



Article Artefacts in Field Trial Research—Lateral Ammonia Fluxes Confound Fertiliser Plot Experiments

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Abstract: Agricultural ammonia (NH₃) emissions can have serious environmental impacts, lower fertiliser nitrogen-use efficiencies, and cause economic losses. NH3 losses may not only occur directly from organic fertilisers such as biogas digestates when applied to crops, the crops themselves may also be a source of ammonia emissions. Wheat yields from 14 years of an organic small plot fertiliser trial fertilised with biogas digestate were analysed to determine if there was significant lateral N transfer between plots. A simple NH₃ loss/gain model was developed to calculate possible N gains and losses via NH₃ volatilisation from the applied digestate. This model was tested using NH₃ volatilisation measurements. In addition, ¹⁵N isotope measurements of crop biomass were used to analyse plant N uptake. While digestate fertilisation increased wheat yields, yield patterns indicated that NH₃ emissions from plots fertilised with biogas digestate affected yields in neighbouring unfertilised plots. Measurements of ammonia losses and gains in the field validated our modelling results, showing that 55% of digestate NH4⁺-N was volatilised. ¹⁵N isotope analysis indicated that crops took up as much as 30 kg ha⁻¹ NH₃-N volatilised from digestate, and that plots closer to fertilised plots took up more of this NH₃-N than crops further away from fertilised plots. Our results imply that neither the results from the fertilised plots nor from the unfertilised plots are without bias. To avoid inadvertently introducing artefacts into fertiliser field trials, plot sizes need to be increased and treatments situated further apart.

Keywords: experimental design; ammonia fluxes; isotope; biogas; digestate; organic agriculture

1. Introduction

It is estimated that globally, only about 50% of the nitrogen (N) applied as fertiliser to crops is actually captured by crops [1], with the remainder immobilised in soil or lost to the environment via denitrification, volatilisation, gaseous plant N loss, leaching, and surface runoff [2]. In Germany, for example, 95% of ammonia (NH₃) emissions come from agriculture [3,4]. Agriculture, therefore, contributes substantially (also via residues from N-fixing crops) to increases in the overall level of reactive N in the environment [5]. N losses can have serious environmental impacts, such as the eutrophication and acidification of terrestrial and aquatic ecosystems [4,6]. Aside from negative effects on the environment, low N-use efficiencies also represent economic losses. Minimising N loss is particularly important in organic agriculture, where the supply of plant-available (i.e., mineral) N is limited. Fertiliser trials are, therefore, used in agricultural research to assess and optimise N-use efficiency by comparing the N uptake of crops in fertilised plots with the N uptake of crops in unfertilised plots. However, these trials need to be carefully designed and managed, particularly with regard to control plots, to ensure that artefacts are not accidentally introduced and/or do not affect experimental outcomes [7]. Artefacts are effects in an experiment that occur as a result of the experiment and are not present naturally to the same degree. For example, fertiliser drag, where a fertiliser treatment is unintentionally moved



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Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). from one plot to another due to the size and control of machinery during application or tillage, can be eliminated by leaving alleys between plots. However, this means that plants at plot borders benefit from the lack of competition for nutrients, water, and light, so these plants are usually not included in any analyses [8]. These measures may still not be enough to prevent fertilised plots from influencing neighbouring plots. If two adjacent plots receive differing amounts of fertiliser, fertiliser can spread to the root zone of the adjacent plot, disadvantaging the plot receiving the higher amount of fertiliser [9]. Depending on the topographic gradient of the field being used, soil erosion, runoff, and subsurface flows containing nitrate (NO_3^-) could also result in N being transferred between plots [10,11]. Nutrient transfer may also occur via gaseous N emissions from one plot to another. Plants are not only able to take up inorganic N from soils but can also use atmospheric N sources such as NH_3 and NO_x [12]. If the stomatal ammonia compensation point of plants is lower than the atmospheric NH₃ concentration, as could be the case in unfertilised control plots, plants can take up ammonia with their leaves, whereas if the stomatal ammonia compensation point is higher than the atmospheric NH_3 concentration, plants may emit ammonia [13,14]. Well-fertilised crops are, thus, a net source of NH₃ to the atmosphere [15]. Therefore, not only the fertilisers but the plants themselves could act as an N source for plants in other plots, in particular, transferring N from fertilised to unfertilised plots.

NH₃ volatilisation involves several steps (NH₃ in equilibrium in solution with NH₄⁺; diffusion of NH₃ to the site of volatilisation; volatilisation of NH₃; diffusion of NH₃ away from the site of volatilisation) during which isotopic fractionation can occur [16]. Due to this fractionation, volatilised NH₃ is depleted in ¹⁵N compared to NH₃ in the original substrate [16]. δ^{15} N isotope analysis can therefore be used to identify the uptake of volatilised NH₃ by crop biomass [17].

The extent of gaseous emissions from fertilisers will depend on the type of fertiliser used. Biogas digestates are organic fertilisers resulting from the anaerobic digestion of biomass for biogas production. Depending on the feedstocks used in the digestion process, biogas digestates can have higher ammonium (NH_4^+) :N ratios than the original feedstocks and crop NH_4^+ -N recoveries equivalent to mineral fertilisers [18]. The fermentation process also increases pH, and pH is one of the main drivers of NH₃ emissions from organic fertilisers [19]. This higher pH, coupled with a higher NH_4^+ -N content, generates an intensive NH₃ volatilisation process when digestates are applied to crops using conventional methods [19,20]. Digestate viscosity and dry matter content also influence NH_3 emissions [15]. While digestates of animal slurries usually have a lower dry matter content than the slurries themselves [21], this is not the case if plant material is used as the feedstock in biogas production [19]. For example, in organic agriculture, clover–grass biomass is often used as a feedstock in biogas production. This results in digestates with a higher viscosity than digestates where animal manures or other crops are used as substrates [22]. With increasing viscosity, digestates infiltrate more slowly into soils, increasing NH₃ losses [23]. Ammonia emissions occur when digestates, like other organic fertilisers, are exposed to air [15], and therefore, if digestates are not incorporated into soils but instead only applied to the soil surface, this will increase NH_3 volatilisation [4]. On the other hand, digestates contain large amounts of small plant particles (presumably even more so if clover-grass was used as a feedstock for biogas production), which, if they remain on the soil surface, dry out and form a thin crust which can reduce NH_3 emissions [19]. NH_3 emissions are also affected by the crop type, with digestate application in wheat showing higher potential NH₃ losses than in maize due to the timing of digestate applications and leaf area index of the crop [22,24]. Losses of up to 40% of the total NH_4^+ -N applied have been reported in wheat crops when digestate was applied using the trailing hose method [25]. Due to these large losses, NH₃ drift from plots fertilised with digestate to other plots in field experiments has been reported [26].

We analysed patterns in wheat (*Triticum aestivum* L.) yield data for the period 2007–2021 from a long-term organic field trial to determine if ammonia emissions from biogas digestate had an effect on yields. In addition, NH₃ emissions as a result of digestate fertilisation

and changes in isotope composition of crop biomass were measured to analyse if these emissions affected neighbouring plots. To date, most studies examining N dynamics and NH₃ emissions from digestate application have been short-term studies where there is a risk that the priming effect (i.e., an increase in soil organic matter decomposition due to the addition of organic matter) and differing weather conditions from year to year may affect results [27]. In addition, after 15 years of digestate fertilisation, the gradient between neighbouring fertilised and unfertilised plots is likely to be higher than in a short-term experiment. The objective of this study was to investigate (a) if there is a significant lateral N transfer between neighbouring plots in small-plot fertiliser trials, (b) the effects of any N transfer on wheat yields and N uptake of crops, and (c) how this affects the interpretation of results from these trials.

2. Materials and Methods

2.1. Experimental Site

Our field trial was situated in Viehhausen, located approximately 8 km west of Freising in southern Germany, 490 m above sea level in the Tertiary hill country, an undulating landscape developed in Tertiary sediments and overlain by a thin loess cover. Using the US soil taxonomy [28], the soil at the experimental site is categorized as a Hapludalf derived from loess with silty loam texture down to at least 1 m. Using the World Reference Base [29], the soil is a Haplic Luvisol (Manganiferric, Siltic). The experiment was situated on a slope facing northeast with a gradient of about 9%. Average annual temperature and precipitation for the period 2007–2022 in Viehhausen were 9.3 °C and 782 mm a⁻¹, respectively.

2.2. Experimental Design

This study looked at two factors, fertilisation and crop rotation (CR). The main factor fertilisation was assigned in two levels to plots within the factor CR; there were 10 different four-year CRs (Table 1), and the 32 plots of each CR were divided into 16 fertilised and 16 unfertilised plots. A detailed plot plan can be found in the Appendix A (Figure A1). The CRs were laid out in a replicated control design. CR1a was located at the start of the field, repeated in the middle (CR1b) and at the opposite end of the field (CR1c) in order to capture any trends in yield potential across the site. Each crop of every CR was cultivated each year; the CRs were, therefore, laid out in columns running down the field, divided into four blocks. The crops moved up a block each year. Within each CR, there were 8 replications in each block (4 fertilised and 4 unfertilised plots). Plots measured 6 m \times 12 m (plots on both outer edges of the trial measured 9 m \times 12 m), with alleys of 3 m between each row of plots and 9 m between each block. The field trial, therefore, had a total size of nearly 4 ha.

The first year in each CR was a clover–grass ley, followed by a year of winter wheat; the CRs differed in the third and fourth years. In this study, we will use the data from winter wheat in year 2, which covers a wide range of conditions given the many years, the different CRs, and the different positions along and across the slope. For the period 2007–2011, the wheat cultivar Enorm was planted; in 2012, Stava; for the period 2013–2021, Florian; and in 2022, Moschus. Planting took place between mid-October and early November after ploughing. Grain yields were determined using a plot combine by harvesting an area within each plot of 15 to 18 m². Yields were adjusted to 86% dry matter content. Due to poor crop development, wheat yields from 2019 were not included in the analysis. In addition, yield data for CR1a from 2020 and 2021 were not included due to changes made to these plots for the ammonia measurements. Therefore, in this study, a total of 14 years (CR1a 12 years) of wheat yield data and N supply data from 13 years (CR1a 11 years) were used.

The N content of grain samples was analysed using an N analyser (Vario Max, Elementar Analysensysteme GmbH (Langenselbold, Germany), Dumas combustion method).

Crop Rotation	Year 1	Year 2	Year 3 Cover Crop	Year 3	Year 4
CR1a	Clover-grass	Wheat (39)	Rye (10)	Maize (51)	Triticale (33)
CR2	Clover–grass	Wheat (28)	Rye (10)	Maize (46)	Wheat (30)
CR3	Clover-grass	Wheat (26)	Legume/non-legume mixture (7)	Lupin ^{1,2}	Wheat ³ (29)
CR4	Clover-grass	Wheat (29)	Legume/non-legume mixture (7)	Soybean ¹	Wheat ⁴ (28)
CR5	Clover-grass	Wheat (40)	White clover (3)	Maize and white clover (42)	Triticale (35)
CR6	Clover–grass	Wheat (35)	White clover (3)	Maize and white clover (37)	Maize and white clover (38)
CR1b	Clover-grass	Wheat (39)	Rye (10)	Maize (51)	Triticale (33)
CR7	Clover–grass	Wheat (35)	Legume/non-legume mixture (6)	Maize (42)	Triticale (35)
CR8	Clover-grass	Wheat (40)	Legume/non-legume mixture (6)	Maize (37)	Maize ⁵ (30)
CR9	Clover–grass (28)	Wheat (64)	Clover–grass	Clover–grass	Triticale (45)
CR10	Clover–grass (46)	Wheat (68)	Clover–grass	Clover–grass	Clover–grass (19)
CR1c	Clover–grass	Wheat (39)	Rye (10)	Maize (51)	Triticale (33)

Table 1. Plan of crop rotations and the mean amount of digestate fertilisation in the fertilised plots 2008–2022 (digestate amount of the fertilised plots in parentheses, $m^3 ha^{-1} a^{-1}$).

¹ Sorghum was grown in this position in the period 2007–2011 and was fertilised with 55 m³ ha⁻¹ a⁻¹ of digestate; ² 2012–2019: Field bean; ³ 2007–2012: Triticale; ⁴ 2007–2012: Sunflower (whole crop) and undersown clover–grass; ⁵ 2007–2018: Sunflower (whole crop) and undersown clover–grass.

2.3. Digestate and Fertilisation

Each unfertilised plot was neighboured laterally by a fertilised plot on one side across the slope and on one side along the slope. There was no difference in treatment between fertilised and unfertilised plots except that fertilised plots were fertilised using biogas digestate. Each CR received a different amount of digestate, depending on the theoretical amount of digestate that could be produced from the estimated crop biomass of clovergrass, rye (whole crop) (Secale cereale L.), silage maize (Zea mays L.) and triticale (whole crop) (× Triticosecale Wittmack) grown in the fertilised plots of each CR. The total amount of digestate was then divided between the crops in the CR according to estimated nutrient requirements (Table 1). Thus, for the period 2007–2022 the wheat crop in year 2 of the CRs received the following average amounts of digestate-N (kg ha⁻¹ a⁻¹), range in parentheses: CR1a 182 (121–275), CR2 138 (96–197), CR3 135 (77–238), CR4 153 (77–298), CR5 210 (115-319), CR6 202 (77-319), CR1b 182 (121-275), CR7 174 (115-243), CR8 210 (115-319), CR9 308 (121–481), CR10 339 (121–542), CR1c 182 (121–275). The wheat crop was fertilised, depending on the CR, weather conditions, and crop development each year, with one or two doses of digestate between mid-March and mid-May. The digestate was applied using a slurry tanker fitted with trailing hoses. The digestate was produced by a local organic farmer from a feedstock mix of, on average, 61% silage from clover–grass and grass leys, and grassland biomass, 30% solid cattle manure, 6% silage maize, and the remainder cereal grains. From 2010 onwards, the digestate was separated into liquid and solid phases, and the liquid phase was used in this field trial. The average dry matter content of the digestate after separation of the solids was 8.1%, with a total N content of 6.3%; 52% of this was NH_4^+ -N (Table 2). The pH of the digestate was 7.8 (data from 2015 only).

Variable	Content (%, Dry Weight Basis, Standard Deviation in Parentheses)
Dry matter	8.1 (1.3)
Loss on ignition	72.3 (2.6)
Total nitrogen	6.3 (0.9)
Ammonium nitrogen	3.3 (0.6)
Phosphate (as P_2O_5)	2.2 (0.3)
Potassium (as K_2O)	8.9 (1.0)
Carbon	39.3 (0.9)
Sulfur	0.5 (0.05)

Table 2. Average digestate composition 2009–2022 (laboratory analysis by Agrolab Labor GmbH).

2.4. NH₃ Volatilisation Model

A simple NH₃ loss/gain model was created to calculate possible N gains and losses of plots via NH₃ volatilisation from the applied digestate. It was assumed that 52% of digestate N was NH₄⁺-N, in accordance with the digestate analysis (Table 2), and that 50% of this NH₄⁺-N was lost via NH₃ volatilisation to directly neighbouring plots, with the remaining 50% staying on the plot to which the digestate was applied. The weaker long-distance transport (to plots other than direct neighbours or beyond the experimental field) was not considered because the fine-grained experimental layout resulted in an almost uniform pattern of long-distance transport that only influenced the basic yield level (intercept of the regression) and would not result in any other detectable differences. In the second step, the assumption of 50% loss to direct neighbours was abandoned, and this fraction was used as a fitting parameter by maximizing R² between N supply and yield.

2.5. Ammonia Volatilisation Measurements

Two methods were used to measure ammonia volatilisation: (a) passive flux samplers (PFS) to measure ammonia gains and (b) the Dräger tube method (DTM) to measure ammonia concentrations in the air [30]. Measurements using PFS took place after digestate fertilisation in the wheat block in CRs 1a, 1b, 1c, 9, and 10 on 31 March 2021 and 14 March 2022. The fertilised plots in CRs 1a, 1b, and 1c received 20 m³ of digestate, and the fertilised plots in CRs 9 and 10 received 30 m³. The PFS were placed in four rows in the field trial, with eight PFS in each row, located at varying distances from a fertilised plot (Figures A1 and A2). The PFS were attached 20 cm above the vegetation with heightadjustable metal rods. The polythene vessels (250 mL capacity) had holes of 2 cm diameter to allow gas exchange and were filled with dilute (0.05 mol) sulfuric acid. The PFS were protected by a mosquito net for the duration of the sampling period to prevent insects from getting into the acid trap. The acid traps were emptied and refilled with sulfuric acid every afternoon for eight days at 4 p.m. The solutions were analysed using an autoanalyzer in the laboratory to determine if the sulfuric acid had reacted with ammonia to form ammonium sulfate and to analyse the resulting ammonium content. The amount of ammonia was then determined from the ammonium content.

In 2021 and 2022, the original plots of CR1a were divided in two to measure ammonia volatilisation according to DTM (Figures A1 and A2). Chambers were placed directly on the ground in vegetation in six fertilised and six unfertilised plots of CR1a (see Figure A3). The fertilised plots were fertilised with 20 m³ ha⁻¹ of digestate on 31 March 2021, 29 April 2021, 14 March 2022, and 28 April 2022 (115, 117, 127, and 99 kg ha⁻¹ N, respectively). Due to missing data, the values from 31 March 2021 are not included in this analysis. Air was sucked from the chambers through PTFE hoses by a portable electric pump into an indicator tube. The indicator tubes contained yellow bromophenol blue granules, which turned blue if ammonia was present in the air removed from the chambers, and also enabled the concentration of ammonia in ppm to be determined. To convert ammonia ppm to kg ha⁻¹, ammonia emission dynamics were calculated according to Pacholski [31] based on the time of day, air temperature, air pressure, wind speed, and number and time of pump strokes

required to achieve a particular colour change in ppm on the indicator tube. Ammonia concentrations measured in the unfertilised plots of CR1a located next to the fertilised plots (see Figures A1 and A2) were assumed to represent the background NH₃ concentration. To calculate the cumulative ammonia emissions from the fertilised soil, this background NH₃ concentration was subtracted from the measurements in the fertilised plots.

For weather conditions during ammonia measurements, see Table A1.

2.6. Measurement of N Uptake

In September 2014, NH₃ uptake was analysed using ¹⁵N isotope analysis in clovergrass plots in year 1 of the rotations. Only CRs 9 and 10 were fertilised with digestate in the clover-grass crop in year 1 (Table 1). This enabled us to analyse how isotopic composition, and therefore NH₃ uptake, changed with distance from a fertilised plot. This was not possible in the wheat crop as all CRs were fertilised. Only clover-grass plots in CRs 8, 9, 10 and 1c were used for the isotopic analysis. The clover-grass plots were sown with a mixture of red clover (*Trifolium pratense* L.), white clover (*Trifolium repens* L.), lucerne (*Medicago sativa* L.), and various grass species. In CR8, the clover-grass was undersown in the sunflower (*Helianthus annuus* L.) crop on 18 June 2013. In CRs 1c and 9, the clover-grass plots were sown with the clover-grass mixture and with oats (*Avena sativa* L.) as a nurse crop on 27 July 2013. In CR10, the clover-grass was sown on 3 August 2011. In 2014, the clover-grass was harvested on 20 May 2014, 7 July 2014, and 20 August 2014, with an additional harvest in CRs 1c and 9 on 24 September 2013. The clover-grass was fertilised with 20 m³ ha⁻¹ of digestate on 26 May 2014, 8 July 2014, and 22 August 2014 in CRs 9 and 10 only. In 2014, these plots received 364 kg ha⁻¹ of digestate-N in total, of which 215 kg ha⁻¹ was NH₄-N.

Biomass samples were collected by hand from five clover–grass plots in CRs 8, 9, 10 and 1c on 29 September 2014 (see Figure A4). Each biomass sample was separated into grass and legume components, and the grass biomass used for further analysis. These samples were dried for two hours at 105 °C and then for 48 h at 60 °C in a drying cabinet. The samples were then ground in a ball mill and dried overnight at 40 °C before being weighed into a tin cup. The N isotope composition of the samples was determined using an elemental analyzer (Carlo Erba NA 1110, Milan, Italy) interfaced (ConFlo II, Finnigan MAT, Bremen, Germany) to an isotope ratio mass spectrometer (Delta Plus, Finnigan MAT). Wheat flour was used as a control and analysed after every tenth sample. Accuracy, measured as the standard deviation of wheat flour replicates, was 0.10‰. Isotope values are presented in δ units, i.e., $\delta^{15}N = R_{sample}/R_{standard} - 1$, where R is the $^{15}N/^{14}N$ ratio, and $R_{standard}$ refers to atmospheric N₂.

Assuming that δ^{15} N in the NH₃ was -20% [17] and that NH₃ uptake in the plot located furthest away from the fertilised plot was negligible (NH₃ fraction = 0), meaning that δ^{15} N of the grass in this plot (2.90‰) reflected NH₃ taken up from the soil, the fraction of NH₃-N was calculated as follows:

Fraction N_{NH3} =
$$(\delta^{15}N_{\text{total}} - \delta^{15}N_{\text{soil}})/(\delta^{15}N_{\text{NH3}} - \delta^{15}N_{\text{soil}})$$
 (1)

Both assumptions are somewhat speculative, but this only influences the absolute value of the fraction of NH₃-N and not the change in NH₃-N with distance from the source. Furthermore, a small N transfer from the clover to the grass is conceivable. However, the fraction of legumes in the unfertilized clover–grass mixtures did not depend on the fertilisation level of the neighbouring plots [32]. Hence, even if some transfer had occurred, it would also have occurred in the plot located furthest away from the fertilised plot, which was used as the baseline value. The calculation of the NH₃-N fraction would not have been affected.

2.7. Statistical Analysis

Most statistical analyses were carried out using R version 4.2.1 [33] and CoStat 6.40 (CoHort Software, Monterey, CA, USA). To examine whether two correlation coefficients differed significantly when different rates of volatilisation were assumed, we applied the

Hotelling test of correlated sample sets [34]. Means are reported with their 95% confidence intervals (CI). Significance is indicated as *, **, *** for p < 0.05, 0.01 and 0.001, respectively.

3. Results

In total, 1328 yields were available. Digestate fertilisation increased wheat grain yields very highly significantly by 50%, with average yields over all CRs and replicates of 4.59 t ha⁻¹ a⁻¹, 95% CI [4.50, 4.68] in the unfertilised plots and 6.89 t ha⁻¹ a⁻¹, 95% CI [6.77, 7.00] in the fertilised plots. Wheat grain N content was also significantly (p < 0.05) higher in fertilised plots (2.01%, 95% CI [1.99, 2.03]) than in unfertilised plots (1.83%, 95% CI [1.81, 1.84]).

N supplied by digestate was a good predictor of yield for the fertilised plots ($R^2 = 0.87$ ***, Figure 1). For example, yields were 23% higher in CR6, which received, on average, 202 kg ha⁻¹ a⁻¹ of digestate N, compared with CR2, which received 138 kg ha⁻¹ a⁻¹ of digestate N. This was an effect of the fertilisation and not of the CR because, in the unfertilised plots, yields were highest in CR1b (4.93 t ha⁻¹ a⁻¹, 95% CI [4.63, 5.23]) and lowest in CR2 (3.89 t ha⁻¹ a⁻¹, 95% CI [3.61, 4.16]), a difference of 27%.



Figure 1. Wheat grain yield of fertilised plots in relation to N supplied by digestate, average for each crop rotation (n = 12; 10 different crop rotations with three replicates of CR1; CR1 a, b, and c are indicated by their letters). Wheat yield and N supply are mean values of four replicates for the entire experimental period. The line is a regression ($y = 2.00 + 0.04 \times -6.31 \times 10^{-5} \times 2^2$; R² = 0.87 ***, n = 12).

Yields in the fertilised plots showed a pronounced spatial pattern, with yields higher in CRs in the middle of the field trial than at either end (Figure 2). There was, however, also a positive relationship between field position and digestate N supply ($R^2 = 0.44 *$), indicating that those CRs receiving higher amounts of digestate had been placed more towards one end of the field. This was mainly due to the high N supply of over 300 kg ha⁻¹ a⁻¹ in positions 10 and 11. Excluding these two CRs meant the relationship with position was no longer significant (R^2 decreased to 0.32). The relationship between position and N supply could not, however, explain why yields were higher in the fertilised plots of CR1b (7.23 t ha⁻¹ a⁻¹, 95% CI [6.86, 7.60]) situated in the middle of the experiment than in identically fertilised 1c (6.77 t ha⁻¹ a⁻¹, 95% CI [6.43, 7.10]) and 1a (6.55 t ha⁻¹ a⁻¹, 95% CI [6.02, 7.08]) situated at either end of the field trial (see also Figure 1). Further, the positive relationship between field position and digestate supply did not explain why the unfertilised plots also had the same spatial pattern (CR1b: 4.93 t ha⁻¹ a⁻¹, 95% CI [4.63, 5.23], CR1c: 4.43 t ha⁻¹ a⁻¹, 95% CI [4.09, 4.77], CR1a: 4.00 t ha⁻¹ a⁻¹, 95% CI [3.58, 4.42]). Regression analysis showed that field position was a good predictor of the mean wheat yield, particularly for the unfertilised plots ($R^2 = 0.89$ *** unfertilised, 0.60 * fertilised; n = 12 in both cases; Figure 2) and that the relationship with field position was very similar for both the unfertilised and fertilised plots, differing mainly in terms of the intercept (unfertilised: $y = 3.48 + 0.36 \times -0.02 \times^2$, fertilised: $y = 5.70 + 0.30 \times -0.01 \times^2$).



Figure 2. Wheat yield in fertilised (black circles) and unfertilised plots (grey triangles) in relation to the position of the crop rotation within the experimental area, mean for four replicates for the experimental period. Lines are second-degree polynomial regressions ($R^2 = 0.60$ * for the fertilised plots and $R^2 = 0.89$ *** for the unfertilised plots). The number next to a point is the mean N supplied by digestate (kg ha⁻¹ a⁻¹, fertilised plots only). Field position is in the same order as crop rotation in Table 1.

We used our NH₃ volatilisation model to test if this spatial pattern had been induced by the experiment itself, using the yield and N supply means for each plot. Yields in the unfertilised plots correlated very highly significantly with the N supplied by neighbouring fertilised plots ($R^2 = 0.25$ ***, n = 192). Yields in the fertilised plots also correlated very highly significantly with N supply when losses and gains according to our model were taken into account ($R^2 = 0.31$ ***, n = 192). Combining both groups yielded $R^2 = 0.66$ *** (n = 384, Figure 3a). Importantly, yields from both fertilised and unfertilised plots showed practically the same response to N supply calculated using our model, even though the slope for the fertilised plots was slightly steeper than the overall slope. Yield response in the unfertilised plots was linear, whereas the overall yield and yield in the fertilised plots had a logarithmic relationship with N supply. This implied that the yield response declined with increasing N supply. Up to an N supply of approximately 200 kg ha⁻¹ a⁻¹, the mean yield response was about 15 kg kg⁻¹, whereas the yield response was only about 5 kg kg⁻¹ for an N supply between 200 to 400 kg ha⁻¹ a⁻¹. This logarithmic relationship was also evident for different years using our model, even though yield levels in these years varied. For example, 2014 was a high-yielding year (mean yield for all plots, fertilised and unfertilised: 8.36 t ha⁻¹ a⁻¹, 95% CI [7.93, 8.80]), and 2010 was a low-yielding year (mean yield for all plots, fertilised and unfertilised: $5.75 \text{ t} \text{ ha}^{-1} \text{ a}^{-1}$, 95% CI [5.51, 5.98]). Nevertheless, the yield-N supply curves for both years show a logarithmic relationship (Figure 3b). For 2010, the regression line had the equation y = ln(x) + 1.04 ($R^2 = 0.69$ ***, n = 96), and for 2014, the regression line had the equation y = 2.01 ln(x) - 0.64 ($R^2 = 0.86$ ***, n = 95).



Figure 3. Wheat grain yield in relation to N supply assuming 50% NH₃ loss from the fertilised plots to their neighbour plots. All lines are logarithmic regressions (for details of the equations, see text). The x-axis is square-root scaled to improve resolution at low N supply levels. (**a**) Yield of all 384 plots averaged over three or four years (solid line; $R^2 = 0.66$ ***). Fertilised plots are black open circles (upper dashed line: n = 192; $R^2 = 0.31$ ***) and unfertilised plots are grey open triangles (lower dashed line: n = 192; $R^2 = 0.25$ ***); (**b**) Comparison of a high-yield year (2014; solid grey triangles; n = 96, $R^2 = 0.86$ ***) and a low-yield year (2010; solid black circles; n = 95; $R^2 = 0.69$ ***).

Growing conditions in each year had a large influence on yield. This was mainly due to precipitation and, as a result, R^2 dropped from 0.66 to 0.46 when data from individual years were used instead of multi-year averages (while n increased from 384 to 1221). Precipitation from April to July, the main wheat growth period, ranged from 260 mm (2015) to 464 mm (2021) and was the best simple parameter describing growing conditions. Yields increased linearly with precipitation from April to July for both the fertilised and the unfertilised plots ($R^2 = 0.02$ *** for n = 1317). The best combination of the influence of precipitation and N supply was hence given by $Y = -1.70 + 0.01 \times P + 1.29 \ln(N)$ ($R^2 = 0.49$ ***, n = 1221), where Y is wheat yield in t ha⁻¹ a⁻¹, P is precipitation from April to July in mm and N is N supply in kg ha⁻¹ a⁻¹ (see Figure A5 in the Appendix A for a depiction of the regression). Omitting the year 2012, during which a different, low-yielding variety was grown, increased R^2 to 0.54 *** while n decreased to 1125 with little change in the equation. The multiple regression indicated that the effect of N fertilisation was higher during the growing period.

When the fraction of NH₄⁺-N in the digestate transferred to neighbour plots was varied, the highest correlation and a very highly significantly steeper slope of response

were found when a loss of 57% was assumed. The coefficient of determination, however, was not significantly different from the peak at 57% when losses ranging from about 30% to almost 70% were assumed (Figure 4). It is, therefore, not possible statistically to say which assumption is best within this range. In contrast, assumed losses below 30% and above 70% were significantly inferior. Assuming no NH₃ transfer between plots was the least suitable assumption.



Figure 4. Coefficient of determination between multi-annual mean wheat grain yield and N supply (n = 384) depending on the assumed NH₃ loss of the fertilised plots. The grey line indicates the range where the coefficient of determination is not significantly lower than the maximum at an NH₃ loss of 57%, according to the Hotelling test.

PFS measurements showed that ammonia concentrations peaked the day after digestate fertilisation (Figure 5a). In 2021, this peak was seen at all measurement sites, including at a distance of 130 m from the fertilisation site (Figure 5a). In 2022, this peak was seen in both measurement directions (Figure 5b). Ammonia concentrations in fertilised plots (distance = 0) were as high as 40.4 ppm. However, the highest ammonia concentration of 58.0 ppm was measured in a plot directly neighbouring a fertilised plot (position C4, 15 March 2022). Ammonia concentrations were highest in rows C and D, while row A had the lowest ammonia concentrations.

Ammonia measurements using the DTM method showed that the fertilised plots emitted between 24.5 and 37.4 kg ha⁻¹ of ammonia during the measurement period (Figure 6; up to 124 h after fertilisation in 2021, 173 h after fertilisation in the first measurement in 2022, and 150 h in the second measurement in 2022). In 2021, the mean cumulative ammonia emissions 124 h after fertilisation were 33.9 kg ha⁻¹ (n = 6), 56% of the NH₄⁺-N applied to the plots with the digestate. In 2022, mean cumulative ammonia emissions were, on average, 53% of the NH₄⁺-N that had been applied with the digestate. Even though the measurement period was much shorter in 2021 than in 2022, cumulative losses were higher in 2021 than in 2022 (Figure 6). The average temperature during the measurement periods in 2022.



Figure 5. Average ammonia concentrations measured using the PFS method. Distance is the distance in metres and direction along the field of a PFS measurement site from the fertilised plots (n = 4, NE = northeast, SW = southwest). (a) Ammonia concentrations in 2021. Fertilisation with digestate took place on 31 March 2021; (b) Ammonia concentrations in 2022. Fertilisation with digestate took place on 14 March 2022.



Figure 6. Cumulative ammonia losses measured in fertilised plots using the DTM method. Fertilisation with digestate took place on 29 April 2021, 14 March 2022 and 28 April 2022 (n = 6).

The emissions from the fertilised plots increased the background concentrations above the unfertilised plots. For example, after fertilisation in April 2021, the NH₃ concentrations in the fertilised plots decreased, with a half-life of 19.8 h from 5 ppm 2.5 h after the fertilisation event to 0.3 ppm 124 h after the event ($y = 6.7 \exp(-0.035 t)$; n = 54; $R^2 = 0.78$). This caused the background NH₃ concentrations, as measured above the unfertilised plots, to be as high as 0.25 ppm after the fertilisation event and to fall below the detection limit after 21 h ($y = 0.3 \exp(-0.065 t)$; n = 6; $R^2 = 0.99$; half-life 10.7 h).

The isotope measurements showed that the grass biomass in the fertilised clover–grass plot had a δ^{15} N value of 6.20‰ and was, therefore, enriched in ¹⁵N compared to grass in the unfertilised clover grass plots (Table A2). ¹⁵N in grass biomass of the unfertilised plots increased with increasing distance from a fertilised clover–grass plot (Table A2), indicating that the plots further away from the fertilised plot took up less gaseous NH₃ than the plots closer to the fertilised plot.

The fraction of total N content in grass biomass originating from NH_3 emissions was calculated to be as high as 0.068 (6.8%) in the plot closest to the fertilised plot (Table A2). This fraction decreased with increasing distance from the fertilised plot. Based

on these proportions, the plot closest to the fertilised plot was calculated to have taken up $30 \text{ kg ha}^{-1} \text{ a}^{-1}$ from NH₃ emitted from neighbouring fertilised plots (Figure 7).



Figure 7. Estimate of gaseous NH₃ uptake (kg ha⁻¹ a⁻¹) in five clover–grass plots depending on their distance from a fertilised clover–grass plot, based on ¹⁵N isotope measurements. The line is a linear regression ($R^2 = 0.78$ *).

4. Discussion

As expected, digestate fertilisation increased wheat yields and N content when compared with an unfertilised control in accordance with the literature (e.g., [21]), but there was a pronounced spatial pattern in both the fertilised and the unfertilised plots. As soil analyses showed no marked differences in soil properties such as Norg content and field capacity across the site [35], the large number of plots and years and the close correlation over all treatments, especially within the unfertilised plots, leaves little doubt that N transfer between neighbouring plots occurred. Our ammonia emission measurements supported this premise, showing that approximately 55% of the NH_4^+ -N that had been applied with the digestate in our experiment was volatilised within a few days, with these ammonia emissions being transported at least as far as 130 m from the original site of fertilisation, and in different directions along the experimental field. Ammonia concentrations were highest at or in the immediate vicinity of the fertilisation site. This was reflected by the 15 N isotope analysis, where δ^{15} N was highest in grass biomass in the fertilised plot and lowest in grass biomass in the plots neighbouring the fertilised plot, increasing again in plots further away from the fertilised plot. This indicated that the grass biomass in the fertilised plot had taken up NH4⁺-N directly from the digestate. In contrast, grass biomass in the unfertilised plots absorbed ¹⁵N-depleted NH₃-N volatilised from the digestate, with grass biomass in plots neighbouring the fertilised plot absorbing more than grass biomass in plots further away from the fertilised plot. We expect that isotope measurements at greater distances from the site of fertilisation would also indicate these effects. The size of these effects will depend on the prevailing wind direction and wind speed at the site. For example, Boaretto et al. [36] observed variation in the absorption of ¹⁵NH₃ in citrus trees equidistant from the fertilisation site, presumably due to wind direction. In an orchard protected from the wind, coffee plants reabsorbed 43% of the volatilised NH₃-N [37].

In principle, processes other than NH_3 volatilisation could have contributed to the spatial pattern seen in the experiment. In particular, N in subsurface water flow may be important [10,38,39]. This would, however, only have caused an N transfer downslope while the plots upslope would not have been affected. This would have split the unfertilised plots into two groups, one group with consistently low yields, even in cases where our

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simple modelling assumed large N transfers from neighbouring plots, while the other group should have responded to digestate application in the fertilised plot in the upslope position. Such a split in the data was not evident. Furthermore, NH₃ volatilisation can explain all the effects that we see in our study. In particular, this explained why CR1a and c, situated at the outer edges of the experiment, had lower yields than CR1b, despite identical treatment. It even explained why CR1c, located next to a CR receiving the highest amounts of digestate-N, had a higher yield than CR1a, which was situated next to CRs receiving the lowest amounts of digestate-N. It explained why, for both the fertilised and the unfertilised plots, the relationship of yield with position was a parabola with the highest yields in the positions in the middle where NH₃ import from both sides was possible, whereas the lowest yields occurred at both edges where import was possible only from one side. N balances calculated for the unfertilised plots show large N deficits [40] without a correspondingly large decrease in yields with time. N transfer via NH₃ volatilisation would explain this effect.

Plot experiments usually aim at minimizing soil heterogeneity to reduce statistical error. This can best be achieved by using small plots and situating treatments in close proximity to each other. In addition, strong gradients between treatments are desirable for producing detectable effects. Both experimental design strategies become undesirable if the NH₃ flow between treatments should be kept at a minimum. The inference from our experiment is that such experiments should use much larger plots with a larger area-to-circumference ratio, and treatments should be separated by larger distances. The average plot size in wheat fertiliser trials is 37.2 m² [41], which is only half the size of the plots in our experiment. The effects would, therefore, normally be even more pronounced. A completely different experimental layout to what is traditionally used would also avoid the inherent problem with traditional designs, which is that replicates are actually pseudoreplicates [42] because they all investigate the same conditions, i.e., soil, weather and microclimate, and, thus, generalization of the results to a wider variety of settings is not possible.

The mean field size in the region where the experiment was situated is 5.7 ha [43], which is three orders of magnitude larger than the typical plot size or the plot size in our experiment. Therefore, the question arises as to which group of plots was more affected by the experimental design, fertilised or unfertilised, and, thus, which result differs more from the response that would occur in farmers' fields. The very flat increase in yield with increasing N in the fertilised plots means that in large fields, the increase in yield may be higher if losses are lower because of a large area-to-circumference ratio. This effect, however, may not be large because there would be no NH₃ sink in the field if the entire crop has reached the stomatal ammonia compensation point after a fertilisation event. Long-distance losses would then result. The steep gradient of the curve for the unfertilised plots indicates that they profit strongly from their fertilised neighbours, but only if such neighbours exist. In farmers' fields, this may only be true at the field edge. In addition, the timing may be less favourable than in our experiment, where a wheat plot was situated beside a wheat plot, and thus emissions from neighbouring plots occurred at a time when a large fertilisation effect for wheat was expected. Such a perfect match will not always be the case in reality. For both reasons, the trapping effect of unfertilised fields will also be smaller in reality, which again would favour long-distance losses. It is, thus, unfortunate that there is a strong trend in modern agriculture of increasing field sizes [44–46], which will increase regional-scale NH₃ emissions and decrease the applicability of results from small-plot experiments. An N surplus causes a shift in δ^{15} N at the farm level, indicative of emissions at a regional scale, rather than field-to-field transfer of NH₃ [47,48].

An even more important question is which measures could, therefore, be taken in agriculture to reduce NH_3 losses. The estimated loss of 57% that performed best in our model was corroborated by our ammonia emission measurements and was similar to the German emission factor for NH_3 for the application of cattle slurry and digestate derived from cattle slurry using trailing hoses without incorporation (46%) [49], and is, therefore, not unrealistic. The reasoning above means that losses would also occur in large plots

and hence, also in fields. We found a consistent effect in many plots and over many years that was even more pronounced in dry years than in wet years. This makes it likely that it will occur under many conditions. Such a consistent effect would not be very likely if it were only caused by the conditions during fertiliser application, which must have varied considerably over the long experimental period. In addition, an N supply above 200 kg ha⁻¹ had barely any effect on yields, with our data indicating that at N supply levels above 280 kg ha⁻¹, wheat yields may start to decrease. Other authors have found similar effects in wheat [50]. This means that the high N inputs could have increased the apoplastic NH₄⁺ concentration in the wheat crop, thereby increasing the NH₃ compensation point and causing NH_3 emission [15,51]. Therefore, even incorporation of the digestate into the soil would not prevent NH₃ losses to the environment, and the same amount of mineral fertiliser could also induce foliar NH₃ losses [52–54]. Splitting fertilisation into two applications would also not have a large effect, as indicated by the flat relationship between N supply and yield. It appears that the best way of reducing NH₃ losses would be a slow-release N source that does not cause peaks in the stomatal NH3 compensation point. Easily degradable carbon compounds are decomposed during the anaerobic digestion process [4]; thus, liquid animal slurries contain more organic matter than digestates and should deliver N more slowly than digestates. Solid manures should be even better in this regard than liquid manures. In the short term, crop N utilization of solid manures and the resulting yields are often the same or lower than for liquid manures or mineral N fertilisers [55,56]. However, in the long term, solid manures show a substantial increase in N utilization [57]. For example, Paul and Beauchamp [58] found that the N availability from composted manure was as high during the third year after a one-off application as in the first year. While these results are often attributed to improvements in soil structure and increases in organic matter stimulating soil fertility over time [55], our results indicate that higher N-use efficiencies may also be due to lower foliar N losses with slow-release N fertilisers. Möller and Stinner [4], for example, found that ammonia emissions were lower after the application of solid farmyard manure than after the application of slurry or digestate. However, total canopy NH₃ emission or deposition depends not only on emission/deposition fluxes from plant stomata and cuticles but also on fluxes from soil and litter and the interactions between these different components and with atmospheric NH_3 concentrations [59]. Soil emission potentials are significantly larger than the stomatal emission potentials, and the soil is, therefore, likely to have the biggest influence on total canopy emissions [60,61].

Our simple model, which explained all data well irrespective of whether we looked at high-yield years or low-yield years or whether we looked at fertilised or unfertilised plots, implies that the availability and the effects on yield are identical for NH₃-N and for other, mainly organic N sources, in the digestate. The relationships described above would disappear if we assumed only NH₃-N or only organic N was available and responsible for our results. It is generally assumed that mineral N (NO₃⁻ and NH₄⁺ ions) is immediately available for plant uptake, whereas organic matter first has to be mineralised before organic N pools can be used by plants [62–64]. The apparently high availability and effectiveness of the organic N were not due to an accumulation in the soil organic matter pool and subsequent slow release, which is likely to occur in a long-term experiment. If this were the case, the effect of the organic N should have increased with time. This was not evident in our field trial. In particular, in the comparison of yields from 2010 (four years after the first fertilisation) and 2014, there was no offset of the plots receiving organic N from the digestate that could be attributed to increased N release from a pool of soil organic N that had accumulated over time. The differences between years were only caused by the different weather conditions and appeared in both the plots that had received organic N and the plots that did not receive organic N. In the long term, N losses in this field trial may increase as N in the soil organic matter pool reaches a stable level and no longer accumulates, but N is still added via digestate fertilisation. This is particularly the case for CRs 9 and 10, both CRs with a high proportion of clover–grass, and, therefore, additional N fixation, that have built up large soil organic matter stocks over the experimental period [65,66].

In our experiment, the decreasing effectiveness of N with increasing N supply indicated that losses of NH₃ (presumably mainly long-distance aerial transport) and organic N (presumably mainly leaching and denitrification losses) were similar for both N sources. This may not be the case at other sites. Soil type may have an effect, for example. In particular, in a considerably drier and warmer climate, NH₃ losses would probably have been higher, and leaching and denitrification losses would probably have been lower. The net effect, however, should be low because both changes cancel each other out.

5. Conclusions

Analysing more than a decade of yields on 384 plots and using a simple N loss/gain model enabled us to see how NH₃ emissions from plots fertilised with biogas digestate affected yields in neighbouring unfertilised plots and to understand the availability of N from biogas digestate. Ammonia volatilisation measurements and ¹⁵N isotope analysis to investigate shoot N absorption validated the results of the model. Our results have important implications, not only for the interpretation of plot experiments but also for agricultural practise. Our results indicate that in order to avoid inadvertently affecting unfertilised control plots, plot sizes need to be increased and treatments situated further apart. Foliar NH_3 emissions and, therefore, agricultural ammonia emissions in general may be reduced if solid rather than liquid organic manures are used; however, the effect of solid manures on the ammonia compensation point of crops needs further investigation. Further investigation in general is needed on how organic fertilisers affect the various components contributing to the overall emission or deposition of NH_3 in agricultural crops. It appears that the organic N in the biogas digestate used in our field trial was highly crop-available, and therefore, there was also a high risk of N losses. Further research on the availability of organic N from digestates is needed to confirm our findings.

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Appendix A

Figure A1. Plot and ammonia volatilisation measurement plan 2021. Row refers to the PFS measurement row, distance is the distance in metres and the direction along the field of a PFS measurement site from the fertilised plots (NE = northeast), red circles indicate the location of the PFS measurement sites, X marks the plots that were fertilised with digestate on 31 March 2021 during PFS measurement. DTM measurements took place in the plots coloured purple.

Table A1. Weather conditions during ammonia measurements in 2021 and 2022.

Date	Mean Temperature (Daily, °C)	Mean Temperature (During DTM Measurement, °C)	Mean Wind Speed (Daily, m s ⁻¹)	Mean Wind Speed (During DTM Measurement, m s ⁻¹)	Wind Direction	Mean Pressure (During DTM Measurement, hPa)
30 March 2021	12.7		0.9		ESE	
31 March 2021	13.4		1		SSE	
1 April 2021	13.7		2		W	
2 April 2021	8.3		3		NW	
3 April 2021	4.4		2.8		NW	
4 April 2021	5.2		1.5		WNW	
5 April 2021	5.3		4.3		SW	
6 April 2021	-0.8		3.3		WSW	
29 April 2021		12.9		2.5		1007.4
30 April 2021		14.1		3.3		1008.4
1 May 2021		10.5		2.8		1009.1
2 May 2021		6.3		4.1		1017.6

Date	Mean Temperature (Daily, °C)	Mean Temperature (During DTM Measurement, °C)	Mean Wind Speed (Daily, m s ⁻¹)	Mean Wind Speed (During DTM Measurement, m s ⁻¹)	Wind Direction	Mean Pressure (During DTM Measurement, hPa)
3 May 2021		10.0		3.1		1021.6
4 May 2021		16.0		4.9		1007.7
14 March 2022	8.2	12.1	2.3	2.1	WSW	1028
15 March 2022	7.1	8.9	1.7	1.1	ENE	1026.0
16 March 2022	7.1	8.9	2.8	2.6	Е	1026.4
17 March 2022	6.1	7.9	1.8	1.3	E	1027.2
18 March 2022	7.5	10.0	3.2	3.4	NE	1036.9
19 March 2022	5.5	8.0	3.5	3.6	NNE	1033.7
20 March 2022	6.7	10.7	2.9	3.3	E	1031.0
21 March 2022	6.3	9.7	1.6	1.8	ENE	1032.8
28 April 2022		14.3		2.3		1027.2
29 April 2022		16.1		2.6		1024.1
30 April 2022		12.2		2.5		1020.7
1 May 2022		10.5		1.7		1020.4
2 May 2022		1.7		15.6		1014.9
3 May 2022		2.0		16.3		1013.7
4 May 2022		2.1		16.6		1015.3





Figure A2. Plot and ammonia volatilisation measurement plan 2022. Row refers to the PFS measurement row, distance is the distance in metres and direction along the field of a PFS measurement site from the fertilised plots (NE = northeast, SW = southwest), red circles indicate the location of the PFS measurement sites, X marks the plots that were fertilised with digestate on 14 March 2022 during PFS measurement. DTM measurements took place in the plots coloured purple.



Figure A3. Ammonia measurement using the DTM method in a fertilised wheat plot.



Figure A4. Plan of clover-grass plots sampled for isotope analysis on 29 September 2014.



Figure A5. Multiple regression of wheat grain yield in relation to N supply and precipitation during the main growing period ($R^2 = 0.48$ *** for n = 1221).

Table A2. δ^{15} N in clover–grass sampled on 29 September 2014.

Distance across the Field from Fertilised Clover–Grass Plot (m)	δ ¹⁵ N Grass Component (‰)	Fraction of Total N in Grass Component Derived from Gaseous NH3 Emissions
3	1.35	0.068
9	1.53	0.060
9	2.24	0.029
15	2.16	0.032
16.5	2.90	0

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