

Review

Recent Advances in Copper-Based Organic Complexes and Nanoparticles for Tumor Theranostics

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Abstract: Treatment of drug-resistant forms of cancer requires consideration of their hallmark features, such as abnormal cell death mechanisms or mutations in drug-responding molecular pathways. Malignant cells differ from their normal counterparts in numerous aspects, including copper metabolism. Intracellular copper levels are elevated in various cancer types, and this phenomenon could be employed for the development of novel oncotherapeutic approaches. Copper maintains the cell oxidation levels, regulates the protein activity and metabolism, and is involved in inflammation. Various copper-based compounds, such as nanoparticles or metal-based organic complexes, show specific activity against cancer cells according to preclinical studies. Herein, we summarize the major principles of copper metabolism in cancer cells and its potential in cancer theranostics.

Keywords: copper; organic complexes; nanoparticles; tumor theranostics

1. Introduction

Copper is a transition metal that plays several important roles crucial for maintenance of cell homeostasis, regulation of cell growth and proliferation, and iron metabolism [1]. Various roles of copper are explained by its ability to act as either a recipient or a donor of electrons depending on the oxidation state: Cu¹⁺ (cuprous ion) and Cu²⁺ (cupric ion). The oxidation state also affects the copper interaction with organic compounds. Thus, Cu¹⁺ preferentially binds to the thiol group in cysteine or the thioether group in methionine, while Cu²⁺ exhibits a high affinity for the secondary carboxyl group in aspartic/glutamic acid or the imidazole nitrogen group in histidine. As a result, copper ions readily form complexes with biomolecules containing these amino acid residues. Copper atoms are involved in a functioning of a wide spectrum of proteins, such as copper/zinc superoxide dismutase (Cu/Zn SOD or SOD1) [2], cytochrome c oxidase (COX) [3], lysyl oxidase (LOX) [4], mitogen-activated protein kinase MEK1 [5], and cAMP-degrading phosphodiesterase PDE3B [6]. In these proteins, copper ions participate in diverse biochemical reactions (especially redox reactions) of donating or accepting of electrons and maintain specific protein structures by coordinating with the abovementioned groups.

Despite its important physiological role, free copper ions are able to damage DNA and protein molecules via generation of reactive oxygen species (ROS) and interaction with cysteine and methionine residues [7]. That is why each cell and whole organisms have distinct mechanisms for the regulation of copper absorbance, distribution, accumulation, and excretion. With the development and propagation of copper-based pharmaceuticals, it is crucial to consider these metabolic and regulatory pathways to improve biocompatibility and efficacy of such compounds. For now, only a small number of studies dedicated to the design of novel copper-containing compounds consider underlying molecular mechanisms of intracellular copper regulation. The present work aims to provide a holistic view of the problem to help researchers boost their work and realize rational approaches in drug development.

2. Copper Intake, Distribution, and Efflux in Normal and Tumor Cells

The major proteins involved in copper maintenance include: CTR1 (copper transport protein), which is responsible for copper intake either from the intestine or blood; metallothioneins and metallothioneins, including ceruloplasmin, which are responsible for metal sequestration, distribution in organisms, and transport to various proteins; ATP7A and ATP7B (ATP-ase copper transporter alpha) responsible for copper excretion via membrane efflux or Golgi apparatus [8]. All these proteins have cysteine- or methionine-rich domains responsible for the binding. A precise description of proteins involved in copper homeostasis and a comparison of copper metabolism in normal and cancer cells are given below.

As it has previously been mentioned, copper intracellular metabolism is precisely regulated by specific protein machinery, which prevents the generation of free copper ions in the cytoplasm or extracellular space and ion-mediated toxicity (Figure 1). CTR1 is a major protein responsible for copper uptake in eukaryotes. CTR1 transporter acts as a pump that facilitates copper import without ATP consumption [9]. The rate of the copper intracellular transport depends on the copper concentration, the presence of other ions (Fe^{3+} , Zn^{2+} , Ag^{+}) and organic compounds (e.g., ascorbate), cell type, and pH. The structure of homotrimeric CTR1 protein contains methionine gates for selective bypass of monovalent copper ions exclusively. However, isoelectric silver ions can compete with copper decreasing its intracellular content [10]. As only monovalent copper can be transported by the CTR1 protein, bivalent copper should first be restored to the monovalent state. This process is facilitated by the reductase proteins, such as STEAP, which are also reported to be overexpressed in several types of cancers and involved in tumorigenesis [11].

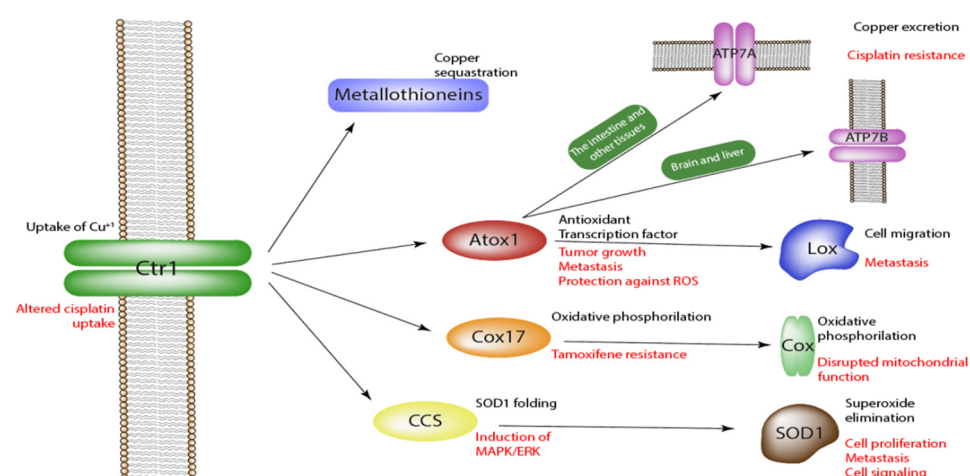


Figure 1. Proteins of copper metabolism. The arrows show how copper is metabolized. In general, CTR1 is responsible for copper intake; numerous metallothioneins and chaperones store the metal and deliver it to the active sites of functional proteins. The black font indicates the role of a protein in normal cells, and the red font indicates the protein function in cancer.

After crossing the plasma membrane, copper ions are readily sequestered by the numerous intracellular metallothioneins, metallochaperones, albumins, glutathione, and ceruloplasmin [12]. Some of these proteins can store the metal for further use, while others serve for intracellular transportation of copper. For example, metallochaperones transfer copper to the active centers of the certain proteins or buffer the metal for further use. Copper chaperon for superoxide dismutase (CCS) delivers copper to superoxide dismutase (SOD1) enzyme, which converts superoxide radical into hydrogen peroxide and oxygen [13,14]. COX17 is another metallochaperone responsible for copper transportation to COX, an important protein involved in oxidative phosphorylation [15].

Atox-1 is a transport protein that delivers copper to ATP7A and ATP7B, which are responsible for copper release into the blood or bile, respectively [16]. Additionally, ATOX1 can migrate to the cell nucleus and act as a transcription factor facilitating cell growth, proliferation, and migration. Another common localization of ATOX1 is in proximity to the plasma membrane, where copper can be transferred to the membrane-associated proteins, such as lysyl oxidase, which is involved in cell migration [16].

3. Copper Regulation in Cancer

Many proteins required for copper metabolism are known to be overexpressed or malfunctioned in cancer cell metabolism. The most known example is participation of these proteins in chemotherapeutic response to conventional drug cisplatin. There is much evidence about CTR1 involvement in the transportation of cisplatin [17,18]. Meta-analysis of gene expression in various cancer types revealed that the reduced expression of the CTR1 gene is associated with the development of cisplatin resistance [19]. The knockout of CTR1 and DMT1 (divalent metal transporter 1) in human H1299 non-small cell lung cancer cells leads to pronounced cisplatin resistance. Moreover, the CTR1 loss decreases expression of COMMD1, XIAP, and NF- κ B, which have a distinct influence on the intracellular homeostasis and signaling [20]. Several works of various research groups also proved a hypothesis about involvement of CTR1 and ATOX1 in cisplatin transport and sequestration [21–23]. However, another study on HEK-293T cells provided evidence about the modest participation of copper-binding proteins (i.e., CTR1, CTR2, ATOX1, and CCS) in cisplatin uptake and distribution [24].

At the same time, a connection between high ATOX1 expression level and survival rate in primary tumor biopsies has been found. Analysis of transcription profiling of 1904 breast cancer patients on METABRIC data set suggests that overexpression of Atox1 may serve as a marker for breast cancer prognosis [25] but only in the hormone receptor-positive tumors. Considering copper involvement in the functioning of the LOX protein [26] which is responsible for cell migration, ATOX1 may facilitate the function of LOX enhancing tumor ability for metastasis [27]. Moreover, Atox1 is also involved in transcription regulation of several genes, as was mentioned earlier. First, upon copper binding ATOX1 can migrate to the cell nucleus and bind the cis element of the cyclin D1 promoter, thus stimulating cell growth and proliferation [21]. Furthermore, a more complex interplay between ATOX1 and p53 has been found [28]. Authors observed increased copper amounts in cell nuclei for HCT116 p53^{+/+} cells compared to p53^{-/-} cells. These facts suggest that Atox1 may play a significant role in cell signaling and regulation of gene expression which should be determined in future studies.

Cytochrome c oxidase copper chaperone (COX17) is also involved in cancer. Inhibiting COX17 in acute leukemia cells results in decreased adenosylhomocysteinase activity leading to disruption of DNA methylation and changes in cell epigenetics [29]. The link between COX17 and cisplatin distribution to mitochondria has been found [30]. The involvement of copper-binding proteins in cisplatin uptake and distribution is probably connected to the similarities in binding affinity of platinum and copper ions. Moreover, glutathione (GSH) seems to attenuate this effect. It was found that 90% of cisplatin bound to GSH is readily transferred to COX17 [31]. This suggests probable involvement of thiol-containing molecules and not only proteins in intracellular cisplatin distribution. It would

be interesting to investigate the effects of combining treatment with cisplatin and thiols or cisplatin-thiol complexes or nanostructures. COX17 was also studied as a prognostic marker for prediction of tamoxifen resistance in breast cancer patients [32]. The authors reported that this protein could be employed as a predictive marker for tumor recurrence and metastasis. These features are also observed for COX5B which is a subunit of COX itself [33]. This correlates with the prognostic value of ATOX1 which was found to possess similar properties in the breast cancer. Another COX nuclear-encoded subunit, COX4, is also shown to be a valuable prognostic and therapeutic marker for medullary thyroid cancer treatment [34]. The role of the COX protein in cancer development and progression as well as its influence on altered signaling and metabolic pathways needs to be further explored.

CCS, a protein involved in copper delivery to SOD1, is also involved in tumorigenesis. SOD1 could serve as a prognostic marker which contributes to worsened prognosis and higher risk of gastric [35] and prostate [36] cancer. Another study indicates SOD1 involvement in cell proliferation and metastasis in non-small cell lung cancer [37]. At the same time, knockdown of CCS leads to decreased cell proliferation and migration of MDA-MB-231 cells but does not affect the MCF-7 cell line [38]. In addition, the MAPK/ERK pathway was inhibited upon loss of CCS activity in MDA-MB-231 cells which also correlated to the increased ROS formation. Inhibition of CCS and Atox1 with specifically designed small molecules is a promising treatment strategy with reduced side effects [39]. The expression of CCS was found to be decreased in human hepatocellular carcinoma (HCC) which is distinct from breast cancer [40]. Despite a statistical significance not being achieved, the study concluded that a low expression level of CCS is a negative prognostic marker for HCC patients. Presumably, copper trafficking in various tissues could be different, as well as the involvement of copper-binding proteins in cancer development, progression, and metastasis. This provides a foundation for further investigation on a wide panel of cancer cell lines.

Copper efflux proteins, ATP7A and ATP7B, are also involved in cancer progression. ATP7A correlates with a poor survival rate and is overexpressed in several tumor types, such as breast, lung, prostate, ovarian, and colon cancer [41]. Another study shows that ATP7A is associated with cisplatin resistance in ovarian cancer and influence effectiveness of treatment with tetrathiomolybdate, which inhibits ATP7A activity [42]. Decreased sequestration of platinum leads to its accumulation in the cell nucleus with subsequent DNA damage. Moreover, the application of tetrathiomolybdate can also result in Ctr1 high expression increasing cisplatin uptake that may be used as a solution for treatment of drug resistance tumors [43]. Another study suggests a greater impact from inhibiting ATP7B compared to ATP7A [44]. A detailed analysis of the ATP7A and ATP7B roles in ovarian cancer are discussed in the review [45]. A study in the breast cancer model reveals the opposite effects of ATP7A and ATP7B in contribution to the cisplatin resistance [46]. ATP7A seems to be more involved in this process, whereas the analysis of ATP7B did not reach statistical significance. To summarize, the above-mentioned ATP7A and ATP7B influence the cisplatin efflux leading to decreased effectiveness of this drug; however, the precise role of each protein should be determined for distinct types of cancer.

Copper takes an active part in the proangiogenic pathways via several mechanisms. First, copper stimulates endothelial cells proliferation and migration. Next, copper is involved in the expression of certain proangiogenic factors (for example, vascular endothelial growth factor VEGF) [47], particularly as a response to hypoxia-inducible factor (HIF-1) signaling [48]. When elevated, copper becomes toxic and may induce side effects leading to genetic disorders (e.g., Wilson's disease) and various types of oncological diseases. However, the exact molecular mechanisms underlying the connection between excessive copper levels and malignant cells are still unknown. It can only be hypothesized, particularly in the early stages, after considering the role copper plays in tumor angiogenesis. Malignant tissues have higher Cu accumulation levels, thus increasing the expression of human copper transporter (hCTR1). hCTR1 regulates the activation of cell-signaling pathways in embryogenesis, which leads to the development and progression of cancers [49].

The above-mentioned impact of copper ions and copper-binding proteins on cell growth, migration, and metabolism suggests that cancer cells require high copper levels to facilitate cell survival and disease progression. Indeed, tumor tissues are enriched with copper suggesting that this metal is one of the diagnostic tools for various oncological disorders [50]. Moreover, copper or copper-binding proteins are essential for the function of important signaling pathways, such as BRAF [51], NF- κ B [52], MAPK [53], and EGFR/Src/VEGF [54]. Hence, the significant role of copper in cancer appearance and progression is starting to emerge in front of researchers. The accumulated data uncover the possibility to improve the efficiency of diagnostic approaches and increase treatment efficacy.

4. Therapeutic Effects of Copper-Based Compounds and Nanocarriers

The disparity in tumor cell and normal cell responses to copper have paved the way for copper complexes to evolve as anticancer agents. Copper-based compounds nowadays are receiving attention due to their target-specific therapeutic properties. Copper compounds influence the activities of several crucial cell organelles, such as the mitochondria and endoplasmic reticulum, leading to the loss of their functions and eventually resulting in cell death (Figure 2).

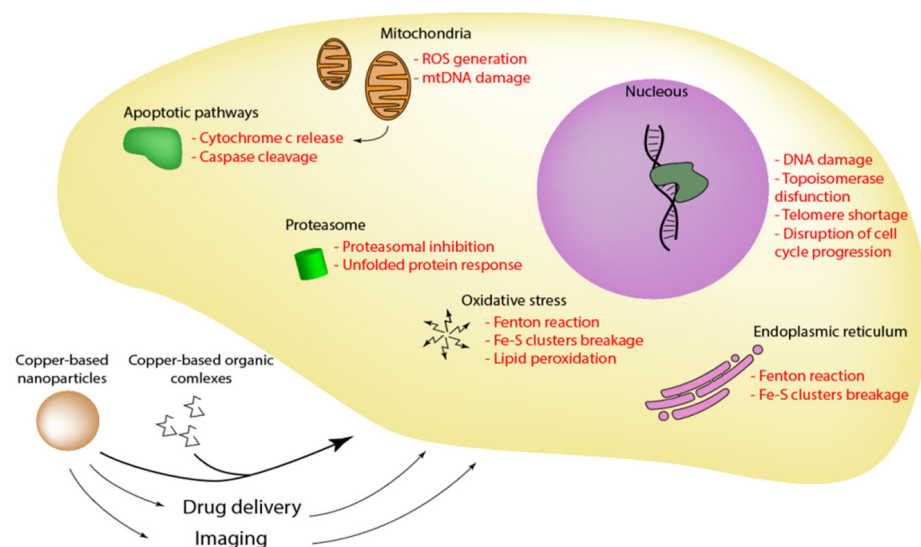


Figure 2. The main effect of copper nanoformulations on cell metabolism. The major impact of copper on cell metabolism is a result of ROS generation and DNA damage. Proteasome, endoplasmic reticulum, and mitochondria also suffer from copper excess.

Nowadays, the increasing number of metal-based compounds and nanoparticles are being investigated due to their promising potential in theranostics, and various iron, zinc, copper-based and other agents are under development and testing for these purposes. For example, superparamagnetic iron oxide nanoparticles (SPIONs) are being actively used as a contrast agent for MRI procedures and in therapy. Currently, there are several running translational studies which explore SPIONs' toxicity and biomedical applications, and ferumoxytol was FDA-approved for clinics [55]. Copper is also attracting the attention of researchers as a possible component for nanocompounds for theranostics and drug delivery. For example, copper is used in PET scanning as a radiotracer agent in cancer diagnostics, and $^{64}\text{CuCl}_2$ has successfully passed clinical studies demonstrating its diagnostic potential [56]. Several studies successfully implemented copper for efficient bone regeneration [57] and anti-inflammatory therapy [58]. Copper-based nanoparticles also found their place in chemodynamic [59] and photothermal therapy [60].

The radiotracer biodistribution has shown that the liver has the highest uptake, followed by the intestine and pancreas, with urinary excretion being insignificant. It is the

first biodistribution and radiation dosimetry trial with healthy volunteers. The estimated absorbance and effective doses were higher than the ones from another report with participants suffering from prostate cancer. The measurement methodology and assumptions used in dose calculation as well as the difference between the biodistribution in cancer patients and healthy volunteers are the main reasons for that disparity [61]. An interesting combination of SPIONs and Cu (II) ions were used as a cell labeling MRI/PET agent. Contrast agents showed good cellular uptake and cell-labeling ability [62]. Furthermore, gold nanoparticles alloyed with copper-64 demonstrate higher sensitivity and stability compared to non-modified gold nanoparticles [63]. Thus, copper presence could improve the effectiveness of the iron or gold nanoparticles, which opened new opportunities for further research in the field of cancer imaging. However, the major limitation and risk factor for wide implication of copper is toxicity of copper ions for cells [64].

Extrinsic and mitochondrial pathways of apoptosis are important in the control of tumor development and could be exploited for therapy [65]. The anticancer properties of Schiff base copper (II) complexes are well-studied and known in the scientific community. For instance, $[\text{Cu}(\text{sal-5-met-L-glu})(\text{H}_2\text{O})]\cdot\text{H}_2\text{O}$, $[\text{Cu}(\text{ethanol})_2(\text{imidazole})_4][\text{Cu}_2(\text{sal-D, L-glu})_2(\text{imidazole})_2]$ and $[\text{Cu}(\text{sal-D, L-glu})(2\text{-methylimidazole})]$ complexes activate the intrinsic pathway, while $[\text{Cu}_2(\text{sal-D, L-glu})_2(\text{isoquinoline})_2]\cdot 2\text{C}_2\text{H}_5\text{OH}$ initiates the extrinsic pathway in human HT-29 colon carcinoma cells, respectively. All these complexes also induce a cytotoxic effect on the HT-20 cell line, and as a result, prove that they might become potential anticancer agents [66]. Structural formulas of the complexes can be found in recent publications [67–69]. Another study shows that accumulation of copper ions inside the cells leads to oxidative stress and apoptosis [70]. Moreover, the usage of 2,2'-dithiodipyridine strongly enhances this effect which is bound to its ability to transport copper through the plasma membrane.

Topoisomerases play an essential role in DNA replication and are relevant in cancer research as a target for novel therapies. There are currently several drugs approved by the FDA targeting topoisomerases (e.g., irinotecan, etoposide, etc.). Thiosemicarbazones are a group of complexes proved to have anticancer activity. "Triapine" (thiosemicarbazone) has been successfully tested for uterine cervix and vaginal cancers in clinical trials phase I and II and is presently under clinical trials phase III [71]. Thiosemicarbazones copper (II) complex $[\text{Cu}(\text{PyCT4BrPh})\text{Cl}]$ was investigated and demonstrated a cytotoxic effect on a leukemia cell line (THP-1) and human breast cancer cell line (MCF-7). It had stronger topoisomerase inhibitor activity and generally more impact on these cell lines than its analogue without copper, which proves how transition metals can increase the effectiveness of the known compound [72].

Copper complexes are shown to influence the endoplasmic reticulum leading to immunogenic cell death in breast cancer stem cells [73–75]. In a recent study, cuprous oxide nanoparticles affect calcium transport leading to its accumulation in intracellular space resulting in oxidative stress, activation of caspases, and apoptosis. Copper complexes are also able to inhibit proteasome function [76]. Other structures allow G-quadruplex telomeric DNA reduction [77]. These effects lead to disturbances in cell cycle, activation of apoptotic pathways, and cancer cell death. One article reports copper complexes are able to accumulate inside mitochondria leading to cytotoxicity by damaging mtDNA [78]. A great variety of induced effects allows copper compounds to be used for various applications in a precisely determined manner of action.

5. Copper Nanoparticles for Cancer Imaging and Drug Delivery

Due to the recent developments in imaging technologies and biology, molecular imaging provides not only the possibility to visualize the tumor, but also to assess the expression and activity of specific molecules (e.g., protein kinases, enzymes, proteases, etc.) and various processes (including metastasis, tumor cell apoptotic death, angiogenesis, etc.) involved in cancer progression, response to therapy, and recurrence [79]. Furthermore, molecular imaging based on CuS NPs enables repetitive assessment of particles biodis-

tribution and biokinetic properties employing positron emission tomography (PET) and photoacoustic imaging (PAI) [80,81].

Photoacoustic (PA) imaging, developed rapidly in the recent decade, represents a noninvasive biomedical imaging method which can be employed for visualization of deeply located tissues tumors, analysis of vasculature [82], or evaluation of neoangiogenesis [83]. Upon the *in vivo* absorbance of a short-pulse laser by various molecules (e.g., water, melanin, RNA, DNA, hemoglobin, cytochromes, lipids, etc.) ultrasonic signals are generated via the mechanism of photothermal conversion [84–86]. Up-to-date gold nanostructures (GNPs) were widely applied as contrast agents for photoacoustic imaging [87]. However, GNPs were reported to have several limitations as contrast agents, including dependence of optical properties on shape, geometry, and size of particles as well as their susceptibility to tumor microenvironmental factors. On the contrary, compared to the maximum absorption between 560 and 840 nm of GNPs, the absorption of copper nanoparticles could be tuned to peak at wavelengths greater than 900 nm, thus providing the improved sensitivity in the NIR region (i.e., stronger PA signal, higher signal-to-noise ratio, greater field-of-view) [88]. Indeed, in the study by Zhou [89] et al., it was shown that polyethylene glycol (PEG)-coated copper(II) sulfide nanoparticles (PEG-CuS NPs) (peak absorption of 1064 nm) could be successfully employed both as a contrast agent for *in vivo* imaging of 4T1 breast tumor vasculature and as a mediator for photothermolysis of cancer cells. However, due to the intrinsic dipole–dipole interactions among Cu-based particles, synthesis of size-tunable, biocompatible, and colloiddally stable suspension of particles remains a challenge. To overcome this problem Ding [90] et al. proposed the aqueous synthesis of PEGylated copper sulfide particles with controllable size between 3 and 7 nm. Subsequent preclinical studies demonstrated that particles, particularly of less than 5 nm, had a higher tumor-imaging potential. Another approach could be based on application of tumor microenvironment-sensitive nanoparticles as was proposed in the work of Wang et al. [91]. The authors developed iron-copper co-doped polyaniline nanoparticles (Fe-Cu@PANI) which upon glutathione (GSH) redox reaction could shift in the absorption spectrum from the visible to the NIR. The etching of Fe-Cu@PANI resulted both in photoacoustic imaging of tumors and efficient photothermal therapy. In recent research by Bindra [92] et al., the authors synthesized a self-assembled nanosystem (SCP-CS) which consisted of a semiconducting polymer (SCP) and encapsulated ultras-small CuS (CS) nanoparticles. This nanosystem demonstrated not only an improved PA-imaging ability but also significant tumor growth inhibition due to the enhanced production of ROS.

In PET apart from traditionally employed positron emitters [⁶⁴Cu]-based NPs were also shown as an efficient radiotracer for tumor diagnostics [93,94]. Thus, Zhou [94] et al. in the U87 human glioblastoma xenograft model demonstrated that a novel class of chelator-free [⁶⁴Cu]CuS nanoparticles (NPs) (PEG-[⁶⁴Cu]CuS NPs) could effectively target the tumor cells providing a potential for image-guided PTA therapy. In a more recent study, more complex indium- and copper-based metal-phenolic nanoparticles (MPNs) (labeled with ¹¹¹In and ⁶⁴Cu) were proposed for *in vivo* multimodal PET/SPECT/CT imaging [95].

Among other applications of Cu-based NPs is their use as a chemotherapeutic drug delivery system. Recently, Zhang [96] et al. proposed hybrid hollow mesoporous organosilica nanoparticles (HMONs) that consisted of ultras-small photothermal CuS particles and disulfiram (DSF). Upon near-infrared (NIR) irradiation, released Cu²⁺ ions from nanoparticles converted the nontoxic DSF into a highly cytotoxic diethyldithiocarbamate (DTC)-copper complex that inhibited tumor growth. In another study, thermo-responsive copper sulfide (CuS) was employed to deliver CRISPR-Cas9 ribonucleoprotein (RNP) and doxorubicin for tumor combination therapy consisting of chemotherapy, gene therapy, and photothermal therapy [97].

6. Clinical Application of Copper-Based Nanoparticles in Oncology

Although some breakthroughs have been made in the treatment of malignant tumors [98,99], therapies, such as chemotherapy and radiotherapy, have become the most

commonly used clinical treatments for tumors. However, the recurrence rate, drug resistance, quality of life, and other issues of cancer patients are still a global challenge [100]. In recent years, nanomaterials can effectively deliver drugs to specific targets, protect blood circulation drugs from endogenous enzymes, extend the half-life of drugs, and have shown great potential in tumor treatment [101,102].

Breast cancer (BC) is the second most common female cancer in the world, second only to lung cancer [99]. Studies have shown that copper-based nanomaterials have broad application prospects in the treatment of BC. For example, Ahamed et al. [103] found that copper ferrite (CuFe_2O_4) nanoparticles (NPs) added to the culture of human breast cancer MCF-7 cells can cause intracellular oxidation stress response, exerting anti-cancer effects, specifically manifested in the production of ROS and the consumption of glutathione (GSH) (Figure 3). Furthermore, Rajagopal et al. [104] found that copper nanoparticles (Wt-CuNPs) have obvious cytotoxic effects on MCF-7 cells. The specific mechanism is mainly due to the release of copper ions from the nanoparticles and the binding of copper ions to tumor cell DNA, causing DNA damage and the resulting apoptotic cell death.

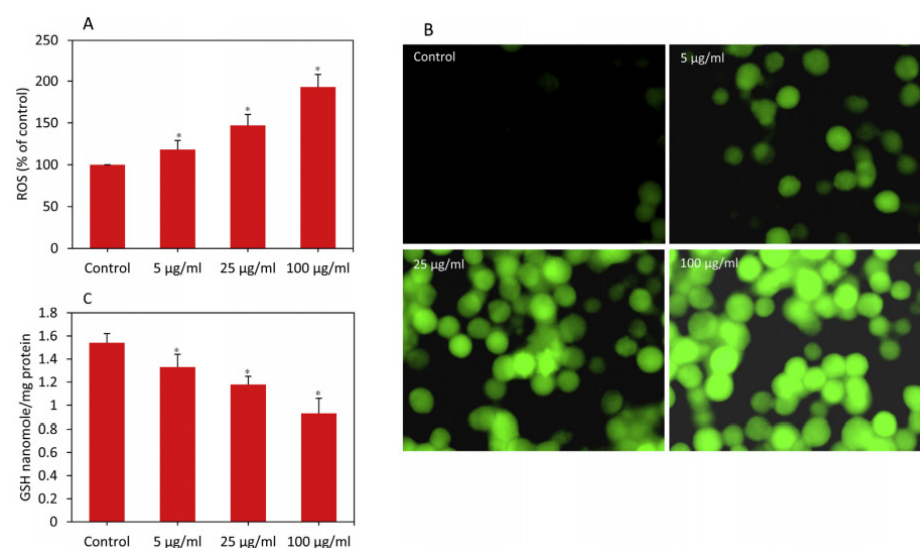


Figure 3. Copper ferrite NP-induced oxidative stress in MCF-7. Cells were exposed to copper ferrite NPs at the dosages of 0, 5, 25, and 100 g/mL for 24 h. At the end of exposure, ROS and GSH levels were determined, as described in materials and methods. (A) Percentage change in ROS level. (B) Fluorescence microscopy image of ROS generation. (C) GSH level. Data represented are mean \pm SD of three identical experiments made in three replicates. * Significant difference as compared to control ($p < 0.05$).

Copper-based nanomaterials have also achieved good results in the treatment of esophageal cancer. Wang et al. [105] covered the silica coating on the Cu_9S_5 nanoparticles to form $\text{Cu}_9\text{S}_5@\text{MS}$ core-shell nanostructures and added $\text{Cu}_9\text{S}_5@\text{MS}$ core-shell nanostructures to human esophageal squamous carcinoma Eca109 and TE8 cells. After co-cultivation and treatment with NIR, it was found that $\text{Cu}_9\text{S}_5@\text{MS}$ + NIR performs active anticancer activity against the EC109 and TE8 cancer cell lines by cell cycle arrest (Figure 4).

Furthermore, Xu et al. [106] optimized the concentration of disulfiram and Cu^{2+} ion for inhibiting esophageal cancer cells and loaded them in hyaluronic acid (HA)/polyethyleneimine (PEI) nanoparticles with specific scales to obtain NP-HPDCu²⁺ nanoparticles to improve the effectiveness and targeting of the drug. In vitro experiments proved that NP-HPDCu²⁺ nanomaterials can significantly promote the occurrence of Eca109 cell apoptosis and inhibit the migration and invasion of Eca109 (Figure 5). At the same time, the nude mouse tumor model proves that NP-HPDCu²⁺ nanomaterials can reduce the tumor volume and keep the weight of nude mice stable. The results of tumor tissue immunohistochemistry, immunofluorescence staining, and western blotting also showed that NP-HPDCu²⁺

nanomaterials can promote apoptosis and inhibit proliferation of esophageal squamous cell carcinoma.

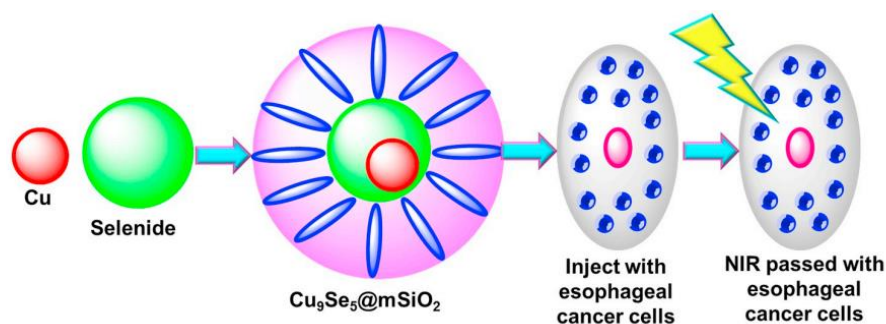


Figure 4. Portrayal of the Cu₉S₅@MS nanoparticles synthesis and application as a dual functional treatment stage for esophageal squamous carcinoma treatment.

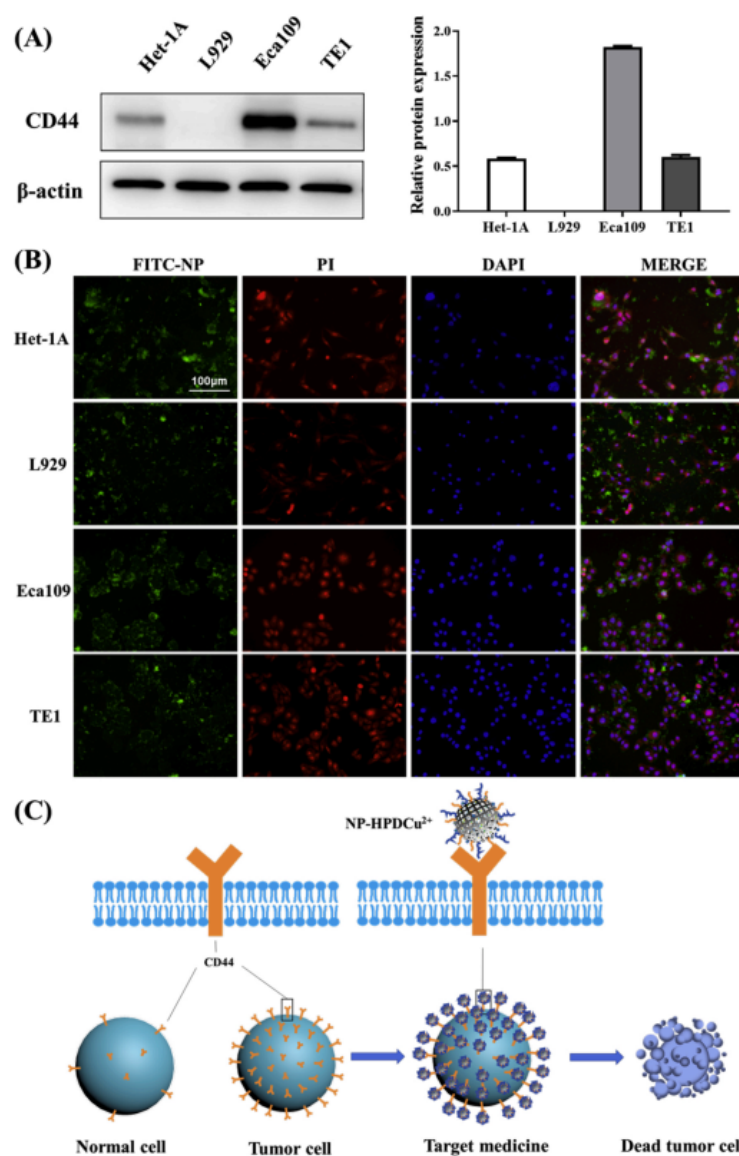


Figure 5. (A) Western blot of CD44 expressed on the Het-1A, L929, Eca109, and TE1 (mean ± SD, n = 3); (B) Fluorescence images of Het-1A, L929, Eca109, and TE1 stained with FITC-labeled NP-HPDCu²⁺ (FITC-NP, green color), PI (apoptosis marker, red color) and DAPI (nucleus marker, blue color); (C) mechanism diagram of targeted killing tumor cells by NP-HPDCu²⁺ nanoparticle.

Lung cancer is the malignant tumor with the highest mortality rate in the world, and non-small cell lung cancer is the most common pathological type in clinic [107,108]. Some researchers have found that copper-based nanomaterials have shown great potential in the treatment of NSCLC. Naatz et al. [109] constructed a new type of nanomaterial, Fe-doped CuO nanomaterial, which can use doped Fe to control the dissolution kinetics of copper-based nanomaterials. Using mouse lung squamous cell KLN-205 to construct a tumor-bearing nude mouse model by regulating the release of Cu^{2+} , the local long-term drug concentration can be maintained, and the occurrence of drug resistance can be reduced. Additionally, these particles can also trigger a systemic anti-cancer immune response, promote the generation of ROS, and increase the rate of tumor cell death, which shows that CuO nanomaterials also have broad prospects for anti-cancer applications (Figure 6). In addition, Kalaiarasi et al. [110] reported that in A549 cells, the anti-cancer effect of CuO copper-based nanomaterials is related to the inhibition of histone deacetylase (HDACs) expression. Specifically, CuO copper-based nanomaterials have a strong inhibitory effect on different types of HDACs, can down-regulate the expression of oncogenes and up-regulate the expression of tumor suppressor genes, and induce apoptosis of cancer cells by activating the caspase cascade pathway to exert anti-cancer effects.

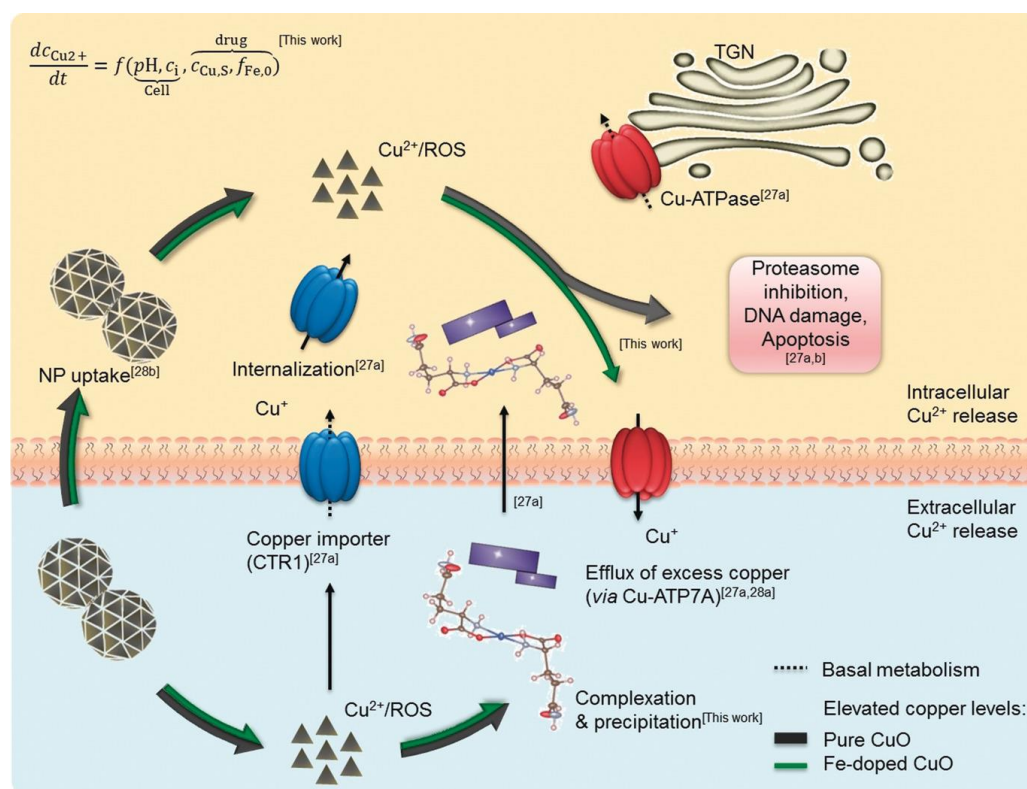


Figure 6. Copper homeostasis and regulatory mechanisms, including extra- and intracellular dissolution of pure and Fe-doped CuO NPs.

In recent years, with the continuous in-depth research of nanomaterials compared with traditional antitumor treatments, nanomaterials have been used in more and more clinical anticancer applications, showing great development potential [111]. For example, in our previous research, we found that some nanoparticles, such as superparamagnetic iron oxide nanoparticles (SPIONs), high-Z gold nanoparticles following intratumoral injection can provide a high local concentration of the agent, reduction of the particle clearance (i.e., renal or hepatic clearance) that increases the bioavailability of nanoparticles and has the effect of radiosensitizer in cancer radiotherapy, which can be used for long-term local anti-tumor therapy [112,113]. As an ideal anti-tumor drug candidate, copper-based nanomaterials have the following advantages: (i) compared with other metals, copper is cheap and rich

in content [114], (ii) copper can induce reactive oxygen species (ROS)-mediated oxidative stress and promote tumor cell apoptosis [115–117], (iii) it has good biocompatibility, biodegradability, antibacterial properties, and selective cytotoxicity to cancer cells [118], and (iv) copper-based nanomaterials have less toxic effects on normal cells, fewer side effects, and are safer and more reliable [119]. Thus, copper-based nanomaterials have attracted more and more attention and have become the current research hotspot. At the same time, the emergence of copper-based nanomaterials has brought down to the treatment of various tumors [120].

However, copper-based nanomaterials also have limitations. For example, the production process of copper-based nanomaterials uses physical and chemical methods that are harmful to the environment and the human body [121]. Additionally, the instability and susceptibility to oxidation of copper-based nanomaterials under physiological conditions may also hinder its anti-tumor effect and reliability [122]. Furthermore, the biological safety of copper-based nanomaterials still requires further cell and molecular studies to avoid any impact on health, since Fahmy et al. [123] found that copper/copper oxide nanoparticles showed cytotoxicity to normal human lung WI-38 cells, resulting in the production of reactive oxygen species and DNA damage and inhibiting the growth and proliferation of WI-38 cells. The stability of copper nanoparticles is also one of the major concerns as copper tends to aggregate to the proteins, specifically cysteine and methionine residues. One work also found a dependency between stability and pH value [124]. However, using green synthesis, the authors successfully designed NPs which are mostly stable at various pH levels.

In short, copper-based nanomaterials are currently ideal anti-tumor drug candidates. With the continuous development of nanomaterials research, it will help provide better cancer treatment strategies in the future.

7. The Combination of Nanoparticles with Other Treatment Modalities

Based on the biological effects of copper and the physical and chemical properties of copper nanoparticles, their applications in the biomedical field mainly include externally triggered nanotherapies (photothermal therapy), drug delivery, antimicrobial applications, tissue regeneration, bioimaging, and bioeffects/biosafety. Therefore, it is reasonable to be expected that the construction of Cu-based biomaterials will have a unique integrated diagnosis and treatment function in clinical medicine. However, due to the complexity of tumors, such as the specific microenvironment and tumor metastasis, it is difficult to eradicate tumors completely through monotherapy alone. Therefore, the development of unique treatment modalities with multiple synergistic therapeutic performance has high prospects for improving therapeutic efficacy. Therefore, rational design of optimal drug combinations is important to achieve optimal synergistic therapeutic effects. Based on this, several unique multifunctional nanosystems involving copper have been constructed to jointly generate multiple nanotherapeutics [125].

Copper chalcogenides (Cu_{2-x}E , E: S, Se, Te, $0 \leq x \leq 1$) have been widely explored in photon-triggered disease therapy, such as photoacoustic imaging and photothermal hyperthermia. With stoichiometric ratios (Cu_{2-x}S), deficient cuprous sulfide exhibits stoichiometric-dependent localized surface plasmon resonance (LSPR) absorption in the near-infrared range and photothermal conversion [126]. The integration of magnetic Fe_3O_4 nanoparticles exerted a magnetic targeting function to enhance tumor accumulation. Importantly, the photonic response of these Fe_3O_4 @CuS composite nanoparticles in the second NIR biological window (1064 nm) achieves higher tissue penetration ability compared to the laser activation of the first NIR biological window. Thus, a higher tumor suppression rate was achieved with no further recurrence (808 nm). In addition to the photothermal conversion efficiency (25.7%) of hydrophilic plate-like Cu₉S₅ nanocrystals at 980 nm [127], the CuS superstructure was exemplified to respond to external 980 nm laser activation for photothermal conversion and subsequent cancer ablation [128]. The cysteine-coated CuS nanoparticles were also irradiated with a 980 nm laser with a high photothermal conversion

efficiency of 38.0%, efficiently inhibiting tumor growth [129]. Furthermore, encapsulation of CuS nanoparticles into zeolite imidazole framework 8 (ZIF-8) resulted in NIR-induced dissociation of ZIF-8 to release loaded chemotherapeutics, aiming to achieve synergistic photothermal ablation and NIR-triggered chemotherapy [130]. Doping iron (Fe^{3+}) can tune the vacancies of Cu_{2-x}S nanoparticles to control NIR absorption, which also enables these semiconductors to have MR-imaging properties [131].

To improve the photothermal conversion efficiency, Cu_{2-x}S and Ag_2S were integrated into one system by producing Cu-Ag₂S/PVP nanoparticles with a high photothermal conversion efficiency of 58.2% under 808 nm laser irradiation, which is much higher than that of Cu_{2-x}S /PVP nanoparticles (27.1%) [132]. The rational integration of plasmonic Au nanoparticles and plasmonic Cu_{2-x}S semiconductors into one matrix can enhance the photothermal properties of Au or Cu_{2-x}S components. The coupled LSPR properties of Au and Cu_{2-x}S can be maximized by designing Au@ Cu_{2-x}S core/shell nanoparticles to enhance the PTT efficacy. Ji et al. synthesized Au@CuS nanoparticles and performed the following cation exchange between Cu^+ and CdS shells, resulting in Au@ Cu_{2-x}S nanostructures [133], which can be formed as nanoparticles or nanorods. The corresponding photothermal conversion efficiencies are calculated to be 59% at 808 nm and 43% at 1064 nm, which rapidly increases the ambient temperature of the Au@ Cu_{2-x}S nanorod aqueous solution. In particular, the design of core/shell Au@ Cu_{2-x}S is more favorable compared to the simple mixture of Au nanorods and Cu_{2-x}S nanoparticles for photothermal conversion. This core/shell design with improved photothermal performance also induced more HeLa cell death compared to the same concentration of Cu_{2-x}S . The Au-Cu₉S₅ plasmonic hybrid nanosystem was established, which enhanced the LSPR of Cu₉S₅ through the coupling effect of LSPR based on the collective vibration of electrons and holes [134]. This Au-Cu₉S₅ hybrid nanosystem exhibits an absorption cross-section enhancement of $1.3 \times 10^8 \text{ m}^{-1} \text{ cm}^{-1}$ and a high photothermal conduction efficiency of 37% for photothermal ablation of tumor tissue. According to the plasmonic coupling effect between core and shell, spherical Au@ Cu_{2-x}S , Au@ Cu_{2-x}S , and rod-shaped Au@ Cu_{2-x}S superparticles were synthesized for photothermal ablation of tumors (4T1 tumor model). It has X-ray-computed, tomography-imaging capabilities because of the presence of Au composition with a large atomic number and an X-ray attenuation coefficient ($5.16 \text{ cm}^{-2} \text{ kg}^{-1}$) [135].

Photothermal therapy exposes materials with the photothermal conversion ability to near-infrared light. These materials can convert the absorbed light energy into thermal energy to kill tumors, showing excellent local tumor treatment effects, but they are less effective for metastatic tumors. The combination of photothermal therapy and radiotherapy in tumor treatment can achieve a synergistic effect. Thus, Zhou et al. [89] synthesized PEG-[⁶⁴Cu]CuS NPs based on a single radioactive copper sulfide nanoparticle. The study demonstrated that inhibition of tumor growth was significantly high when both methods, radiotherapy and hyperthermia, were employed.

Photothermal therapy (PTT) mainly uses photothermal materials accumulated at the tumor site, which can convert the absorbed light energy into heat energy (above 45 °C) under near-infrared irradiation. Combining tumor photothermal therapy and immunotherapy could further improve the therapeutic potency of PTT [136]. Another approach could be based on the combination of PTT with chemotherapy. Thus, Wu et al. [137] demonstrated that encapsulation of CuS nanoparticles into the zeolite imidazole framework 8 (ZIF-8) resulted in NIR-induced dissociation of ZIF-8 to release loaded chemotherapeutics, which in turn provided synergistic photothermal ablation and NIR-triggered chemotherapy.

The tumor microenvironment is usually characterized by low pH [138], altered redox states [139], hypoxia [140], and expression of particular enzymes that could be employed for the development of stimuli-responsive nanoparticles. Based on the fact that the hydrogen sulfide (H_2S)-producing enzyme of cystathionine- β -synthase (CBS) is upregulated in colon cancer, H_2S concentrations in tumors reach approximately 0.3 to 3.4 $\text{mmol}\cdot\text{L}^{-1}$. Therefore, using this overexpressed endogenous H_2S to convert cuprous oxide (Cu_2O) to copper sulfide in situ can activate PA imaging and photothermal tumor ablation [141]. It is

exemplified that the use of S-adenosyl-1-methionine (SAM) as an allosteric CBS activator accelerates the in situ reaction between H₂S and Cu₂O, resulting in significantly enhanced PA-imaging signal and photothermal effect. In contrast, the use of aminoxyacetic acid (AOAA) as a CBS inhibitor reduced the production of H₂S and subsequently the conversion of Cu₂O to copper sulfide, showing no significant PA signal and negligible temperature change in tumors. However, the photothermal conversion efficiency after high-dose copper sulfide conversion is low, and the ideal photon therapy effect cannot be obtained. To address this critical issue, based on the LSPR-coupling effect between noble metals and plasmonic semiconductors, Tao et al. constructed Au@Cu₂O plasmonic hybrids to enhance in situ H₂S-triggered post-conversion photothermal performance [142]. Similar to the conversion of Cu₂O to Cu₉S₈, tumor-accumulated Au@Cu₂O nanoparticles were also converted into Au@Cu₉S₈ nanoagents to achieve PA-enhanced contrast agents and photothermal tumor ablation by increasing tumor temperature. The LSPR-coupling effect induces nearly 2.1-fold stronger NIR absorption and 1.2-fold higher photothermal conversion efficiency, enabling the utilization of low nanoparticle doses with desirable therapeutic properties. These two paradigms provide another strategy for realizing photothermal hyperthermia involving copper-based nanoagents by in situ generation of copper-based nanoagents with unique photothermal properties. Cheng Y. et al. [143] took advantage of the ordered large-pore structure and easily chemically modified the property of DLMSNs, the copper sulfide (CuS) nanoparticles with high photothermal conversion efficiency. A homogenous cancer cell membrane was coated on the surfaces of these DLMSNs, followed by conjugation with the anti-PD-1 peptide. The thus-obtained AM@DLMSN@CuS/R848 was applied to holistically treat metastatic TNBC in vitro and in vivo. The data showed that AM@DLMSN@CuS/R848 had a high TNBC-targeting ability and induced efficient photothermal ablation on primary TNBC tumors under 980 nm laser irradiation. Tumor antigens thus generated and increasingly released R848 by response to the photothermal effect, combined with AUNP-12 detached from AM@DLMSN@CuS/R848 in the weakly acidic tumor microenvironment and synergistically exerted an anti-tumor effect, thus preventing TNBC recurrence and metastasis.

Table 1 summarizes the above information presenting major classes of therapeutics and some examples for detail consideration. The unique features of copper allow to create a wide spectrum of various nanostructures with great diversity of their applications.

Table 1. Copper-based compounds and nanoparticles with various applications and mechanisms of action.

	Copper-Based Compound	Mechanism of Action
Diagnostic tool	64-CuCl ₂ [64]	Contrast agent in PET/MRI scanning
	Combination of SPIONs and Cu(II) [62] Gold-copper alloyed NPs [63]	
Therapeutic agent	Schiff base copper (II) complexes [66]	Activation of extrinsic or intrinsic apoptotic pathways
	Copper-based nanoparticles [96,103]	Copper ions release, oxidative stress, DNA damage
	Thiosemicarbazones copper (II) complex [72]	Topoisomerase inhibition
	Polypyridyl-Schiff-base copper complex [74]	Targets endoplasmic reticulum leading to immunogenic cell death
	G-quadruplex-targeting copper complex [77]	Rapid reduction of telomeres in cancer cells
	Ferrocenyl terpyridine copper complexes [78]	Targets mitochondria, causes mtDNA damage

Table 1. Cont.

	Copper-Based Compound	Mechanism of Action
	Copper chalcogenides [126] Alloyed CuAg or CuAu NPs [132,133]	Photothermal ablation and NIR-triggered chemotherapy
Combined approach	PEG-[64Cu]CuS NPs [94]	Combined radiotherapy and hyperthermia against metastatic tumor cells
	Copper-doped iron NPs [109,131]	Magnetic guidance and copper release with subsequent oxidative stress

8. Conclusions

Copper is an essential trace element in cell metabolism with distinct features. Participation of copper in oxidation–reduction reactions has an important impact on cell metabolism, survival, and growth. Free copper ions could exert a cytotoxic effect; however, most of the copper is bound to the enzymes, metallochaperones, and metallothioneins. These proteins, despite their direct function, could influence functionality of other proteins affecting cell signaling and gene expression, interfering in the anti-cancer chemotherapies. Recent studies demonstrate that copper-based nanocarriers due to their unique physio-chemical properties could be efficiently employed for tumor theranostics as a monotherapeutic approach or in combination with other treatment modalities. Constant development and modification of existing systems have great potential in clinic. Some limitations, which include ROS generation and free ion emergence, should be considered. However, an understanding of the underlying molecular regulation of copper intracellular distribution and metabolism will help to improve the current development of copper-based therapeutics and nanostructures for further efficient clinical application.

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References

- Shanbhag, V.C.; Gudekar, N.; Jasmer, K.; Papageorgiou, C.; Singh, K.; Petris, M.J. Copper metabolism as a unique vulnerability in cancer. *Biochim. Biophys. Acta Mol. Cell Res.* **2021**, *1868*, 118893. [[CrossRef](#)] [[PubMed](#)]
- Skopp, A.; Boyd, S.D.; Ullrich, M.S.; Liu, L.; Winkler, D.D. Copper–zinc superoxide dismutase (Sod1) activation terminates interaction between its copper chaperone (Ccs) and the cytosolic metal-binding domain of the copper importer Ctr1. *Biomaterials* **2019**, *32*, 695–705. [[CrossRef](#)] [[PubMed](#)]
- Nývltová, E.; Dietz, J.V.; Seravalli, J.; Khalimonchuk, O.; Barrientos, A. Coordination of metal center biogenesis in human cytochrome c oxidase. *Nat. Commun.* **2022**, *13*, 3615. [[CrossRef](#)] [[PubMed](#)]
- Postma, G.C.; Nicastro, C.N.; Valdez, L.B.; Mikusic, I.A.R.; Grecco, A.; Minatel, L. Decrease lysyl oxidase activity in hearts of copper-deficient bovines. *J. Trace Elem. Med. Biol.* **2021**, *65*, 126715. [[CrossRef](#)]
- Grasso, M.; Bond, G.J.; Kim, Y.J.; Boyd, S.; Dzebo, M.M.; Valenzuela, S.; Tsang, T.; Schibrowsky, N.A.; Alwan, K.B.; Blackburn, N.J.; et al. The copper chaperone CCS facilitates copper binding to MEK1/2 to promote kinase activation. *JBC* **2021**, *297*, 101314. [[CrossRef](#)] [[PubMed](#)]

6. Krishnamoorthy, L.; Cotruvo, J.A., Jr.; Chan, J.; Kaluarachchi, H.; Muchenditsi, A.; Pendyala, V.S.; Jia, S.; Aron, A.T.; Ackerman, C.M.; Vander Wal, M.N.; et al. Copper regulates cyclic-AMP-dependent lipolysis. *Nat. Chem. Biol.* **2016**, *12*, 586–592. [[CrossRef](#)]
7. Guo, H.; Li, K.; Wang, W.; Wang, C.; Shen, Y. Effects of copper on hemocyte apoptosis, ROS production, and gene expression in white shrimp *Litopenaeus vannamei*. *Biol. Trace Elem. Res.* **2017**, *179*, 318–326. [[CrossRef](#)]
8. Chen, J.; Jiang, Y.; Shi, H.; Peng, Y.; Fan, X.; Li, C. The molecular mechanisms of copper metabolism and its roles in human diseases. *Pflug Arch. Eur. J. Phys.* **2020**, *472*, 1415–1429. [[CrossRef](#)]
9. Lee, J.; Petris, M.J.; Thiele, D.J. Biochemical characterization of the human copper transporter Ctr1. *J. Biol. Chem.* **2002**, *277*, 4380–4387. [[CrossRef](#)]
10. Ren, F.; Logeman, B., L.; Zhang, X.; Liu, Y.; Thiele, D., J.; Yuan, P. X-ray structures of the high-affinity copper transporter Ctr1. *Nat. Commun.* **2019**, *10*, 1386. [[CrossRef](#)]
11. Chen, H.; Xu, C.; Yu, Q.; Zhong, C.; Peng, Y.; Chen, J.; Chen, G. Comprehensive landscape of STEAP family functions and prognostic prediction value in glioblastoma. *J. Cell. Physiol.* **2021**, *236*, 2988–3000. [[CrossRef](#)] [[PubMed](#)]
12. Inesi, G. Molecular features of copper binding proteins involved in copper homeostasis. *IUBMB Life* **2017**, *69*, 211–217. [[CrossRef](#)] [[PubMed](#)]
13. Luchinat, E.; Barbieri, L.; Banci, L. A molecular chaperone activity of CCS restores the maturation of SOD1 fALS mutants. *Sci. Rep.* **2017**, *7*, 17433. [[CrossRef](#)]
14. Banks, C.J.; Andersen, J.L. Mechanisms of SOD1 regulation by post-translational modifications. *Redox Biol.* **2019**, *26*, 101270. [[CrossRef](#)] [[PubMed](#)]
15. Vanišová, M.; Burská, D.; Křížová, J.; Daňhelovská, T.; Dosoudilová, Ž.; Zeman, J.; Stibůrek, L.; Hansíková, H. Stable COX17 downregulation leads to alterations in mitochondrial ultrastructure, decreased copper content and impaired cytochrome c oxidase biogenesis in HEK293 cells. *Folia Biol.* **2019**, *65*, 181–187.
16. La Fontaine, S.; Mercer, J.F. Trafficking of the copper-ATPases, ATP7A and ATP7B: Role in copper homeostasis. *Arch. Biochem. Biophys.* **2007**, *463*, 149–167. [[CrossRef](#)]
17. Zhang, W.; Shi, H.; Chen, C.; Ren, K.; Xu, Y.; Liu, X.; He, L. 2 Curcumin enhances cisplatin sensitivity of human NSCLC cell lines through influencing Cu-Sp1-CTR1 regulatory loop. *Phytomedicine* **2018**, *48*, 51–61. [[CrossRef](#)]
18. Sinani, D.; Adle, D.J.; Kim, H.; Lee, J. Distinct mechanisms for Ctr1-mediated copper and cisplatin transport. *J. Biol. Chem.* **2007**, *282*, 26775–26785. [[CrossRef](#)]
19. Sun, S.; Cai, J.; Yang, Q.; Zhao, S.; Wang, Z. The association between copper transporters and the prognosis of cancer patients undergoing chemotherapy: A meta-analysis of literatures and datasets. *Oncotarget* **2017**, *8*, 16036. [[CrossRef](#)]
20. Ilyechova, E.Y.; Bonaldi, E.; Orlov, I.A.; Skomorokhova, E.A.; Puchkova, L.V.; Broggin, M. CRISP-R/Cas9 mediated deletion of copper transport genes CTR1 and DMT1 in NSCLC cell line H1299. Biological and pharmacological consequences. *Cells* **2019**, *8*, 322. [[CrossRef](#)]
21. Akerfeldt, M.C.; Tran, C.M.N.; Shen, C.; Hambley, T.W.; New, E.J. Interactions of cisplatin and the copper transporter CTR1 in human colon cancer cells. *J. Biol. Inorg. Chem.* **2017**, *22*, 765–774. [[CrossRef](#)] [[PubMed](#)]
22. Ishida, S.; Lee, J.; Thiele, D.J.; Herskowitz, I. Uptake of the anticancer drug cisplatin mediated by the copper transporter Ctr1 in yeast and mammals. *PNAS* **2002**, *99*, 14298–14302. [[CrossRef](#)] [[PubMed](#)]
23. Safaei, R.; Maktabi, M.H.; Blair, B.G.; Larson, C.A.; Howell, S. Effects of the loss of Atox1 on the cellular pharmacology of cisplatin. *J. Inorg. Biochem.* **2009**, *103*, 333–341. [[CrossRef](#)]
24. Bompiani, K.M.; Tsai, C.Y.; Achatz, F.P.; Liebig, J.K.; Howell, S.B. Copper transporters and chaperones CTR1, CTR2, ATOX1, and CCS as determinants of cisplatin sensitivity. *Metallomics* **2016**, *8*, 951–962. [[CrossRef](#)] [[PubMed](#)]
25. Blockhuys, S.; Brady, D.C.; Wittung-Stafshede, P. Evaluation of copper chaperone ATOX1 as prognostic biomarker in breast cancer. *Breast Cancer* **2020**, *27*, 505–509. [[CrossRef](#)]
26. Blockhuys, S.; Wittung-Stafshede, P. Copper chaperone Atox1 plays role in breast cancer cell migration. *Biochem. Biophys. Res. Commun.* **2017**, *483*, 301–304. [[CrossRef](#)]
27. Itoh, S.; Kim, H.W.; Nakagawa, O.; Ozumi, K.; Lessner, S.M.; Aoki, H.; Akram, K.; McKinney, R.D.; Ushio-Fukai, M.; Fukai, T. Novel role of antioxidant-1 (Atox1) as a copper-dependent transcription factor involved in cell proliferation. *J. Biol. Chem.* **2008**, *283*, 9157–9167. [[CrossRef](#)]
28. Beaino, W.; Guo, Y.; Chang, A.J.; Anderson, C.J. Roles of Atox1 and p53 in the trafficking of copper-64 to tumor cell nuclei: Implications for cancer therapy. *J. Biol. Inorg. Chem.* **2014**, *19*, 427–438. [[CrossRef](#)]
29. Singh, R.P.; Jeyaraju, D.V.; Voisin, V.; Xu, C.; Barghout, S.H.; Khan, D.H.; Hurren, R.; Gronda, M.; Wang, X.; Jitkova, Y.; et al. Targeting the Mitochondrial Metallochaperone Cox17 Reduces DNA Methylation and Promotes AML Differentiation through a Copper Dependent Mechanism. *Blood* **2018**, *132*, 1339. [[CrossRef](#)]
30. Zhao, L.; Cheng, Q.; Wang, Z.; Xi, Z.; Xu, D.; Liu, Y. Cisplatin binds to human copper chaperone Cox17: The mechanistic implication of drug delivery to mitochondria. *Chem. Commun.* **2014**, *50*, 2667–2669. [[CrossRef](#)]
31. Zhao, L.; Wang, Z.; Wu, H.; Xi, Z.; Liu, Y. Glutathione selectively modulates the binding of platinum drugs to human copper chaperone Cox17. *Biochem. J.* **2015**, *472*, 217–223. [[CrossRef](#)] [[PubMed](#)]
32. Sotgia, F.; Fiorillo, M.; Lisanti, M.P. Mitochondrial markers predict recurrence, metastasis and tamoxifen-resistance in breast cancer patients: Early detection of treatment failure with companion diagnostics. *Oncotarget* **2017**, *8*, 68730–68745. [[CrossRef](#)] [[PubMed](#)]

33. Gao, S.P.; Sun, H.F.; Fu, W.Y.; Li, L.D.; Zhao, Y.; Chen, M.T.; Jin, W. High expression of COX5B is associated with poor prognosis in breast cancer. *Future Oncol.* **2017**, *13*, 1711–1719. [[CrossRef](#)] [[PubMed](#)]
34. Bikas, A.; Jensen, K.; Patel, A.; Costello, J.; Reynolds, S.M.; Mendonca-Torres, M.C.; Thakur, S.; Klubo-Gwiedzinska, J.; Ylli, D.; Wartofsky, L.; et al. Cytochrome C Oxidase Subunit 4 (COX4): A Potential Therapeutic Target for the Treatment of Medullary Thyroid Cancer. *Cancers* **2020**, *12*, 2548. [[CrossRef](#)]
35. Yi, J.F.; Li, Y.-M.; Liu, T.; He, W.-T.; Li, X.; Zhou, W.-C.; Kang, S.-L.; Zeng, X.-T.; Zhang, J.-Q. Mn-SOD and CuZn-SOD polymorphisms and interactions with risk factors in gastric cancer. *World J. Gastroenterol.* **2010**, *16*, 4738–4746. [[CrossRef](#)]
36. Ahmed, A.S.; Eryilmaz, R.; Demir, H.; Aykan, S.; Demir, C. Determination of oxidative stress levels and some antioxidant enzyme activities in prostate cancer. *Aging Male* **2019**, *22*, 198–206. [[CrossRef](#)]
37. Liu, S.; Li, B.; Xu, J.; Hu, S.; Zhan, N.; Wang, H.; Gao, C.; Li, J.; Xu, X. SOD1 Promotes Cell Proliferation and Metastasis in Non-small Cell Lung Cancer via an miR-409-3p/SOD1/SETDB1 Epigenetic Regulatory Feedforward Loop. *Front. Cell Dev. Biol.* **2020**, *8*, 213. [[CrossRef](#)]
38. Li, Y.; Liang, R.; Zhang, X.; Wang, J.; Shan, C.; Liu, S.; Li, L.; Zhang, S. Copper Chaperone for Superoxide Dismutase Promotes Breast Cancer Cell Proliferation and Migration via ROS-Mediated MAPK/ERK Signaling. *Front. Pharmacol.* **2019**, *10*, 356. [[CrossRef](#)]
39. Wang, J.; Luo, C.; Shan, C.; You, Q.; Lu, J.; Elf, S.; Zhou, Y.; Wen, Y.; Vinkenborg, J.L.; Fan, J.; et al. Inhibition of human copper trafficking by a small molecule significantly attenuates cancer cell proliferation. *Nat. Chem.* **2015**, *7*, 968–979. [[CrossRef](#)]
40. Wen, C.; Shan, C.L.; Sun, W.J.; Wan, Y.; Lin, R.; Chen, B.; Dai, H.-T.; Tang, K.-Y.; Xiang, X.-H.; Yang, J.-Y.; et al. Copper chaperone for superoxide dismutase expression is down-regulated and correlated with more malignant tumoral features and poor prognosis in human hepatocellular carcinoma. 2021, *preprint*. [[CrossRef](#)]
41. Samimi, G.; Varki, N.M.; Wilczynski, S.; Safaei, R.; Alberts, D.S.; Howell, S.B. Increase in expression of the copper transporter ATP7A during platinum drug-based treatment is associated with poor survival in ovarian cancer patients. *Clin. Cancer Res.* **2003**, *9*, 5853–5859.
42. Chisholm, C.L.; Wang, H.; Wong, A.H.; Vazquez-Ortiz, G.; Chen, W.; Xu, X.; Deng, C.X. Ammonium tetrathiomolybdate treatment targets the copper transporter ATP7A and enhances sensitivity of breast cancer to cisplatin. *Oncotarget* **2016**, *7*, 84439. [[CrossRef](#)] [[PubMed](#)]
43. Seiko, I.; Frank, M.; Karen, S.; Douglas, H. Enhancing Tumor-Specific Uptake of the Anticancer Drug Cisplatin with a Copper Chelator. *Cancer Cell* **2010**, *17*, 574–583. [[CrossRef](#)]
44. Mangala, L.S.; Zuzel, V.; Schmandt, R.; Leshane, E.S.; Halder, J.B.; Armaiz-Pena, G.N.; Spannuth, W.A.; Tanaka, T.; Shahzad, M.M.K.; Lin, Y.G.; et al. Therapeutic Targeting of ATP7B in Ovarian Carcinoma. *Clin. Cancer Res.* **2009**, *15*, 3770–3780. [[CrossRef](#)] [[PubMed](#)]
45. David, L.; Maruša, H.; Borut, K.; Katarina, Č. The contribution of copper efflux transporters ATP7A and ATP7B to chemo-resistance and personalized medicine in ovarian cancer. *Biomed. Pharmacother.* **2020**, *129*, 110401. [[CrossRef](#)]
46. Yu, Z.; Cao, W.; Ren, Y.; Zhang, Q.; Liu, J. ATPase copper transporter A, negatively regulated by miR-148a-3p, contributes to cisplatin resistance in breast cancer cells. *Clin. Transl. Med.* **2020**, *10*, 57–73. [[CrossRef](#)]
47. Sen, C.K.; Khanna, S.; Venojarvi, M.; Trikha, P.; Ellison, E.C.; Hunt, T.K.; Roy, S. Copper-induced vascular endothelial growth factor expression and wound healing. *Am. J. Physiol. Cell Physiol.* **2002**, *282*, 9157–9167. [[CrossRef](#)]
48. Wu, Z.; Zhang, W.; Kang, Y.J. Copper affects the binding of HIF-1 α to the critical motifs of its target genes. *Metallomics* **2019**, *11*, 429–438. [[CrossRef](#)]
49. Wee, N.K.; Weinstein, D.C.; Fraser, S.T.; Assinder, S.J. The mammalian copper transporters CTR1 and CTR2 and their roles in development and disease. *Int. J. Biochem. Cell Biol.* **2013**, *45*, 960–963. [[CrossRef](#)]
50. Wang, W.; Wang, X.; Luo, J.; Chen, X.; Ma, K.; He, H.; Li, W.; Cui, J. Serum Copper Level and the Copper-to-Zinc Ratio Could Be Useful in the Prediction of Lung Cancer and Its Prognosis: A Case-Control Study in Northeast China. *Nutr. Cancer* **2021**, *73*, 1908–1915. [[CrossRef](#)]
51. Brady, D.C.; Crowe, M.S.; Turski, M.L.; Hobbs, G.A.; Yao, X.; Chaikuad, A.; Knapp, S.; Xiao, K.; Campbell, S.L.; Thiele, D.J.; et al. Copper is required for oncogenic BRAF signaling and tumorigenesis. *Nature* **2014**, *509*, 492–496. [[CrossRef](#)] [[PubMed](#)]
52. Kim, D.W.; Shin, M.J.; Choi, Y.J.; Kwon, H.J.; Lee, S.H.; Lee, S.; Lee, S.; Park, J.; Han, K.H.; Eum, W.S.; et al. Tat-ATOX1 inhibits inflammatory responses via regulation of MAPK and NF- κ B pathways. *BMB Rep.* **2018**, *51*, 654–659. [[CrossRef](#)] [[PubMed](#)]
53. Tsai, C.Y.; Finley, J.C.; Ali, S.S.; Patel, H.H.; Howell, S.B. Copper influx transporter 1 is required for FGF, PDGF and EGF-induced MAPK signaling. *Biochem. Pharmacol.* **2012**, *84*, 1007–1013. [[CrossRef](#)]
54. Li, Y.; Fu, S.Y.; Wang, L.H.; Wang, F.Y.; Wang, N.N.; Cao, Q.; Wang, Y.-T.; Yangab, J.-Y.; Wu, C.-F. Copper improves the anti-angiogenic activity of disulfiram through the EGFR/Src/VEGF pathway in gliomas. *Cancer Lett.* **2015**, *369*, 86–96. [[CrossRef](#)] [[PubMed](#)]
55. Thakor, A.S.; Jokerst, J.V.; Ghanouni, P.; Campbell, J.L.; Mitra, E.; Gambhir, S.S. Clinically Approved Nanoparticle Imaging Agents. *J. Nucl. Med.* **2016**, *57*, 1833–1837. [[CrossRef](#)]
56. Gutfilen, B.; Souza, S.A.; Valentini, G. Copper-64: A real theranostic agent. *Drug Des. Devel. Ther.* **2018**, *12*, 3235–3245. [[CrossRef](#)]
57. Xue, X.; Zhang, H.; Liu, H.; Wang, S.; Li, J.; Zhou, Q.; Chen, X.; Ren, X.; Jing, Y.; Deng, Y.; et al. Rational Design of Multifunctional CuS Nanoparticle-PEG Composite Soft Hydrogel-Coated 3D Hard Polycaprolactone Scaffolds for Efficient Bone Regeneration. *Adv. Funct. Mater.* **2022**, *32*, 2202470. [[CrossRef](#)]

58. Xue, X.; Liu, H.; Wang, S.; Hu, Y.; Huang, B.; Li, M.; Chen, X.; Ren, X.; Jing, Y.; Deng, Y.; et al. Neutrophil-erythrocyte hybrid membrane-coated hollow copper sulfide nanoparticles for targeted and photothermal/anti-inflammatory therapy of osteoarthritis. *Compos. B Eng.* **2022**, *237*, 109855. [[CrossRef](#)]
59. Lu, H.; Xu, S.; Ge, G.; Guo, Z.; Zhao, M.; Liu, Z. Boosting Chemodynamic Therapy by Tumor-Targeting and Cellular Redox Homeostasis-Disrupting Nanoparticles. *ACS Appl. Mater.* **2022**, *14*, 44098–44110. [[CrossRef](#)]
60. Fanizza, E.; Mastrogiacomo, R.; Pugliese, O.; Guglielmelli, A.; De Sio, L.; Castaldo, R.; Scavo, M.P.; Giancaspro, M.; Rizzi, F.; Gentile, G.; et al. NIR-Absorbing Mesoporous Silica-Coated Copper Sulphide Nanostructures for Light-to-Thermal Energy Conversion. *Nanomaterials* **2022**, *12*, 2545. [[CrossRef](#)]
61. Avila-Rodriguez, M.A.; Rios, C.; Carrasco-Hernandez, J.; Manrique-Arias, J.C.; Martinez-Hernandez, R.; Garcia-Perez, F.O.; Martinez-Rodriguez, E.; Romero-Piña, M.E.; Diaz-Ruiz, A. Biodistribution and radiation dosimetry of [(64)Cu]copper dichloride: First-in-human study in healthy volunteers. *EJNMMI Res.* **2017**, *7*, 98. [[CrossRef](#)] [[PubMed](#)]
62. Patel, D.; Kell, A.; Simard, B.; Xiang, B.; Lin, H.Y.; Tian, G. The cell labeling efficacy, cytotoxicity and relaxivity of copper-activated MRI/PET imaging contrast agents. *Biomaterials* **2011**, *32*, 1167–1176. [[CrossRef](#)] [[PubMed](#)]
63. Zhao, Y.; Sultan, D.; Detering, L.; Cho, S.; Sun, G.; Pierce, R.; Wooley, K.L.; Liu, Y. Copper-64-alloyed gold nanoparticles for cancer imaging: Improved radiolabel stability and diagnostic accuracy. *Angew. Chem. Int. Ed. Engl.* **2014**, *53*, 156–159. [[CrossRef](#)]
64. Zhou, M.; Tian, M.; Li, C. Copper-Based Nanomaterials for Cancer Imaging and Therapy. *Bioconjug. Chem.* **2016**, *27*, 1188–1199. [[CrossRef](#)]
65. Fulda, S.; Debatin, K.M. Extrinsic versus intrinsic apoptosis pathways in anticancer chemotherapy. *Oncogene* **2006**, *25*, 4798–4811. [[CrossRef](#)] [[PubMed](#)]
66. Konarikova, K.; Perdikaris, G.A.; Gbelcova, H.; Andrezalova, L.; Sveda, M.; Ruml, T.; Laubertová, L.; Žitňanová, I. Effect of Schiff base Cu(II) complexes on signaling pathways in HT-29 cells. *Mol. Med. Rep.* **2016**, *14*, 4436–4444. [[CrossRef](#)] [[PubMed](#)]
67. Langer, V.; Gyepesová, D.; Scholtzová, E.; Mach, P.; Kohútová, M.; Valent, A.; Smrčok, L.U. Crystal and electronic structure of aqua (N-salicylidene-methylester-L-glutamato) Cu (II) monohydrate. *Z. Kristallogr. Cryst. Mater.* **2004**, *219*, 112–116. [[CrossRef](#)]
68. Nakao, Y.; Sakurai, K.I.; Nakahara, A. Copper (II) chelates of Schiff bases derived from salicylaldehyde and various α -amino acids. *Bull. Chem. Soc. Jpn.* **1967**, *40*, 1536–1538. [[CrossRef](#)]
69. Krätzmár-Šmogrovič, J.; Pavelčík, F.; Soldánová, J.; Sivy, J.; Seressová, V.; Žemlička, M. The Crystal and Molecular Structure and Properties of Diaqua [N-salicylidene-(S)-(+)–glutamato] copper (II) Monohydrate. *Z. Nat. B* **1991**, *46*, 1323–1327. [[CrossRef](#)]
70. Zhang, J.; Duan, D.; Xu, J.; Fang, J. Redox-Dependent Copper Carrier Promotes Cellular Copper Uptake and Oxidative Stress-Mediated Apoptosis of Cancer Cells. *ACS Appl. Mater. Interfaces* **2018**, *10*, 33010–33021. [[CrossRef](#)]
71. Kunos, C.A.; Andrews, S.J.; Moore, K.N.; Chon, H.S.; Ivy, S.P. Randomized Phase II Trial of Triapine-Cisplatin-Radiotherapy for Locally Advanced Stage Uterine Cervix or Vaginal Cancers. *Front. Oncol.* **2019**, *9*, 1067. [[CrossRef](#)] [[PubMed](#)]
72. Vutey, V.; Castelli, S.; D’Annessa, I.; Samia, L.B.; Souza-Fagundes, E.M.; Beraldo, H.; Desideria, A. Human topoisomerase IB is a target of a thiosemicarbazone copper(II) complex. *Arch. Biochem. Biophys.* **2016**, *606*, 34–40. [[CrossRef](#)]
73. Kaur, P.; Johnson, A.; Northcote-Smith, J.; Lu, C.; Suntharalingam, K. Immunogenic Cell Death of Breast Cancer Stem Cells Induced by an Endoplasmic Reticulum-Targeting Copper(II) Complex. *Chembiochem* **2020**, *21*, 3618–3624. [[CrossRef](#)]
74. Tardito, S.; Isella, C.; Medico, E.; Marchio, L.; Bevilacqua, E.; Hatzoglou, M.; Bussolati, O.; Franchi-Gazzola, R. The thioxotriazole copper(II) complex A0 induces endoplasmic reticulum stress and paraptotic death in human cancer cells. *J. Biol. Chem.* **2009**, *284*, 24306–24319. [[CrossRef](#)] [[PubMed](#)]
75. Passeri, G.; Northcote-Smith, J.; Suntharalingam, K. Delivery of an immunogenic cell death-inducing copper complex to cancer stem cells using polymeric nanoparticles. *RSC Adv.* **2022**, *12*, 5290–5299. [[CrossRef](#)] [[PubMed](#)]
76. Li, D.D.; Yague, E.; Wang, L.Y.; Dai, L.L.; Yang, Z.B.; Zhi, S.; Zhang, N.; Zhao, X.-M.; Hu, Y.-H. Novel Copper Complexes That Inhibit the Proteasome and Trigger Apoptosis in Triple-Negative Breast Cancer Cells. *ACS Med. Chem. Lett.* **2019**, *10*, 1328–1335. [[CrossRef](#)] [[PubMed](#)]
77. Yu, Z.; Fenk, K.D.; Huang, D.; Sen, S.; Cowan, J.A. Rapid Telomere Reduction in Cancer Cells Induced by G-Quadruplex-Targeting Copper Complexes. *J. Med. Chem.* **2019**, *62*, 5040–5048. [[CrossRef](#)] [[PubMed](#)]
78. Deka, B.; Sarkar, T.; Banerjee, S.; Kumar, A.; Mukherjee, S.; Deka, S.; Saikia, K.K.; Hussain, A. Novel mitochondria targeted copper(ii) complexes of ferrocenyl terpyridine and anticancer active 8-hydroxyquinolines showing remarkable cytotoxicity, DNA and protein binding affinity. *Dalton Trans.* **2017**, *46*, 396–409. [[CrossRef](#)]
79. Weissleder, R. Molecular imaging in cancer. *Science* **2006**, *312*, 1168–1171. [[CrossRef](#)]
80. Louie, A. Multimodality imaging probes: Design and challenges. *Chem. Rev.* **2010**, *110*, 3146–3195. [[CrossRef](#)]
81. Ku, G.; Chen, J.; Vittal, J.J. Copper sulfide nanoparticles as a new class of photoacoustic contrast agent for deep tissue imaging at 1064 nm. *ACS Nano.* **2012**, *6*, 7489–7496. [[CrossRef](#)] [[PubMed](#)]
82. Zhang, H.F.; Maslov, K.; Stoica, G.; Wang, L.V. Functional photoacoustic microscopy for high-resolution and noninvasive in vivo imaging. *Nat. Biotechnol.* **2006**, *24*, 848–851. [[CrossRef](#)] [[PubMed](#)]
83. Siphanto, R.; Thumma, K.K.; Kolkman, R.G.M.; van Leeuwen, T.G.; de Mul, F.F.M.; van Neck, J.W.; van Adrichem, L.N.A.; Steenbergen, W. Serial noninvasive photoacoustic imaging of neovascularization in tumor angiogenesis. *Opt. Express* **2005**, *13*, 89–95. [[CrossRef](#)] [[PubMed](#)]
84. Yao, J.; Wang, L.V. Photoacoustic tomography: Fundamentals, advances and prospects. *Contrast Media Mol. Imaging* **2011**, *6*, 332–345. [[CrossRef](#)]

85. Zha, Z.; Deng, Z.; Li, Y.; Li, C.; Wang, J.; Wang, S.; Quc, E.; Dai, Z. Biocompatible polypyrrole nanoparticles as a novel organic photoacoustic contrast agent for deep tissue imaging. *Nanoscale* **2013**, *5*, 4462–4467. [[CrossRef](#)] [[PubMed](#)]
86. Bao, B.; Yang, Z.; Liu, Y.; Xu, Y.; Gu, B.; Chen, J.; Su, P.; Tong, L.; Wang, L. Two-photon semiconducting polymer nanoparticles as a new platform for imaging of intracellular pH variation. *Biosens. Bioelectron.* **2019**, *126*, 129–135. [[CrossRef](#)] [[PubMed](#)]
87. Manohar, S.; Ungureanu, C.; van Leeuwen, T.G. Gold nanorods as molecular contrast agents in photoacoustic imaging: The promises and the caveats. *Contrast Media Mol. Imaging* **2011**, *6*, 389–400. [[CrossRef](#)]
88. Cherukula, K.; Manickavasagam Lekshmi, K.; Uthaman, S.; Cho, K.; Cho, C.S.; Park, I.K. Multifunctional inorganic nanoparticles: Recent progress in thermal therapy and imaging. *Nanomaterials* **2016**, *6*, 76. [[CrossRef](#)]
89. Zhou, M.; Ku, G.; Pigeon, L.; Li, C. Theranostic probe for simultaneous in vivo photoacoustic imaging and confined photothermolysis by pulsed laser at 1064 nm in 4T1 breast cancer model. *Nanoscale* **2014**, *6*, 15228–15235. [[CrossRef](#)]
90. Ding, K.; Zeng, J.; Jing, L.; Qiao, R.; Liu, C.; Jiao, M.; Libc, Z.; Gao, M. Aqueous synthesis of PEGylated copper sulfide nanoparticles for photoacoustic imaging of tumors. *Nanoscale* **2015**, *7*, 11075–11081. [[CrossRef](#)]
91. Wang, S.; Zhang, L.; Zhao, J.; He, M.; Huang, Y.; Zhao, S. A tumor microenvironment-induced absorption red-shifted polymer nanoparticle for simultaneously activated photoacoustic imaging and photothermal therapy. *Sci. Adv.* **2021**, *7*, eabe3588. [[CrossRef](#)] [[PubMed](#)]
92. Bindra, A.K.; Wang, D.; Zheng, Z.; Jana, D.; Zhou, W.; Yan, S.; Wuac, H.; Zheng, Y.; Zhao, Y. Self-assembled semiconducting polymer based hybrid nanoagents for synergistic tumor treatment. *Biomaterials* **2021**, *279*, 121188. [[CrossRef](#)] [[PubMed](#)]
93. Phelps, M.E. Positron emission tomography provides molecular imaging of biological processes. *PNAS* **2000**, *97*, 9226–9233. [[CrossRef](#)]
94. Zhou, M.; Zhang, R.; Huang, M.; Lu, W.; Song, S.; Melancon, M.P.; Tian, M.; Liang, D.; Li, C. A chelator-free multifunctional [64Cu] CuS nanoparticle platform for simultaneous micro-PET/CT imaging and photothermal ablation therapy. *J. Am. Chem. Soc.* **2010**, *132*, 15351–15358. [[CrossRef](#)] [[PubMed](#)]
95. Suárez-García, S.; Esposito, T.V.; Neufeld-Peters, J.; Bergamo, M.; Yang, H.; Saatchi, K.; Schaffer, P.; Häfeli, U.O.; Ruiz-Molina, D.; Rodríguez-Rodríguez, C. Hybrid Metal–Phenol Nanoparticles with Polydopamine-like Coating for PET/SPECT/CT Imaging. *ACS Appl. Mater. Interfaces* **2021**, *13*, 10705–10718. [[CrossRef](#)]
96. Zhang, H.; Song, F.; Dong, C.; Yu, L.; Chang, C.; Chen, Y. Co-delivery of nanoparticle and molecular drug by hollow mesoporous organosilica for tumor-activated and photothermal-augmented chemotherapy of breast cancer. *J. Nanobiotechnology* **2021**, *19*, 290. [[CrossRef](#)]
97. Chen, C.; Ma, Y.; Du, S.; Wu, Y.; Shen, P.; Yan, T.; Li, X.; Song, Y.; Zha, Z.; Han, X. Controlled CRISPR-Cas9 Ribonucleoprotein Delivery for Sensitized Photothermal Therapy. *Small* **2021**, *17*, 2101155. [[CrossRef](#)]
98. Druzhkova, I.N.; Shirmanova, M.V.; Kuznetsova, D.S.; Lukina, M.M.; Zagaynova, E.V. Modern Approaches to Testing Drug Sensitivity of Patients' Tumors (Review). *Sovrem Tekhnologii Med.* **2021**, *12*, 91–102. [[CrossRef](#)]
99. Ganesan, K.; Wang, Y.; Gao, F.; Liu, Q.; Zhang, C.; Li, P.; Zhang, L.; Chen, J. Targeting Engineered Nanoparticles for Breast Cancer Therapy. *Pharmaceutics* **2021**, *13*, 1829. [[CrossRef](#)]
100. Es-Haghi, A.; Taghavizadeh, Y.M.; Sharifalhosseini, M.; Baghani, M.; Yousefi, E.; Rahdar, A.; Baino, F. Application of Response Surface Methodology for Optimizing the Therapeutic Activity of ZnO Nanoparticles Biosynthesized from *Aspergillus niger*. *Biomimetics* **2021**, *6*, 34. [[CrossRef](#)]
101. Sanaei, M.J.; Pourbagheri-Sigaroodi, A.; Kaveh, V.; Sheikholeslami, S.A.; Salari, S.; Bashash, D. The application of nano-medicine to overcome the challenges related to immune checkpoint blockades in cancer immunotherapy: Recent advances and opportunities. *Crit. Rev. Oncol. Hematol.* **2021**, *157*, 103160. [[CrossRef](#)] [[PubMed](#)]
102. Park, W.; Heo, Y.J.; Han, D.K. New opportunities for nanoparticles in cancer immunotherapy. *Biomater. Res.* **2018**, *22*, 24. [[CrossRef](#)] [[PubMed](#)]
103. Ahamed, M.; Akhtar, M.J.; Alhadlaq, H.A.; Alshamsan, A. Copper ferrite nanoparticle-induced cytotoxicity and oxidative stress in human breast cancer MCF-7 cells. *Colloids Surf. B. Biointerfaces* **2016**, *142*, 46–54. [[CrossRef](#)]
104. Rajagopal, G.; Nivetha, A.; Sundar, M.; Panneerselvam, T.; Murugesan, S.; Parasuraman, P.; Kumar, S.; Ilango, S.; Kunjiappan, S. Mixed phytochemicals mediated synthesis of copper nanoparticles for anticancer and larvicidal applications. *Heliyon* **2021**, *7*, e7360. [[CrossRef](#)] [[PubMed](#)]
105. Wang, S.; Liu, J.; Qiu, S.; Yu, J. Facile fabrication of Cu9-S5 loaded core-shell nanoparticles for near infrared radiation mediated tumor therapeutic strategy in human esophageal squamous carcinoma cells nursing care of esophageal cancer patients. *J. Photochem. Photobiol. B.* **2019**, *199*, 111583. [[CrossRef](#)] [[PubMed](#)]
106. Xu, R.; Zhang, K.; Liang, J.; Gao, F.; Li, J.; Guan, F. Hyaluronic acid/polyethyleneimine nanoparticles loaded with copper ion and disulfiram for esophageal cancer. *Carbohydr. Polym.* **2021**, *261*, 117846. [[CrossRef](#)]
107. Imyanitov, E.N.; Iyevleva, A.G.; Levchenko, E.V. Molecular testing and targeted therapy for non-small cell lung cancer: Current status and perspectives. *Crit. Rev. Oncol. Hematol.* **2021**, *157*, 103194. [[CrossRef](#)]
108. Herbst, R.S.; Morgensztern, D.; Boshoff, C. The biology and management of non-small cell lung cancer. *Nature* **2018**, *553*, 446–454. [[CrossRef](#)]
109. Naatz, H.; Manshian, B.B.; Rios, L.C.; Tsikourkitoudi, V.; Deligiannakis, Y.; Birkenstock, J.; Pokhrel, S.; Mädler, L. Model-Based Nanoengineered Pharmacokinetics of Iron-Doped Copper Oxide for Nanomedical Applications. *Angew. Chem. Int. Ed. Engl.* **2020**, *59*, 1828–1836. [[CrossRef](#)]

110. Kalaiarasi, A.; Sankar, R.; Anusha, C.; Saravanan, K.; Aarthy, K.; Karthic, S.; Ravikumar, V. Copper oxide nanoparticles induce anticancer activity in A549 lung cancer cells by inhibition of histone deacetylase. *Biotechnol. Lett.* **2018**, *40*, 249–256. [[CrossRef](#)]
111. Giri, R.K.; Chaki, S.; Khimani, A.J.; Vaidya, Y.H.; Thakor, P.; Thakkar, A.B.; Pandya, S.J.; Deshpande, M.P. Biocompatible CuInS₂ Nanoparticles as Potential Antimicrobial, Antioxidant, and Cytotoxic Agents. *ACS Omega* **2021**, *6*, 26533–26544. [[CrossRef](#)] [[PubMed](#)]
112. Li, W.B.; Stangl, S.; Klapproth, A.; Shevtsov, M.; Hernandez, A.; Kimm, M.A.; Schuemann, J.; Qiu, R.; Michalke, B.; Bernal, M.A.; et al. Application of High-Z Gold Nanoparticles in Targeted Cancer Radiotherapy-Pharmacokinetic Modeling, Monte Carlo Simulation and Radiobiological Effect Modeling. *Cancers* **2021**, *13*, 5370. [[CrossRef](#)] [[PubMed](#)]
113. Klapproth, A.P.; Shevtsov, M.; Stangl, S.; Li, W.B.; Multhoff, G. A New Pharmacokinetic Model Describing the Biodistribution of Intravenously and Intratumorally Administered Superparamagnetic Iron Oxide Nanoparticles (SPIONs) in a GL261 Xenograft Glioblastoma Model. *Int. J. Nanomed.* **2020**, *15*, 4677–4689. [[CrossRef](#)] [[PubMed](#)]
114. Wang, L.; Hu, C.; Shao, L. The antimicrobial activity of nanoparticles: Present situation and prospects for the future. *Int. J. Nanomed.* **2017**, *12*, 1227–1249. [[CrossRef](#)] [[PubMed](#)]
115. Zheng, R.; Cheng, Y.; Qi, F.; Wu, Y.; Han, X.; Yan, J.; Zhang, H. Biodegradable Copper-Based Nanoparticles Augmented Chemodynamic Therapy through Deep Penetration and Suppressing Antioxidant Activity in Tumors. *Adv. Healthc. Mater.* **2021**, *10*, e2100412. [[CrossRef](#)]
116. Koh, J.Y.; Lee, S.J. Metallothionein-3 as a multifunctional player in the control of cellular processes and diseases. *Mol. Brain* **2020**, *13*, 116. [[CrossRef](#)]
117. Lelievre, P.; Sancey, L.; Coll, J.L.; Deniaud, A.; Busser, B. The Multifaceted Roles of Copper in Cancer: A Trace Metal Element with Dysregulated Metabolism, but Also a Target or a Bullet for Therapy. *Cancers* **2020**, *12*, 3594. [[CrossRef](#)]
118. Camats, M.; Pla, D.; Gomez, M. Copper nanocatalysts applied in coupling reactions: A mechanistic insight. *Nanoscale* **2021**, *13*, 18817–18838. [[CrossRef](#)]
119. Mehdizadeh, T.; Zamani, A.; Abtahi, F.S. Preparation of Cu nanoparticles fixed on cellulosic walnut shell material and investigation of its antibacterial, antioxidant and anticancer effects. *Heliyon* **2020**, *6*, e3528. [[CrossRef](#)]
120. Naikoo, G.; Al-Mashali, F.; Arshad, F.; Al-Maashani, N.; Hassan, I.U.; Al-Baraami, Z.; Faruck, L.H.; Qurashi, A.; Ahmed, W.; Asiri, A.M. An Overview of Copper Nanoparticles: Synthesis, Characterisation and Anticancer Activity. *Curr. Pharm. Des.* **2021**, *27*, 4416–4432. [[CrossRef](#)]
121. Akter, M.; Sikder, M.T.; Rahman, M.M.; Ullah, A.; Hossain, K.; Banik, S.; Hosokawa, T.; Saito, T.; Kurasaki, M. A systematic review on silver nanoparticles-induced cytotoxicity: Physicochemical properties and perspectives. *J. Adv. Res.* **2018**, *9*, 1–16. [[CrossRef](#)] [[PubMed](#)]
122. Da, S.D.; De Luca, A.; Squitti, R.; Rongioletti, M.; Rossi, L.; Machado, C.; Cerchiaro, G. Copper in tumors and the use of copper-based compounds in cancer treatment. *J. Inorg. Biochem.* **2022**, *226*, 111634. [[CrossRef](#)]
123. Fahmy, H.M.; Ebrahim, N.M.; Gaber, M.H. In-vitro evaluation of copper/copper oxide nanoparticles cytotoxicity and genotoxicity in normal and cancer lung cell lines. *J. Trace. Elem. Med. Biol.* **2020**, *60*, 126481. [[CrossRef](#)] [[PubMed](#)]
124. Prasad, P.R.; Kanchi, S.; Naidoo, E.B. In-vitro evaluation of copper nanoparticles cytotoxicity on prostate cancer cell lines and their antioxidant, sensing and catalytic activity: One-pot green approach. *J. Photochem. Photobiol. B Biol.* **2016**, *161*, 375–382. [[CrossRef](#)] [[PubMed](#)]
125. Dong, C.; Feng, W.; Xu, W.; Yu, L.; Xiang, H.; Chen, Y.; Zhou, J. The Copper Age: Copper (Cu)-Involved Nanotheranostics. *Adv. Sci.* **2020**, *21*, 2001549. [[CrossRef](#)]
126. Li, W.; Zamani, R.; Rivera Gil, P.; Pelaz, B.; Ibáñez, M.; Cadavid, D.; Shavel, A.; Alvarez-Puebla, R.A.; Parak, W.J.; Arbiol, J.; et al. CuTe Nanocrystals: Shape and Size Control, Plasmonic Properties, and Use as SERS Probes and Photothermal Agents. *J. Am. Chem. Soc.* **2013**, *135*, 7098–7101. [[CrossRef](#)] [[PubMed](#)]
127. Tian, Q.; Jiang, F.; Zou, R.; Liu, Q.; Chen, Z.; Zhu, M.; Yang, S.; Wang, J.; Wang, J.; Hu, J. Hydrophilic Cu₉S₅ Nanocrystals: A Photothermal Agent with a 25.7% Heat Conversion Efficiency for Photothermal Ablation of Cancer Cells in Vivo. *ACS Nano* **2011**, *5*, 9761–9771. [[CrossRef](#)]
128. Tian, Q.; Tang, M.; Sun, Y.; Zou, R.J.; Chen, Z.G.; Zhu, M.F.; Yang, S.P.; Wang, J.L.; Wang, J.H.; Hu, J.Q. Hydrophilic Flower-Like CuS Superstructures as an Efficient 980 nm Laser-Driven Photothermal Agent for Ablation of Cancer Cells. *Adv. Mater.* **2011**, *23*, 3542–3547. [[CrossRef](#)]
129. Liu, X.; Li, B.; Fu, F.; Xu, K.; Zou, R.; Wang, Q.; Zhang, B.; Chena, Z.; Hu, J. Facile synthesis of biocompatible cysteine-coated CuS nanoparticles with high photothermal conversion efficiency for cancer therapy. *Dalton Trans.* **2014**, *43*, 11709–11715. [[CrossRef](#)]
130. Wang, Z.; Tang, X.; Wang, X.; Yang, D.; Yang, C.; Lou, Y.; Chen, J.; He, N. Near-infrared light-induced dissociation of zeolitic imidazole framework-8 (ZIF-8) with encapsulated CuS nanoparticles and their application as a therapeutic nanoplatform. *Chem. Comm.* **2016**, *52*, 12210–12213. [[CrossRef](#)]
131. Zhang, S.; Huang, Q.; Zhang, L.; Zhang, H.; Han, Y.; Sun, Q.; Cheng, Z.; Qin, H.; Doub, S.; Li, Z. Vacancy engineering of Cu_{2-x}Se nanoparticles with tunable LSPR and magnetism for dual-modal imaging guided photothermal therapy of cancer. *Nanoscale* **2018**, *10*, 3130–3143. [[CrossRef](#)] [[PubMed](#)]
132. Dong, L.; Ji, G.; Liu, Y.; Xu, X.; Lei, P.; Du, K.; Song, S.; Feng, J.; Zhang, H. Multifunctional Cu–Ag₂S nanoparticles with high photothermal conversion efficiency for photoacoustic imaging-guided photothermal therapy in vivo. *Nanoscale* **2018**, *10*, 825–831. [[CrossRef](#)] [[PubMed](#)]

133. Ji, M.; Xu, M.; Zhang, W.; Yang, Z.; Huang, L.; Liu, J.; Zhang, Y.; Gu, L.; Yu, Y.; Hao, W.; et al. Structurally Well-Defined Au@Cu₂-xS Core-Shell Nanocrystals for Improved Cancer Treatment Based on Enhanced Photothermal Efficiency. *Adv. Mater.* **2016**, *28*, 3094–3101. [[CrossRef](#)] [[PubMed](#)]
134. Ding, X.; Liow, C.H.; Zhang, M.; Huang, R.; Li, C.; Shen, H.; Liu, M.; Zou, Y.; Gao, N.; Zhang, Z.; et al. Surface Plasmon Resonance Enhanced Light Absorption and Photothermal Therapy in the Second Near-Infrared Window. *J. Am. Chem. Soc.* **2014**, *136*, 15684–15693. [[CrossRef](#)]
135. Zhu, H.; Wang, Y.; Chen, C.; Ma, M.; Zeng, J.; Li, S.; Xia, Y.; Gao, M. Monodisperse Dual Plasmonic Au@Cu₂-xE (E= S, Se) Core@Shell Supraparticles: Aqueous Fabrication, Multimodal Imaging, and Tumor Therapy at in Vivo Level. *ACS Nano.* **2017**, *11*, 8273–8281. [[CrossRef](#)]
136. Chen, W.; Qin, M.; Chen, X.; Wang, Q.; Zhang, Z.; Sun, X. Combining photothermal therapy and immunotherapy against melanoma by polydopamine-coated Al₂O₃ nanoparticles. *Theranostics* **2018**, *8*, 2229–2241. [[CrossRef](#)]
137. Wu, Z.-C.; Li, W.-P.; Luo, C.-H.; Su, C.H.; Yeh, C.S. Rattle-Type Fe₃O₄@CuS Developed to Conduct Magnetically Guided Photoinduced Hyperthermia at First and Second NIR Biological Windows. *Adv. Funct. Mater.* **2015**, *25*, 6527–6537. [[CrossRef](#)]
138. Webb, B.A.; Chimenti, M.; Jacobson, M.P.; Barber, D.L. Dysregulated pH: A perfect storm for cancer progression. *Nat. Rev. Cancer* **2011**, *11*, 671–677. [[CrossRef](#)]
139. Estrela, J.M.; Ortega, A.; Obrador, E. Glutathione in Cancer Biology and Therapy. *Crit. Rev. Clin. Lab. Sci.* **2006**, *43*, 143–181. [[CrossRef](#)]
140. Harris, A.L. Hypoxia—a key regulatory factor in tumour growth. *Nat. Rev. Cancer* **2002**, *2*, 38–47. [[CrossRef](#)]
141. An, L.; Wang, X.; Rui, X.; Lin, J.; Yang, H.; Tian, Q.; Tao, C.; Yang, S. The In Situ Sulfidation of Cu₂O by Endogenous H₂S for Colon Cancer Theranostics. *Angew. Chem. Int. Ed* **2018**, *57*, 15782–15786. [[CrossRef](#)] [[PubMed](#)]
142. Tao, C.; An, L.; Lin, J.; Tian, Q.; Yang, S. Surface Plasmon Resonance-Enhanced Photoacoustic Imaging and Photothermal Therapy of Endogenous H₂S-Triggered Au@Cu₂O. *Small* **2019**, *15*, 1903473. [[CrossRef](#)] [[PubMed](#)]
143. Cheng, Y.; Chen, Q.; Guo, Z.; Li, M.; Yang, X.; Wan, G.; Chen, H.; Zhang, Q.; Wang, Y. An Intelligent Biomimetic Nanoplatform for Holistic Treatment of Metastatic Triple-Negative Breast Cancer via Photothermal Ablation and Immune Remodeling. *ACS Nano.* **2020**, *14*, 15161–15181. [[CrossRef](#)] [[PubMed](#)]