, I., Chen, C.C., et al., 2005. Des-acyl ghrelin acts by CRF type 2 receptors to disrupt fasted stomach motility in conscious rats. *Gastroenterology* 129:8–25.
- [322] Broglio, F., Gottero, C., Prodam, F., Gauna, C., Muccioli, G., Papotti, M., et al., 2004. Non-acylated ghrelin counteracts the metabolic but not the neuroendocrine response to acylated ghrelin in humans. *The Journal of Clinical Endocrinology and Metabolism* 89:3062–3065.
- [323] Barazzoni, R., Zanetti, M., Ferreira, C., Vinci, P., Pirulli, A., Mucci, M., et al., 2007. Relationships between desacylated and acylated ghrelin and insulin sensitivity in the metabolic syndrome. *The Journal of Clinical Endocrinology and Metabolism* 92:3935–3940.
- [324] Cederberg, H., Koivisto, V.M., Jokelainen, J., Surcel, H.M., Keinanen-Kiukaanniemi, S., Rajala, U., 2012. Unacylated ghrelin is associated with changes in insulin sensitivity and lipid profile during an exercise intervention. *Clinical Endocrinology* 76:39–45.
- [325] Tong, J., Davis, H.W., Summer, S., Benoit, S.C., Haque, A., Bidlingmaier, M., et al., 2014. Acute administration of unacylated ghrelin has no effect on Basal or stimulated insulin secretion in healthy humans. *Diabetes* 63:2309–2319.
- [326] Asakawa, A., Inui, A., Fujimiya, M., Sakamaki, R., Shinfuku, N., Ueta, Y., et al., 2005. Stomach regulates energy balance via acylated ghrelin and desacyl ghrelin. *Gut* 54:18–24.
- [327] Delhanty, P.J., Sun, Y., Visser, J.A., van Kerkwijk, A., Huisman, M., van Ijcken, W.F., et al., 2010. Unacylated ghrelin rapidly modulates lipogenic and insulin signaling pathway gene expression in metabolically active tissues of GHSR deleted mice. *PLoS One* 5:e11749.
- [328] Delhanty, P.J., Huisman, M., Baldeon-Rojas, L.Y., van den Berge, I., Greffhorst, A., Aribat, T., et al., 2013. Des-acyl ghrelin analogs prevent high-fat-diet-induced dysregulation of glucose homeostasis. *FASEB Journal: Official Publication of the Federation of American Societies for Experimental Biology* 27:1690–1700.
- [329] Granata, R., Settanni, F., Julien, M., Nano, R., Togliatto, G., Trombetta, A., et al., 2012. Des-acyl ghrelin fragments and analogues promote survival of pancreatic beta-cells and human pancreatic islets and prevent diabetes in streptozotocin-treated rats. *Journal of Medicinal Chemistry* 55:2585–2596.
- [330] Granata, R., Settanni, F., Biancone, L., Trovato, L., Nano, R., Bertuzzi, F., et al., 2007. Acylated and unacylated ghrelin promote proliferation and inhibit apoptosis of pancreatic beta-cells and human islets: involvement of 3',5'-

- cyclic adenosine monophosphate/protein kinase A, extracellular signal-regulated kinase 1/2, and phosphatidylinositol 3-Kinase/Akt signaling. *Endocrinology* 148:512–529.
- [331] Granata, R., Settanni, F., Trovato, L., Destefanis, S., Gallo, D., Martinetti, M., et al., 2006. Unacylated as well as acylated ghrelin promotes cell survival and inhibit apoptosis in HIT-T15 pancreatic beta-cells. *Journal of Endocrinological Investigation* 29:RC19–22.
- [332] Granata, R., Volante, M., Settanni, F., Gauna, C., Ghe, C., Annunziata, M., et al., 2010. Unacylated ghrelin and obestatin increase islet cell mass and prevent diabetes in streptozotocin-treated newborn rats. *Journal of Molecular Endocrinology* 45:9–17.
- [333] Heppner, K.M., Piechowski, C.L., Muller, A., Ottaway, N., Sisley, S., Smiley, D.L., et al., 2014. Both acyl and des-acyl ghrelin regulate adiposity and glucose metabolism via central nervous system ghrelin receptors. *Diabetes* 63:122–131.
- [334] Carlini, V.P., Varas, M.M., Cragolini, A.B., Schioth, H.B., Scimone, T.N., de Barioglio, S.R., 2004. Differential role of the hippocampus, amygdala, and dorsal raphe nucleus in regulating feeding, memory, and anxiety-like behavioral responses to ghrelin. *Biochemical and Biophysical Research Communications* 313:635–641.
- [335] Hewson, A.K., Dickson, S.L., 2000. Systemic administration of ghrelin induces Fos and Egr-1 proteins in the hypothalamic arcuate nucleus of fasted and fed rats. *Journal of Neuroendocrinology* 12:1047–1049.
- [336] Schaeffer, M., Langlet, F., Lafont, C., Molino, F., Hodson, D.J., Roux, T., et al., 2013. Rapid sensing of circulating ghrelin by hypothalamic appetite-modifying neurons. *Proceedings of the National Academy of Sciences of the United States of America* 110:1512–1517.
- [337] Zorrilla, E.P., Iwasaki, S., Moss, J.A., Chang, J., Otsuji, J., Inoue, K., et al., 2006. Vaccination against weight gain. *Proceedings of the National Academy of Sciences of the United States of America* 103:13226–13231.
- [338] Chapman, I.M., Bach, M.A., Van Cauter, E., Farmer, M., Krupa, D., Taylor, A.M., et al., 1996. Stimulation of the growth hormone (GH)-insulin-like growth factor I axis by daily oral administration of a GH secretagogue (MK-677) in healthy elderly subjects. *The Journal of Clinical Endocrinology and Metabolism* 81:4249–4257.
- [339] Murphy, M.G., Plunkett, L.M., Gertz, B.J., He, W., Wittreich, J., Polvino, W.M., et al., 1998. MK-677, an orally active growth hormone secretagogue, reverses diet-induced catabolism. *The Journal of Clinical Endocrinology and Metabolism* 83:320–325.
- [340] Inoue, H., Sakamoto, Y., Kangawa, N., Kimura, C., Ogata, T., Fujieda, K., et al., 2011. Analysis of expression and structure of the rat GH-secretagogue/ghrelin receptor (Ghsr) gene: roles of epigenetic modifications in transcriptional regulation. *Molecular and Cellular Endocrinology* 345:1–15.
- [341] Bach, M.A., Rockwood, K., Zetterberg, C., Thamsborg, G., Hebert, R., Devogelaer, J.P., et al., 2004. The effects of MK-0677, an oral growth hormone secretagogue, in patients with hip fracture. *Journal of the American Geriatrics Society* 52:516–523.
- [342] Lainscak, M., von Haehling, S., Doehner, W., Anker, S.D., 2012. The obesity paradox in chronic disease: facts and numbers. *Journal of Cachexia, Sarcopenia and Muscle* 3:1–4.
- [343] von Haehling, S., Morley, J.E., Anker, S.D., 2012. From muscle wasting to sarcopenia and myopenia: update 2012. *Journal of Cachexia, Sarcopenia and Muscle* 3:213–217.
- [344] Anker, M.S., von Haehling, S., Springer, J., Banach, M., Anker, S.D., 2013. Highlights of the mechanistic and therapeutic cachexia and sarcopenia research 2010 to 2012 and their relevance for cardiology. *International Journal of Cardiology* 162:73–76.
- [345] Miki, K., Maekura, R., Nagaya, N., Nakazato, M., Kimura, H., Murakami, S., et al., 2012. Ghrelin treatment of cachectic patients with chronic obstructive pulmonary disease: a multicenter, randomized, double-blind, placebo-controlled trial. *PLoS One* 7:e35708.
- [346] Akashi, Y.J., Palus, S., Datta, R., Halem, H., Taylor, J.E., Thoene-Reineke, C., et al., 2009. No effects of human ghrelin on cardiac function despite profound effects on body composition in a rat model of heart failure. *International Journal of Cardiology* 137:267–275.
- [347] Palus, S., von Haehling, S., Doehner, W., Datta, R., Zhang, J., Dong, J.Z., et al., 2013. Effect of application route of the ghrelin analog BIM-28131 (RM-131) on body weight and body composition in a rat heart failure model. *International Journal of Cardiology* 168:2369–2374.
- [348] Lenk, K., Palus, S., Schur, R., Datta, R., Dong, J., Culler, M.D., et al., 2013. Effect of ghrelin and its analogues, BIM-28131 and BIM-28125, on the expression of myostatin in a rat heart failure model. *Journal of Cachexia, Sarcopenia and Muscle* 4:63–69.
- [349] Vila, G., Grimm, G., Resl, M., Heinisch, B., Einwallner, E., Esterbauer, H., et al., 2012. B-type natriuretic peptide modulates ghrelin, hunger, and satiety in healthy men. *Diabetes* 61:2592–2596.
- [350] Lund, L.H., Williams, J.J., Freda, P., LaManca, J.J., LeJemtel, T.H., Mancini, D.M., 2009. Ghrelin resistance occurs in severe heart failure and resolves after heart transplantation. *European Journal of Heart Failure* 11:789–794.
- [351] Strassburg, S., Anker, S.D., Castaneda, T.R., Burget, L., Perez-Tilve, D., Pfluger, P.T., et al., 2008. Long-term effects of ghrelin and ghrelin receptor agonists on energy balance in rats. *American Journal of Physiology Endocrinology and Metabolism* 295:E78–E84.
- [352] Koch, L., 2013. Growth hormone in health and disease: novel ghrelin mimetic is safe and effective as a GH stimulation test. *Nature Reviews Endocrinology* 9:315.
- [353] Currow, D.C., Abernethy, A.P., 2014. Anamorelin hydrochloride in the treatment of cancer anorexia-cachexia syndrome. *Future Oncology* 10:789–802.
- [354] Camilleri, M., Acosta, A., 2014. Emerging treatments in Neurogastroenterology: relamorelin: a novel gastrocolokinetic synthetic ghrelin agonist. *Neurogastroenterology and Motility: The Official Journal of the European Gastrointestinal Motility Society* 27:324–332.
- [355] Sha, L.V.d.P., L.H., Szurszewski, J., 2014. Effects of RM-131 on circular smooth muscle in human and mouse colon and colonic intraluminal pressure in conscious mice. *Gastroenterology* 146:S-363.
- [356] Shin, A., Camilleri, M., Busciglio, I., Burton, D., Smith, S.A., Vella, A., et al., 2013. The ghrelin agonist RM-131 accelerates gastric emptying of solids and reduces symptoms in patients with type 1 diabetes mellitus. *Clinical Gastroenterology and Hepatology: The Official Clinical Practice Journal of the American Gastroenterological Association* 11:1453–1459 e1454.
- [357] Shin, A., Camilleri, M., Busciglio, I., Burton, D., Stoner, E., Noonan, P., et al., 2013. Randomized controlled phase Ib study of ghrelin agonist, RM-131, in type 2 diabetic women with delayed gastric emptying: pharmacokinetics and pharmacodynamics. *Diabetes Care* 36:41–48.
- [358] Dass, N.B., Munonyara, M., Bassil, A.K., Hervieu, G.J., Osbourne, S., Corcoran, S., et al., 2003. Growth hormone secretagogue receptors in rat and human gastrointestinal tract and the effects of ghrelin. *Neuroscience* 120:443–453.
- [359] Deboer, M.D., 2011. Use of ghrelin as a treatment for inflammatory bowel disease: mechanistic considerations. *International Journal of Peptides* 2011:189242.