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# Bundle-sheath leakiness and intrinsic water use efficiency of a perennial $C_4$ grass are increased at high vapour pressure deficit during growth

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# Abstract

Bundle-sheath leakiness ( $\phi$ ) is a key parameter of the CO<sub>2</sub>-concentrating mechanism of C<sub>4</sub> photosynthesis and is related to leaf-level intrinsic water use efficiency (WUE<sub>i</sub>). This work studied short-term dynamic responses of  $\phi$  to alterations of atmospheric CO<sub>2</sub> concentration in *Cleistogenes squarrosa*, a perennial grass, grown at high (1.6 kPa) or low (0.6 kPa) vapour pressure deficit (VPD) combined with high or low N supply in controlled environment experiments.  $\phi$  was determined by concurrent measurements of photosynthetic gas exchange and on-line carbon isotope discrimination, using a new protocol. Growth at high VPD led to an increase of  $\phi$  by 0.13 and a concurrent increase of WUE<sub>i</sub> by 14%, with similar effects at both N levels.  $\phi$  responded dynamically to intercellular CO<sub>2</sub> concentration (C<sub>i</sub>), increasing with C<sub>i</sub>. Across treatments,  $\phi$  was negatively correlated to the ratio of CO<sub>2</sub> saturated assimilation rate to carboxylation efficiency (a proxy of the relative activities of Rubisco and phosphoenolpyruvate carboxylase) indicating that the long-term environmental effect on  $\phi$  was related to the balance between C<sub>3</sub> and C<sub>4</sub> cycles. Our study revealed considerable dynamic and long-term variation in  $\phi$  of *C. squarrosa*, suggesting that  $\phi$  should be determined when carbon isotope discrimination is used to assess WUE<sub>i</sub>. Also, the data indicate a trade-off between WUE<sub>i</sub> and energetic efficiency in *C. squarrosa*.

**Key words:** C<sub>4</sub> photosynthesis, CO<sub>2</sub>-concentrating mechanism, carbon isotope discrimination, gas exchange, nitrogen nutrition, vapour pressure deficit.

# Introduction

The CO<sub>2</sub> concentrating mechanism (CCM) is a specialized feature of C<sub>4</sub> photosynthesis and enables the maintenance of a very high CO<sub>2</sub> partial pressure at the site of ribulose-1,5-bi-sphosphate carboxylase/oxygenase (Rubisco), thus effectively minimizing photorespiration (Hatch, 1987). The CCM underlies the higher quantum yield, and higher water- and nitrogen-use efficiency of C<sub>4</sub> plants relative to C<sub>3</sub> plants under high temperature and/or low intercellular CO<sub>2</sub> (Ehleringer *et al.*,

1997; Long, 1999). In C<sub>4</sub> photosynthesis, CO<sub>2</sub> is first fixed by phosphoenolpyruvate carboxylase (PEPc) in mesophyll cells to form C<sub>4</sub> acids (the C<sub>4</sub> cycle), which are transported into bundle-sheath cells, where the acids are decarboxylated and CO<sub>2</sub> is finally fixed by Rubisco (the C<sub>3</sub> cycle). Effective CCM requires a strong 'CO<sub>2</sub> pump' (a high rate of the C<sub>4</sub> cycle). However, leakage of CO<sub>2</sub> from bundle-sheath cells back to mesophyll cells represents an energetic inefficiency that is

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related to the cost of the regeneration of phosphoenolpyruvate (PEP; Hatch, 1987; Furbank *et al.*, 1990). The ratio of this leakage rate to the C<sub>4</sub> cycle rate is termed bundle-sheath leakiness (Farquhar, 1983), or leakiness ( $\phi$ ), and is a key parameter of C<sub>4</sub> photosynthesis.

Leakiness is generally quantified using a carbon isotope approach based on the model of carbon isotope discrimination ( $\Delta$ ) of C<sub>4</sub> photosynthesis (Farquhar, 1983). The simplified version of the C<sub>4</sub> discrimination model indicates that  $\Delta$ is determined by  $\phi$  and C<sub>i</sub>/C<sub>a</sub>, the ratio of intercellular CO<sub>2</sub> concentration (C<sub>i</sub>) to atmospheric CO<sub>2</sub> concentration (C<sub>a</sub>):

$$\Delta = a + (b_4 + \phi(b_3 - s) - a)C_i / C_a,$$
(1)

with *a* the fractionation during diffusion of CO<sub>2</sub> in air,  $b_4$  the combined fractionation of PEP carboxylation and the preceding fractionation associated with dissolution of CO<sub>2</sub> and conversion to HCO<sub>3</sub><sup>-</sup>,  $b_3$  the fractionation by Rubisco and *s* the fractionation during leakage of inorganic C out of the bundle sheath. Since leaf-level intrinsic water use efficiency (WUE<sub>i</sub>; net CO<sub>2</sub> assimilation rate/stomatal conductance) is a function of  $C_i$  and  $C_a$ , combined with Eqn 1 we have:

WUE<sub>i</sub> = 
$$\frac{C_a}{1.6} \left( 1 - \frac{\Delta - a}{b_4 + (b_3 - s)\phi - a} \right)$$
, (2)

showing that WUE<sub>i</sub> can be estimated from  $\Delta$  if  $\phi$  is known. Similar to the application of  $\Delta$  to quantify WUE<sub>i</sub> of C<sub>3</sub> plants (Farquhar and Richards, 1984; Yang *et al.*, 2016*b*),  $\Delta$  was suggested as a potentially promising screening tool for breeding and improvement of  $C_4$  crops (Henderson *et al.*, 1998; Gresset et al., 2014; von Caemmerer et al., 2014). However, such endeavours require knowledge of the magnitude and variability of  $\phi$  and relevant environmental drivers. Early studies led to the notion that leakiness is relatively constant within a species with moderate short-term environmental variation, e.g. in response to light,  $CO_2$ , and temperature (Henderson *et al.*, 1992). A constant leakiness would largely simplify the relationship between  $WUE_i$  and  $\Delta$ . However, more recent studies suggest dynamic variation of leakiness in response to shortterm variation of irradiation (Kromdijk et al., 2010; Pengelly et al., 2010; Ubierna et al., 2013; Bellasio and Griffiths, 2014) or temperature (von Caemmerer et al., 2014).

Analysis of dynamic changes of leakiness in response to short-term variation of atmospheric CO<sub>2</sub> provides an opportunity for studying the functioning of the CCM. However, model analyses suggested an increase of leakiness with increasing  $C_i$  (von Caemmerer and Furbank, 1999; Kiirats *et al.*, 2002; Ghannoum, 2009; Yin *et al.*, 2011), while (the few) experimental studies using the carbon isotope approach showed no clear response of leakiness to  $C_i$  (Henderson *et al.*, 1992; Pengelly *et al.*, 2012).

Long-term effects of environmental drivers on leakiness have been observed. Thus, drought stress was shown to increase leakiness of several species (Saliendra *et al.*, 1996; Williams *et al.*, 2001; Fravolini *et al.*, 2002), and this effect was attributed to the relative activities of Rubisco/ PEPc (Saliendra *et al.*, 1996). N stress was reported to affect leakiness of several sugarcane species (Meinzer and Zhu, 1998), but another study with different species found no effect of N fertilizer supply (Fravolini *et al.*, 2002). Increased CO<sub>2</sub> concentration was shown to increase (Watling *et al.*, 2000; Fravolini *et al.*, 2002) or have no effect (Williams *et al.*, 2001) on leakiness. Currently, it is unknown if vapour pressure deficit (VPD) during plant growth has an effect on leakiness. Studies on long-term effects of N nutrition, water supply, or CO<sub>2</sub> concentration during growth on leakiness using biomass-based  $\Delta$  have considerable uncertainty related to post-photosynthetic fractionation and the temporal mismatch between biomass formation and gas exchange measurements as discussed by many authors (Henderson *et al.*, 1992; Cousins *et al.*, 2008; von Caemmerer *et al.*, 2014).

Scarcity of experimental evidence is partially due to the fact that quantifying leakiness using combined measurements of gas exchange and  $\Delta$  ('on-line'  $\Delta$ ) is a laborious task. Measurement artefacts, i.e. CO<sub>2</sub> leak fluxes between the leaf cuvette and the surrounding air and isotopic disequilibria between photosynthetic and respiratory CO<sub>2</sub> fluxes, can lead to errors of measured  $\Delta$  of several permil (Gong *et al.*, 2015). This technical issue is particularly relevant for C<sub>4</sub> species, as the  $\Delta$  of C<sub>4</sub> plants is generally much lower than that of C<sub>3</sub> plants (Farquhar, 1983).

A previous study in our lab demonstrated that *Cleistogenes* squarrosa (Trin.) Keng-a perennial C<sub>4</sub> (NAD-ME) grass and co-dominant species in the semiarid steppe of Inner Mongolia—had a large and variable biomass-based  $\Delta$  of leaves (6–9‰; Yang et al., 2011), potentially indicating a high and variable  $\phi$ . As  $\phi$  is related to WUE<sub>i</sub> (Eqn 2) and potentially driven by environmental factors, we were interested to unravel the mechanisms governing  $\Delta$  in this C<sub>4</sub> species. Furthermore, understanding the physiological response of C. squarrosa to N nutrition and VPD may provide new insight into the adaptation of this species to its habitat given that: (i) water and N availability are the major limiting factors that determine primary production and resource use efficiency of plants in the semiarid steppes of Inner Monglia (Gong et al., 2011) and (ii) the abundance of  $C_4$  plants has increased substantially during the past several decades in the Mongolian plateau (Wittmer et al., 2010). Given the lack of knowledge on long-term effects of VPD or N nutrition and short-term dynamic effects of  $CO_2$  on  $\phi$ , we performed controlled environment experiments with C. squarrosa grown under a low or high level of VPD combined with a low or high level of N fertilizer supply. Leakiness was determined using combined measurements of gas exchange and  $\Delta$  (on-line  $\Delta$ ) following the protocols of Gong et al. (2015).

## Materials and methods

Plant material and growth conditions

*Cleistogenes squarrosa* is an NAD-ME type (determined by <sup>14</sup>C labelling) C<sub>4</sub> grass, with a 'chloridoid' type of Kranz anatomy (Pyankov *et al.*, 2000). Stands of *C. squarrosa* were grown from seed in plastic pots (5 cm diameter, 35 cm depth) filled with quartz sand. Pots were placed in growth chambers (PGR15, Conviron, Winnipeg, Canada) at a density of 236 plants m<sup>-2</sup>. A photosynthetic photon

flux density (PPFD) of 800 µmol m<sup>-2</sup> s<sup>-1</sup>, provided by cool white fluorescent tubes, was maintained at the top of the canopies during the 16 h-long photoperiods. Air temperature was maintained at 25 °C during photo- and dark periods. A CO<sub>2</sub> concentration ([CO<sub>2</sub>]) of 386±3 µmol mol<sup>-1</sup> (mean±SD, *n*>400) in chamber air was maintained during the photoperiods (cf. Schnyder *et al.*, 2003). A modified Hoagland nutrient solution was supplied three times per day by an automatic irrigation system throughout the entire experiment similar to Lehmeier *et al.* (2008). The time of first watering is referred to as imbibition of seeds. Before germination, all chambers were run with a VPD of 0.63 kPa (corresponding to a relative humidity (RH) of 80%).

#### N nutrition and VPD treatments

The study had a  $2 \times 2$  factorial design, with N fertilizer supply and VPD as factors, two levels (low and high, see below) for each factor, and four replicates (four replicate stands) for each combination of N and VPD level. About 1 week after the imbibition of seeds (i.e. first watering), VPD and N treatment were implemented until the end of the experiment. The nutrient solution with 7.5 mM N in the form of equimolar concentrations of calcium nitrate and potassium nitrate was supplied to chambers of the low N treatment (N1), while another nutrient solution with 22.5 mM N was supplied to chambers of the high N treatment (N2). The concentration of other nutrients was the same in both nutrient solutions: 1.0 mM MgSO<sub>4</sub>, 0.5 mM KH<sub>2</sub>PO<sub>4</sub>, 1 mM NaCl, 125 µM Fe-EDTA, 46 µM H<sub>3</sub>BO<sub>3</sub>, 9 µM MnSO<sub>4</sub>, 1 µM ZnSO<sub>4</sub>, 0.3 µM CuSO<sub>4</sub>, 0.1 µM Na<sub>2</sub>MoO<sub>4</sub>. VPD (difference between actual vapour pressure and the saturation vapour pressure) was maintained at 0.63 kPa (corresponding to an RH of 80%) in the chambers of the low VPD treatment (V1) or at 1.58 kPa (corresponding to an RH of 50%) in the chambers of the high VPD treatment (V2). Each chamber (or each stand) was assigned to one of the treatments: N1V1, N1V2, N2V1, and N2V2. As we had a total of four chambers, in order to repeat each treatment four times, all replicates were accommodated in four sequential experimental runs (in total, 16 stands were grown) (cf. Table S1 in Liu et al. 2016). In each treatment, two replicate chambers were supplied with CO<sub>2</sub> from a mineral source ( $\delta^{13}C_{CO_2}$  of -6%) and the other two chambers with CO<sub>2</sub> from an organic source ( $\delta^{13}C_{CO_2}$  of -33%). Carbon isotope discrimination by the canopies led to some  $^{13}$ C-enrichment of the CO<sub>2</sub> inside the growth chambers. However, that effect was small due to a high rate of air flow through the chambers: quasi-continuous on-line measurements of  $\delta^{13}C_{CO_2}$  inside the growth chambers demonstrated a  $\delta^{13}C_{CO_2}$  inside the chambers with the <sup>13</sup>C-enriched CO<sub>2</sub> source of  $-5.2\pm0.1\%$  (mean $\pm$ SD, n=23) and  $-31.7\pm0.1\%$  (*n*=23) inside the chambers with the <sup>13</sup>C-depleted CO<sub>2</sub>.

#### Gas exchange measurement system

The method reported by Gong et al. (2015) was used for leaf-level combined measurements of gas exchange and <sup>13</sup>C discrimination (on-line  $\Delta$ ). Measurements were performed using a leaf-level  $^{13}\text{CO}_2/^{12}\text{CO}_2$  gas exchange and labelling system, which included a portable CO<sub>2</sub> exchange system (LI-6400, LI-COR Inc., Lincoln, NE, USA) housed in a gas exchange mesocosm (Gong et al., 2015). The air supply to the mesocosm and LI-6400 was mixed from CO<sub>2</sub>-free, dry air, and CO<sub>2</sub> of know  $\delta^{13}C_{CO_2}$  (cf. Schnyder *et al.*, 2003). [CO<sub>2</sub>] inside the mesocosm (growth chamber) was monitored with an infrared gas analyser (LI-6262, LI-COR Inc.). During the measurement, the plant to be measured and the sensor head of the LI-6400 were placed inside the mesocosm, with concentration and  $\delta^{13}C$  of CO<sub>2</sub> in both gas exchange facilities monitored and controlled. Using this set-up, we separately controlled the concentration and  $\delta^{13}$ C in both facilities (sensor head and growth chamber) for purposes of gas exchange measurement or <sup>13</sup>C labelling (see below).

The mesocosm and leaf cuvette systems were coupled to a continuous-flow isotope ratio mass spectrometer (IRMS; Delta<sup>plus</sup> Advantage equipped with GasBench II, ThermoFinnigan, Bremen, Germany) via a stainless steel capillary. Sample air was drawn through the capillary via a peristaltic pump and passed through a 0.25 ml sample loop attached to the 8-port Valco valve of the GasBench II. Sample air in the loop was introduced into the IRMS via an open split after passage of a dryer (Nafion) and a GC column (25 m×0.32 mm Poraplot Q; Chrompack, Middleburg, The Netherlands). After every second sample a VPDB-gauged CO<sub>2</sub> reference gas was injected into the IRMS via the Open Split. The whole-system precision (SD) of repeated measurements was 0.10‰ (n=50). For further details of the method see Gong *et al.* (2015).

# Gas exchange and concurrent on-line <sup>13</sup>C discrimination measurements

Gas exchange and on-line  $\Delta$  measurements followed the optimized protocols suggested by Gong *et al.* (2015). In order to minimize the effects of any CO<sub>2</sub> leak artefact across the gaskets of the clamp-on leaf cuvette and the surrounding (mesocosm) air, [CO<sub>2</sub>] inside the mesocosm was maintained close to that of the leaf cuvette of the LI-6400 (with the difference <30 µmol mol<sup>-1</sup>), and the same CO<sub>2</sub> source was supplied to the mesocosm and LI-6400. By these means, the diffusive gradient of <sup>13</sup>CO<sub>2</sub> or <sup>12</sup>CO<sub>2</sub> between leaf cuvette and the surrounding air was effectively minimized (Gong *et al.*, 2015). In order to also minimize the isotopic disequilibrium artefact between the photosynthetic and respiratory CO<sub>2</sub> flux, the same CO<sub>2</sub> source was used for growing plants and for on-line leaf-level  $\Delta$  measurements (Gong *et al.*, 2015).

Gas exchange and on-line  $\Delta$  measurements were performed on the fully expanded young leaves near the tip of major tillers (the second youngest fully expanded leaf), started at about 38 days and ended at about 53 days after the imbibition of seeds. We used the standard 2 × 3 cm cuvette of LI-6400. Two leaf blades from separate major tillers of the same plant were carefully placed in the cuvette. This was done to compensate for the narrow leaf width of *C. squarrosa* (about 3 mm). The enclosed leaf area was determined after gas exchange measurements by photographing the leaf blades and analysis with ImageJ software (National Institutes of Health, Bethesda, MD, USA). All gas exchange parameters were recomputed using the measured enclosed leaf area.

Photosynthetic CO<sub>2</sub> response curves were taken with the successive CO<sub>2</sub> steps of 390, 800, 390, 260, 180, 120, 90, 60, and 30 µmol  $mol^{-1}$  ([CO<sub>2</sub>] in the cuvette) on at least two plants randomly chosen from each growth chamber. In total, 42 plants were measured. The data of five plants were eliminated from further analysis due to extremely low leaf N contents. Finally, each treatment had nine to ten replicates of individual plants. During the gas exchange measurements, the environmental parameters were maintained close to the growth conditions: leaf temperature was maintained at 25 °C, PPFD at 800  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>, leaf-to-air VPD at 0.76–0.96 kPa (V1 treatment) or 1.4–1.6 kPa (V2 treatment; Supplementary Fig. S1 at JXB online) at all CO<sub>2</sub> levels. To generate the appropriate VPD for gas exchange measurements, the inflow air stream (well-mixed dry air) was passed through a gas washing bottle placed in a temperature-controlled water bath. For each step of the CO<sub>2</sub> response, the [CO<sub>2</sub>] in the mesocosm was also adjusted to be close to that of the cuvette. Leaves were acclimated for at least 30 min before the start of gas exchange measurements. After  $CO_2$  exchange rates reached a steady state at each step of the CO2 response, gas exchange parameters (measured by LI-6400), e.g. net  $CO_2$  assimilation rate (A), transpiration rate (E), and stomatal conductance to vapour  $(g_s)$  were recorded. Instantaneous water use efficiency was calculated as  $WUE_{ins} = A/E$ ; intrinsic water use efficiency was calculated as  $WUE_i = A/g_s$ .

Among the 37 CO<sub>2</sub> response curves, 22 plants were concurrently measured for on-line <sup>13</sup>C discrimination. For this, the [CO<sub>2</sub>] and  $\delta^{13}$ C of the incoming ( $C_{in}$  and  $\delta_{in}$ ) and outgoing cuvette air ( $C_{out}$  and  $\delta_{out}$ ) were measured on five to six plants per treatment combination. All measurements of  $\delta^{13}$ C were corrected for the non-linearity effect of the IRMS (Elsig and Leuenberger, 2010).

<sup>13</sup>C discrimination during net CO<sub>2</sub> assimilation ( $\Delta$ ) was calculated according to Evans *et al.* (1986):

$$\Delta = \frac{\xi(\delta_{out} - \delta_{in})}{1 + \delta_{out} - \xi(\delta_{out} - \delta_{in})},\tag{3}$$

where  $\xi = C_{\rm in}/(C_{\rm in} - C_{\rm out})$ .

#### Determination of leaf respiration rate in light

Additional measurements were performed to determine leaf respiration rate in light ( $R_L$ ). This was done with leaves of the same age class as used for CO<sub>2</sub> response measurements using the <sup>13</sup>C labelling method reported by Gong *et al.* (2015). All measurements were performed under growth conditions (i.e. at  $C_a$  of 390 µmol mol<sup>-1</sup>, PPFD of 800 µmol m<sup>-2</sup> s<sup>-1</sup>). Briefly,  $\Delta$  of each leaf was measured first with the same CO<sub>2</sub> source as used for growing plants, then the same measurement was repeated with the other CO<sub>2</sub> source (i.e. the <sup>13</sup>C-enriched or -depleted CO<sub>2</sub> source). Making use of isotopic disequilibria between photosynthetic and respiratory CO<sub>2</sub> fluxes derived from the two sets of on-line  $\Delta$  measurements, the ratio of  $R_L$  to net CO<sub>2</sub> assimilation rate (A) was solved as:

$$R_{\rm L} / A = \left(\delta_{\rm AE} - \delta_{\rm AD}\right) / \left(\delta_{\rm outE} - \delta_{\rm outD}\right) - 1, \tag{4}$$

$$\delta_{\rm A} = \left(\delta_{\rm in} C_{\rm in} - \delta_{\rm out} C_{\rm out}\right) / \left(C_{\rm in} - C_{\rm out}\right),\tag{5}$$

where  $\delta_A$  and  $\delta_{out}$  are the  $\delta^{13}$ C of net CO<sub>2</sub> assimilation and that of CO<sub>2</sub> in the cuvette, respectively, and subscripts E and D indicate the relevant parameters measured with the <sup>13</sup>C-enriched or the <sup>13</sup>C-depleted sources of CO<sub>2</sub>, respectively.  $R_L$  was measured on six individuals in each treatment.  $R_L$  of other leaves of the same treatment was estimated by multiplying A of that leaf by the treatmentspecific mean of  $R_L/A$ . For further details of the methodology see Gong *et al.* (2015).

#### Bundle-sheath leakiness ( $\phi$ ), $C_3$ cycle rate and $C_4$ cycle rate

Bundle-sheath leakiness was determined using the complete version of the Farquhar  $C_4$  discrimination model (Farquhar, 1983), modified to include the ternary correction (Farquhar and Cernusak, 2012):

$$\Delta = \frac{1}{1-t} \left[ a_{\rm b} \frac{C_{\rm a} - C_{\rm L}}{C_{\rm a}} + a \frac{C_{\rm L} - C_{\rm i}}{C_{\rm a}} \right] + \frac{1+t}{1-t} \left[ \left( e_{\rm s} + a_{\rm l} \right) \frac{C_{\rm i} - C_{\rm m}}{C_{\rm a}} + \frac{b_{\rm 4} + \phi \left( \frac{b_{\rm 3}C_{\rm bs}}{C_{\rm bs} - C_{\rm m}} - s \right)}{1 + \frac{\phi C_{\rm m}}{C_{\rm bs} - C_{\rm m}}} \frac{C_{\rm m}}{C_{\rm a}} \right],$$
(6)

where  $C_a$ ,  $C_L$ ,  $C_i$ ,  $C_m$ , and  $C_{bs}$  are the CO<sub>2</sub> concentrations in air ( $C_a = C_{out}$  during gas exchange measurements), at the leaf surface, in the intercellular airspace, in the mesophyll cell, and in the bundle-sheath cell, respectively.  $C_L$  was calculated as  $C_L = C_a - 1.37A/g_{BL}$ , with  $g_{BL}$  the boundary layer conductance to vapour calculated by the LI-6400 software.  $a_b = 2.9\%$ , a = 4.4%,  $e_s = 1.1\%$ ,  $a_i = 0.7\%$ , and s = 1.8% denote fractionation constants associated with CO<sub>2</sub> diffusion across boundary layer, across stomata, dissolution in water, diffusion in liquid phase, and leaking out of bundle sheath cells, respectively. t represents the ternary correction factor (Farquhar and Cernusak, 2012):

$$t = (1 + \overline{a}) E / (2g_{\rm sc}), \tag{7}$$

where *E* is the transpiration rate and  $g_{sc}$  is the total conductance to CO<sub>2</sub>:  $g_{sc}=1/(1.6/g_s+1.37/g_{BL})$ , where  $g_s$  is the stomatal conductance

to vapour.  $\bar{a}$  is the weighted fractionation across boundary layer  $(a_b=2.9\%_0)$  and stomata  $(a=4.4\%_0)$ :  $\bar{a}=[a_b(C_a-C_L)+a(C_L-C_i)]/(C_a-C_i)$ .  $b_3$  (<sup>13</sup>C fractionation during carboxylation by Rubisco including respiratory fractionation) and  $b_4$  (<sup>13</sup>C fractionation by CO<sub>2</sub> dissolution, hydration, and PEPc carboxylation including respiratory fractionation) were calculated according to Farquhar (1983):

$$b_3 = b'_3 - eR_L / V_c - 0.5 fV_o / V_c$$
, and (8)

$$b_4 = b'_4 - 0.5eR_{\rm L} \left(1 - \phi\right) / \left(A + 0.5R_{\rm L}\right),\tag{9}$$

where  $b'_3=30\%$ ,  $b'_4=-5.7\%$ , e=-6% (Ghashghaie *et al.*, 2003; Kromdijk *et al.*, 2010), f=11.6% (Ghashghaie *et al.*, 2003; Lanigan *et al.*, 2008), and  $V_c$  is the rate of Rubisco carboxylation and  $V_o$  is the rate of oxygenation. In our experiments, the same source CO<sub>2</sub> was used for growing plants and for online  $\Delta$  measurements; therefore an additional term (cf. *e'* in Ubierna *et al.* 2013) accounting for the discrepancy of  $\delta^{13}C_{CO_2}$  between growth environment and gas exchange measurements was not included. Here, it is assumed that respiration in mesophyll cells ( $R_m$ ) and bundle-sheath cells ( $R_{bs}$ ) both equal 0.5 $R_L$  (von Caemmerer and Furbank, 1999; Pengelly *et al.*, 2010).

To solve  $\phi$  using Eqns 7–9,  $C_{\rm m}$ ,  $C_{\rm bs}$ ,  $V_{\rm o}$ , and  $V_{\rm c}$  must be estimated. Those parameters were determined using the equations of the C<sub>4</sub> photosynthesis model of von Caemmerer and Furbank (1999).  $C_{\rm m}$  can be calculated from:

$$g_{\rm m} = A / (C_{\rm i} - C_{\rm m}), \qquad (10)$$

if mesophyll conductance to CO<sub>2</sub> is known. It has been generally assumed that  $g_m$  is high and non-limiting for CO<sub>2</sub> assimilation in C<sub>4</sub> photosynthesis. Thus, in many published works on  $\phi$ ,  $g_{\rm m}$ was assumed to be infinite, i.e.  $C_i = C_m$ . However, a recent study using a new oxygen isotope approach found that  $g_m$  of  $C_4$  species ranged between 0.66 and 1.8 mol m<sup>-2</sup> s<sup>-1</sup> measured at a leaf temperature of about 32 °C (Barbour et al., 2016). We calculated the minimum  $g_m$  as:  $g_{mmin} = A/C_i$  (von Caemmerer *et al.*, 2014). These results showed that  $g_{mmin}$  decreased with increasing  $C_i$  (see Supplementary Fig. S2). Further, we assumed that the young leaves of C. squarrosa had a  $g_m$  of 0.66 mol m<sup>-2</sup> s<sup>-1</sup> at the growth  $C_{\rm a}$  of 390 µmol mol<sup>-1</sup>. Appling a constant ratio of  $g_{\rm m}/g_{\rm mmin}$  across the CO<sub>2</sub> response curve, we estimated the response of  $g_{\rm m}$  to  $C_{\rm i}$ . The estimates of  $g_{\rm m}$  ranged between 0.2 and 2.4 mol m<sup>-2</sup> s<sup>-1</sup> and decreased with increasing  $C_i$  (Supplementary Fig. S2); similarly, decrease of  $g_m$  with increasing  $C_i$  was found in other  $C_4$  species using the oxygen isotope approach (Osborn et al., 2016) or an 'in vitro V<sub>pmax</sub>' method (N. Ubierna, personal communication).

A simplified version of the C<sub>4</sub> photosynthesis model for enzymelimited CO<sub>2</sub> assimilation rate (von Caemmerer and Furbank, 1999) can be used to model the photosynthetic CO<sub>2</sub> response curve:  $A=\min[(V_p-R_m+g_{bs}C_m), (V_{cmax}-R_L)]$ , where  $V_P$  is the rate of PEP carboxylation,  $V_{cmax}$  is the maximum rate of Rubisco carboxylation, and  $g_{bs}$  is the bundle-sheath conductance to CO<sub>2</sub>. *A* is determined by the term  $V_{cmax}-R_L$  at high CO<sub>2</sub> concentrations. According to this model, we assumed that  $V_{cmax}=A_{max}+R_L$ , where  $A_{max}$  is the maximum net assimilation rate observed in the photosynthetic light response curves measured on leaves of the same age at ambient CO<sub>2</sub> (data shown in Supplementary Fig. S3). Treatment-specific  $V_{cmax}$  of *C. squarrosa* ranged between 16 and 19 µmol m<sup>-2</sup> s<sup>-1</sup>, which is in the range of  $V_{cmax}$  of C<sub>4</sub> grass used in Earth System Models (13–33 µmol m<sup>-2</sup> s<sup>-1</sup>; Rogers, 2014).  $C_{bs}$  was estimated using equations of von Caemmerer and Furbank (1999):

$$C_{\rm bs} = \frac{\gamma^* O_{\rm bs} + K_{\rm c} (1 + \frac{O_{\rm bs}}{K_{\rm o}}) (A + R_{\rm L}) / V_{\rm cmax}}{1 - (A + R_{\rm L}) / V_{\rm cmax}},$$
 (11)

$$O_{\rm bs} = \alpha A / (0.047g_{\rm bs}) + O_{\rm m},$$
 (12)

where  $O_{\rm bs}$  is the bundle-sheath oxygen concentration, and  $O_{\rm m}$ (210 mmol mol<sup>-1</sup>) is the oxygen concentration in mesophyll cells;  $\gamma$ (0.000193) is half of the reciprocal of Rubisco specificity;  $K_c$  and  $K_o$ are the Michaelis–Menten constants of Rubisco for CO<sub>2</sub> (650 µmol mol<sup>-1</sup>) and O<sub>2</sub> (450 µmol mol<sup>-1</sup>) at 25 °C, respectively;  $\alpha$  (0.1) is the fraction of PSII activity in the bundle-sheath;  $g_{bs}$ =0.003 mol m<sup>-2</sup> s<sup>-1</sup>. Furthermore,  $V_o/V_c$  was calculated as  $V_o/V_c=2\gamma^*O_{bs}/C_{bs}$ . Knowing  $V_o/V_c$ ,  $V_o$  and  $V_c$  were calculated using the equation:  $V_c=A+R_L+0.5V_o$ . The values of model parameters and equations were derived from von Caemmerer and Furbank (1999). Knowing  $\phi$ and applying the assumption that  $R_{\rm m}=R_{\rm bs}=0.5R_{\rm L}$  (von Caemmerer and Furbank, 1999; Pengelly *et al.*, 2010), the rate of PEP carboxylation  $(V_p)$  or Rubisco carboxylation  $(V_c)$  was estimated (see Supplementary Fig. S4). As  $C_{\rm m}$  and  $C_{\rm bs}$  cannot be directly measured, the determination of both parameters in this study was associated with some uncertainty. Another commonly used assumption in calculating leakiness using the complete or simplified version of Eqn 6 is that  $CO_2$  is in equilibrium with  $HCO_3^-$  in the mesophyll cytoplasm. The extent of this equilibrium is determined by the relative rate of PEP carboxylation and hydration of  $CO_2(V_h)$ . If  $CO_2$ is not in equilibrium with HCO<sub>3</sub><sup>-</sup>,  $b'_4$  of -5.7% in Eqn 9 should be replaced by  $b'_4$ =-5.7 + 7.9 $V_p/V_h$  (Farquhar 1983). Thus, we calculated  $\phi$  using C<sub>4</sub> discrimination models with different assumptions on  $g_{\rm m}$ ,  $C_{\rm bs}$ , and  $V_{\rm p}/V_{\rm h}$  (Supplementary Fig. S5). These results are compared and discussed below.

#### Fitting of CO<sub>2</sub> response curves

For each plant, measured A was plotted against the respective intercellular  $CO_2$  concentration. A model of  $C_4$  photosynthesis was used to fit the  $CO_2$  response curves (Wang *et al.*, 2012):

$$A = a'(1 - e^{-b'x}) + c'$$
(13)

where x is  $C_i$  and c' is  $R_L$ . Using the fitted parameters (a' and b') of each CO<sub>2</sub> response curve, CO<sub>2</sub>-saturated net assimilation rate ( $A_{sat}$ ) was obtained as (a'+c'), and carboxylation efficiency (CE) was calculated as b'(a'+c') (the initial slope of the  $A-C_i$  curve). For CO<sub>2</sub>

response curves of C<sub>4</sub> leaves,  $A_{sat}$  is proportional to Rubisco activity, while CE is proportional to the PEP carboxylase activity (von Caemmerer and Furbank, 1999; Yin *et al.*, 2011). The ratio of  $A_{sat}$  to CE was calculated to indicate the ratio of Rubisco to PEPc activities.

#### Leaf trait parameters and nitrogen nutrition index

N content in dry mass of leaves previously used for gas exchange measurements was measured using an elemental analyser (NA 1110, Carlo Erba, Milan, Italy). For sample preparation, leaves were dried at 60 °C for 48 h, weighed and then milled. N content on a mass (N<sub>mass</sub>, %DM) and leaf area basis (N<sub>area</sub>, g m<sup>-2</sup>) were calculated. Specific leaf area (SLA, cm<sup>2</sup> mg<sup>-1</sup>) was obtained as SLA=leaf area/leaf dry mass. In each experiment, a set of individual plants (four plants per chamber) were sampled four or five times (twice per week) after canopy closure (when leaf area index was >2) for the determination of standing aboveground biomass, and N content in total aboveground dry mass. Those data were used to calculate nitrogen nutrition index (NNI) according to Lemaire *et al.* (2008).

#### Statistical analysis

Two-way ANOVA was used to test the effects of N supply, VPD, and their interactions on leaf traits and gas exchange parameters, using general linear models of SAS (SAS 9.1, SAS Institute, USA). The effects of N nutrition and VPD on the relationship between  $\ln C_i$  and leakiness were analysed using a dummy regression approach; the parallelism and coincidence of linear regressions of different treatments were tested according to Berenson *et al.* (1983). Non-linear regression analysis on photosynthetic CO<sub>2</sub> response curve of individual plants was performed using the Nlin procedure of SAS.

## Results

#### Leaf traits and photosynthetic CO<sub>2</sub> assimilation

N supply increased  $N_{mass}$  of fully expanded young leaves by 25% (*P*<0.05) and also tended to increase  $N_{area}$  (*P*=0.06), but had no effect on SLA (Table 1). N supply also significantly increased

**Table 1.** Leaf trait and gas exchange parameters of C. squarrosa measured under the same environmental conditions as during growth (leaf temperature 25 °C, PPFD 800  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>, [CO<sub>2</sub>] 390  $\mu$ mol mol<sup>-1</sup>, VPD 0.8 kPa for V1 and 1.6 kPa for V2)

Plants were grown at low or high N fertilizer supply (N1 or N2) combined with low or high VPD (V1 or V2). Leaf trait parameters include N content per unit dry mass (N<sub>mass</sub>, % DM) and per leaf area (N<sub>area</sub>, g m<sup>-2</sup>), and specific leaf area (SLA, cm<sup>2</sup> mg<sup>-1</sup>). Gas exchange parameters: net  $CO_2$  assimilation rate (A, µmol m<sup>-2</sup> s<sup>-1</sup>), transpiration rate (E, mmol m<sup>-2</sup> s<sup>-1</sup>), stomatal conductance to water vapor ( $g_s$ , mol m<sup>-2</sup> s<sup>-1</sup>), respiration rate in light ( $R_L$ , µmol m<sup>-2</sup> s<sup>-1</sup>), the ratio of internal to atmospheric  $CO_2$  concentration ( $C_1/C_a$ ), instantaneous water use efficiency (WUE<sub>ins</sub>= $A/E_s$ , µmol mol<sup>-1</sup>), and intrinsic water use efficiency (WUE<sub>i</sub>= $A/g_s$ , µmol mol<sup>-1</sup>). Data are shown as mean±SE (n=6 for  $R_L$ , n=9-10 for the other parameters). Significant treatment effects (P<0.05) are shown in bold.

	N1		N2		<i>P</i> -value		
	V1	V2	V1	V2	N	VPD	N×VPD
N <sub>mass</sub>	2.4 ± 0.1	2.4 ± 0.1	3.0 ± 0.1	3.0 ± 0.2	0.01	0.81	0.79
N <sub>area</sub>	$1.2 \pm 0.2$	$1.3 \pm 0.2$	$1.6 \pm 0.2$	$1.6 \pm 0.2$	0.06	0.82	0.92
SLA	$0.21 \pm 0.02$	$0.20 \pm 0.02$	$0.21 \pm 0.02$	$0.21 \pm 0.02$	0.91	0.82	0.85
A	$14.8 \pm 0.9$	$13.1 \pm 0.9$	$14.9 \pm 0.9$	$12.3 \pm 1.0$	0.81	0.03	0.58
E	$0.93 \pm 0.08$	$1.14 \pm 0.09$	$0.93 \pm 0.08$	$1.23 \pm 0.09$	0.69	0.01	0.60
$g_{s}$	$0.10 \pm 0.01$	$0.08 \pm 0.01$	$0.11 \pm 0.01$	$0.07 \pm 0.01$	0.69	0.01	0.45
RL	0.81 ± 0.26	$0.62 \pm 0.30$	$0.53 \pm 0.10$	$0.51 \pm 0.10$	0.36	0.62	0.71
$C_i/C_a$	$0.35 \pm 0.03$	$0.26 \pm 0.04$	$0.35 \pm 0.03$	$0.25 \pm 0.04$	0.93	0.01	0.91
WUE <sub>ins</sub>	$15.9 \pm 0.6$	11.7 ± 0.6	$16.8 \pm 0.6$	$10.0 \pm 0.6$	0.82	0.01	0.04
WUEi	$153 \pm 9$	$173 \pm 9$	$154 \pm 9$	178 ± 10	0.79	0.02	0.82

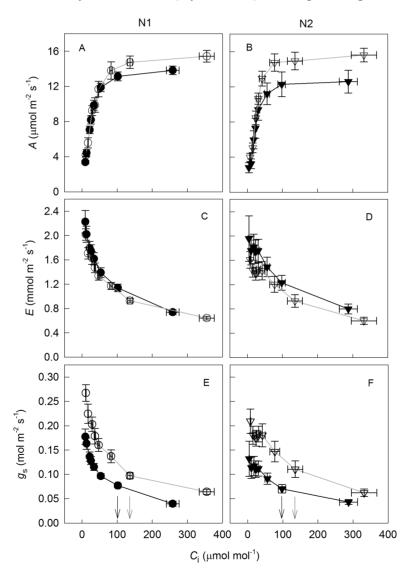
nitrogen nutrition index (NNI; determined from N content and aboveground standing biomass of each stand): NNI was 0.88  $\pm$  0.05 (mean $\pm$ SD, *n*=4 replicate stands) for the N1V1 treatment, 0.71 $\pm$ 0.06 for the N1V2 treatment, 1.25 $\pm$ 0.06 for the N2V1 treatment, and 1.32 $\pm$ 0.05 for the N2V2 treatment. The effects of VPD and the interaction of VPD and N supply on N<sub>mass</sub>, N<sub>area</sub> and SLA were all non-significant (*P*>0.05) (Table 1).

The effects of N supply on N<sub>mass</sub> and N<sub>area</sub> were not associated with parallel effects on gas exchange parameters measured at growth CO<sub>2</sub> level (390 µmol mol<sup>-1</sup>). Thus, N supply had no effect on *A*, *E*,  $g_s$ ,  $C_i/C_a$ , and WUE<sub>ins</sub>, and there was only a small interactive effect of N and VPD on WUE<sub>i</sub> (Table 1). Conversely, VPD during growth had significant effects on all these parameters, but none of these effects demonstrated an interaction with N supply except for WUE<sub>ins</sub>. High VPD decreased *A* by 14%,  $g_s$  by 29%,  $C_i/C_a$  by 27% at both low and high N, and WUE<sub>ins</sub> by 26% at low N and 40% at high N. Conversely, high VPD increased *E* by 27% and WUE<sub>i</sub> by 14% independently of N levels (Table 1). The rate of leaf respiration in light ( $R_L$ ) did not differ between treatments.

The response of photosynthetic gas exchange parameters to dynamic changes of CO<sub>2</sub> concentration (Fig. 1) showed typical CO<sub>2</sub> responses of net CO<sub>2</sub> assimilation rate. VPD had clear effects on gas exchange parameters: compared with low VPD, *A* was 20% lower (Fig. 1A, B), *E* was 21% higher (Fig. 1C, D), and  $g_s$  was 32% lower (Fig. 1E, F) at high VPD averaged over N levels and CO<sub>2</sub> concentrations.

# Carbon isotope discrimination and bundle-sheath leakiness

Carbon isotope discrimination ( $\Delta$ ) during net CO<sub>2</sub> exchange revealed strong dynamic responses to short-term variation of CO<sub>2</sub> concentration;  $\Delta$  increased with increasing C<sub>i</sub> in all treatments, and ranged from 4 to 8‰ (Fig. 2A, B). Compared at the same C<sub>i</sub>,  $\Delta$  was higher at high VPD than at low VPD. Estimates



**Fig. 1.** Net CO<sub>2</sub> assimilation rate (*A*; A, B), transpiration rate (*E*; C, D) and stomatal conductance to water vapour ( $g_s$ ; E, F) in response to short-term variation of intercellular CO<sub>2</sub> (*C*<sub>1</sub>) under low (N1, circles; A, C, E) or high N supply (N2, triangles; B, D, F) combined with low (V1, open symbols) or high VPD (V2, filled symbols). Data are shown as the mean±SE (n=9–10). Operating conditions of gas exchange measurements were the same as conditions in growth chambers (leaf temperature 25 °C, PPFD 800 µmol m<sup>-2</sup> s<sup>-1</sup>, VPD 0.8 kPa for V1 and 1.6 kPa for V2). The corresponding  $C_i$  values at growth CO<sub>2</sub> concentration (390 µmol mol<sup>-1</sup>) are indicated by grey (low VPD) and black arrows (high VPD) in (E) and (F).

of  $\phi$  also increased with increasing C<sub>i</sub> in all treatments, and ranged from 0.23 to 0.81 (Fig. 2C, D). The relationship between  $C_{\rm i}$  and  $\phi$  followed a logarithmic function. Regression analyses between  $\ln C_i$  and  $\phi$  including VPD as a dummy variable indicated a significant VPD effect on  $\phi$ :  $\phi$  was higher by 0.13 at high VPD than at low VPD at both N levels (Fig. 3, Table 2). Plants grown with high N supply had a higher  $\phi$  at moderate to low  $C_i$  ( $C_a \leq 390$ ) than plants with low N supply (Fig. 3). Accordingly, the slope of the linear regression between  $\ln C_i$ and  $\phi$  was significantly lower for the high N treatment than for the low N treatment (Table 2). The increase of  $\phi$  along the gradient of  $C_i$  was apparently related to the discrepancy between rates of  $C_3$  and  $C_4$  cycles:  $V_c$  already reached the maximum at a  $C_i < 100 \,\mu\text{mol mol}^{-1}$ , while  $V_p$  increased throughout the range of measured  $C_i$  (Supplementary Fig. S4). We also calculated  $\phi$  using C<sub>4</sub> discrimination models with different assumptions on  $g_{\rm m}$ ,  $C_{\rm bs}$ , and  $V_{\rm p}/V_{\rm h}$ , and our conclusions on short-term response to CO<sub>2</sub> and long-term response to VPD and N were not altered (see Supplementary Fig. S5).

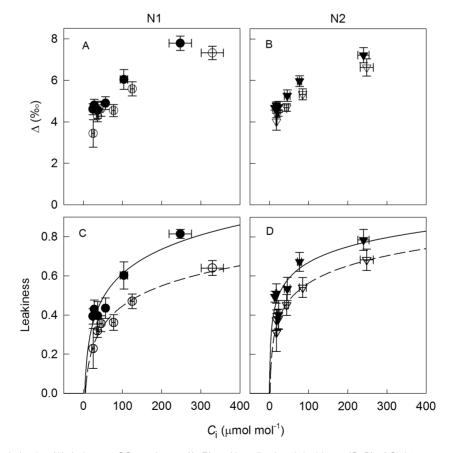
# Relationships between leaf traits and gas exchange parameters

VPD had a clear effect on  $A_{sat}$  (Fig. 4A): averaged over N treatments, high VPD caused a 15% reduction of  $A_{sat}$ .

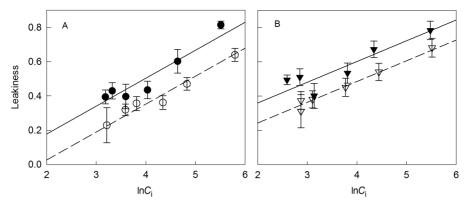
N supply and N×VPD interactions had no effect on  $A_{sat}$ . CE of leaves seemed to be slightly higher at high (0.71 mol  $m^{-2} s^{-1}$ ) than low N (0.53 mol  $m^{-2} s^{-1}$ ), although this effect was only significant at P=0.10 (Fig. 4B). Asat/CE was calculated to indicate the ratio of Rubisco to PEPc activities. Treatment-specific mean Asat/CE was negatively correlated with  $\phi$  (P<0.05, Fig. 4C). Across all treatments, we found no correlation between  $A_{\text{sat}}$  and  $N_{\text{mass}}$  (P=0.41, Fig. 5A), but a positive correlation between CE and  $N_{mass}$  ( $r^2=0.43$ , P < 0.001, Fig. 5B) and a negative correlation between  $A_{sat}$ CE and  $N_{mass}$  ( $r^2=0.29$ , P<0.001, Fig. 5C). WUE<sub>i</sub> was negatively correlated with  $A_{\text{sat}}$  ( $r^2=0.34$ , P<0.001, Fig. 6A) and  $A_{\text{sat}}$ /CE ( $r^2$ =0.49, P<0.001, Fig. 6C), but positively with CE  $(r^2=0.19, P<0.001, Fig. 6B)$  and leakiness  $(r^2=0.19, P<0.05, P<0.05, P<0.05)$ Fig. 6D). Furthermore, a positive correlation between WUE<sub>i</sub> and leakiness was also found during short-term response to  $CO_2$  ( $r^2 > 0.9$ , P < 0.01, Supplementary Fig. S6).

## Discussion

This study indicates an effect of VPD of the growth environment on bundle-sheath leakiness:  $\phi$  was higher at high VPD than at low VPD with an absolute difference of 0.13 at both N supply levels. This effect appeared to be constitutive,



**Fig. 2.** Carbon isotope discrimination ( $\Delta$ ) during net CO<sub>2</sub> exchange (A, B) and bundle sheath leakiness (C, D) of *Cleistogenes squarrosa* leaves in response to short-term variation of intercellular CO<sub>2</sub> (*C*). Plants were grown under low (N1, circles; A, C) or high N supply (N2, triangles; B, D) combined with low (V1, open symbols, dashed lines) or high VPD (V2, filled symbols, solid lines). Operating conditions of gas exchange measurements were the same as conditions in growth chambers (leaf temperature 25 °C, PPFD 800  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>, VPD 0.8 kPa for V1 and 1.6 kPa for V2). Data are shown as the mean±SE (*n*=5–6). The regressions were fitted using a function of *y*=*y*<sub>0</sub>+*a*ln(*x*); all regressions have *r*<sup>2</sup>>0.8.



**Fig. 3.** Relationships between bundle sheath leakiness and  $ln(C_i)$  under low (N1, circles; A) or high N supply (N2, triangles; B) combined with low (V1, open symbols, dashed lines) or high VPD (V2, filled symbols, solid lines). Operating conditions of gas exchange measurements were the same as conditions in growth chambers (leaf temperature 25 °C, PPFD 800  $\mu$ mol m<sup>-2</sup> s<sup>-1</sup>, VPD 0.8 kPa for V1 and 1.6 kPa for V2). Data are shown as the mean $\pm$ SE (*n*=5–6). The data of each N level were fitted by linear regression including a dummy variable indicating VPD treatments (*V*) after the confirmation of parallelism of the two regressions of VPD levels: with *V*=0 for low VPD and *V*=1 for high VPD (for equations see Table 2).

**Table 2.** Results of regression analysis on the relationship between  $C_i$  and leakiness ( $\phi$ )

The data for each N level were fitted by linear regression including a dummy variable indicating VPD treatments (*V*) after the confirmation of parallelism of the two regressions of VPD levels, with *V*=0 for low VPD and *V*=1 for high VPD; 95% confidence intervals of slope and coefficient of dummy variable (*V*) are shown. The coefficient of *V* quantifies the mean difference in  $\phi$  between high and low VPD treatments across *C*<sub>i</sub> levels. Both regressions are significant (*P*<0.001).

N level	equation	r <sup>2</sup>	95% CI slope	95% CI coeff. V
N1	$\phi = -0.30 + 0.16 \ln C_{\rm i} + 0.15 V$	0.94	0.13~0.20	0.09~0.21
N2	$\phi$ =0.001+0.12lnC <sub>i</sub> +0.12V	0.91	0.09~0.15	0.05~0.18

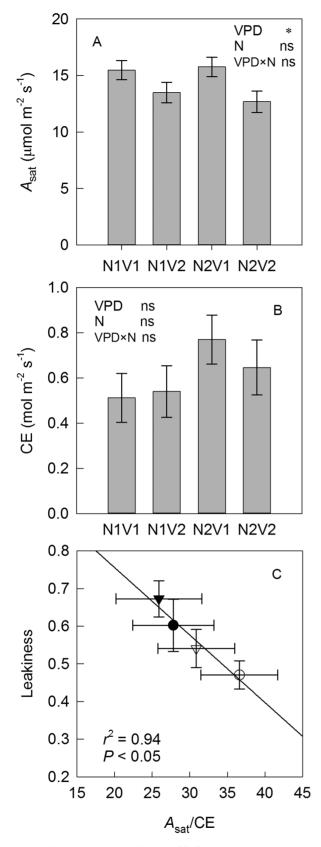
as it was manifest throughout a wide range of short-term variations of CO<sub>2</sub> concentration. Remarkably, the higher leakiness at high VPD was associated with a 14% improved WUE<sub>i</sub>. Also, we observed an N effect on  $\phi$  that responded to short-term variation of CO<sub>2</sub> levels: high N supply increased  $\phi$  of leaves at ambient to low CO<sub>2</sub> levels ( $C_a \leq 390 \,\mu$ mol mol<sup>-1</sup>). Further,  $\phi$  responded dynamically to short-term changes of [CO<sub>2</sub>], in support of theoretical predictions of C<sub>4</sub> photosynthesis models (von Caemmerer and Furbank, 1999; Yin *et al.*, 2011). Lastly, across treatments, a positive correlation between WUE<sub>i</sub> and leakiness was evident, pointing to a trade-off between WUE<sub>i</sub> and energy use efficiency of this species.

Unfortunately, leakiness cannot be measured directly, and the online  $\Delta^{13}$ C approach for estimations of leakiness in conjunction with the C<sub>4</sub> photosynthetic discrimination model of Farquhar (1983) can lead to inaccurate estimates of leakiness if improper simplifications are made (Ubierna *et al.*, 2011, 2013; Gong *et al.*, 2015). One commonly used assumption is that  $g_m$  of C<sub>4</sub> plants is infinite, and thus  $C_i = C_m$ . However, a recent study indicated that  $g_m$  of C<sub>4</sub> plants is finite, although close to the high-end values of  $g_m$  of C<sub>3</sub> plants (Barbour *et al.*, 2016). In our study, assuming a lower range of  $g_m$ across  $C_i$  levels (see Supplementary Fig. S5), or a constant  $g_{\rm m}$  of 1.8 mol m<sup>-2</sup> s<sup>-1</sup> (data not shown) or an infinite  $g_{\rm m}$  (Supplementary Fig. S5) did not modify observed treatment effects. Only if  $g_{\rm m}$  was much lower at low compared with high VPD (e.g.  $g_{\rm m}$  at N1V1 was 60% lower than at N1V2 treatment), would VPD effects on leakiness disappear. However, it is unlikely that *C. squarrosa* had lower  $g_{\rm m}$  at low than at high VPD, as studies with C<sub>3</sub> species generally showed a decrease of  $g_{\rm m}$  by high VPD or water stress (Flexas *et al.*, 2008). Thus, our estimates of treatment effects on leakiness appeared to be largely insensitive to assumptions on  $g_{\rm m}$ .

Many studies using Eqn 6 have assumed that  $CO_2$  is in equilibrium with  $HCO_3^{-}$  in the mesophyll cytoplasm. However, according to Cousins et al. (2008) that assumption might not be proper, especially for C<sub>4</sub> grasses, which have lower carbonic anhydrase (CA) activity than C4 dicots. Our observations with the same species in the same experiment suggested that, under growth conditions, assimilated CO<sub>2</sub> was in equilibrium with HCO<sub>3</sub><sup>-</sup>, as indicated by the fact that  $\delta^{18}$ O of leaf cellulose did not differ between plants growing in the presence of CO<sub>2</sub> with contrasting  $\delta^{18}$ O (Liu *et al.*, 2016). Thus the observed treatment effects on leakiness of C. squarrosa seemed not to be related to CA activity. Further, we calculated leakiness using different assumptions on  $V_p/V_h$ :  $V_p/V_h=0$ or  $V_p/V_h=0.23$  (mean of NAD-ME grasses in Cousins *et al.*, 2008) and observed a minor effect on leakiness (less than 0.06). Thus, the short-term response of leakiness to  $CO_2$  was not explained by changes in  $V_p/V_h$ . Applying another commonly used simplification that  $C_{bs}$  is sufficiently higher than  $C_{\rm m}$ , so that  $C_{\rm bs}/(C_{\rm bs}-C_{\rm m})=1$  and  $C_{\rm m}/(C_{\rm bs}-C_{\rm m})=0$ , also had no influence on our conclusions on effects of environmental factors on leakiness.

# Effects of VPD and N nutrition on bundle-sheath leakiness

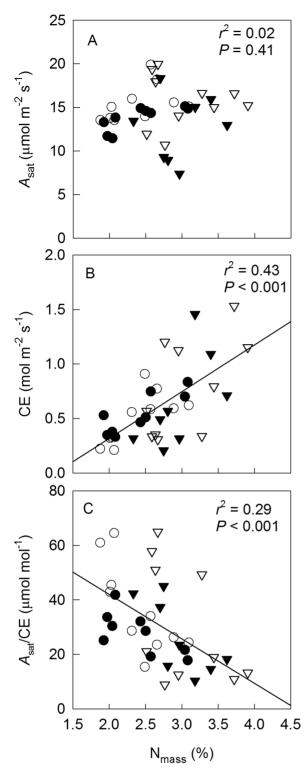
The study demonstrated a higher leakiness of plants grown at high VPD than at low VPD, and this effect was not modified by N nutrition. The present effect was not related to drought stress, as the elongation rate of phytomers (Yang *et al.*, 2016*a*) and the growth rate of individuals (Liu *et al.*, 2016) were not



**Fig. 4.**  $A_{sat}$  (A), carboxylation efficiency (CE; B) and the relationship between leakiness measured under growth conditions (390 µmol mol<sup>-1</sup>) and  $A_{sat}$ /CE (µmol mol<sup>-1</sup>; C) under low or high N fertilizer supply (N1, circles; or N2, triangles) combined with low or high VPD (V1, open symbols; or V2, filled symbols). Data are shown as the mean±SE (n=9–10 for  $A_{sat}$  and CE, n=5–6 for leakiness). \* indicates the treatment effect was significant at the *P*-level of 0.05, while ns indicates no significant effect.

influenced by VPD. The long-term response of leakiness to VPD was related to the decrease of  $C_i/C_a$  and the increase of  $\Delta$  (0.55‰ at ambient CO<sub>2</sub> level) at high VPD, although the direct and indirect influences cannot be clearly distinguished due to the intrinsic correlation between  $C_i/C_a$  and  $\Delta$ . The latter observation was also supported by the <sup>13</sup>C discrimination of leaf dry mass, which was significantly higher at high VPD (5.0±0.2‰, averaged over N levels) than at low VPD treatment  $(4.4\pm0.2\%)$ . Likely, the higher leakiness at high VPD resulted from an enhanced capacity of the C<sub>4</sub> cycle relative to the C<sub>3</sub> cycle at low  $g_s$ , as was supported by the lower  $A_{sat}/CE$ ratio and the lower  $C_i/C_a$  at high VPD relative to low VPD. This interpretation is in line with the findings that the relative activities of Rubisco/PEPc decreased at low  $g_s$  under water stress, with the adjustment achieved by increasing activity of PEPc (Saliendra et al., 1996; Foyer et al., 1998) or decreasing sensitivity of PEPc to the inhibitor malate (Foyer et al., 1998). Also, the data may shed light on the mechanism that underlies the increased leakiness under limited water supply (Saliendra et al., 1996; Williams et al., 2001; Fravolini et al., 2002): that response may have been triggered by decreased stomatal conductance at high VPD. Although a VPD effect on this relationship has not yet been reported, the adjustment of Rubisco/PEPc activities seems to be an important mechanism for plants to counteract the reduced  $g_s$  under longterm low water supply (Saliendra et al., 1996) or high VPD. Maintaining a powerful  $C_4$  pump (at the expense of increased leakiness) might reduce the quantum yield due to the additional ATP requirement for PEP regeneration (Hatch, 1987; Furbank et al., 1990). However, this mechanism may be beneficial for plants to maintain a high WUE; under high VPD or drought stress.

In this work, high N nutrition increased leakiness in shortterm exposures to subambient to ambient [CO<sub>2</sub>], but not to elevated  $[CO_2]$  of 800 µmol mol<sup>-1</sup>. This result is at variance with previous reports of a higher leakiness of plants growing with a low N supply (Meinzer and Zhu, 1998; Fravolini et al., 2002). Other studies also reported non-significant effects of N supply on leakiness (Fravolini et al., 2002; Wang et al., 2012). In the present study, the plants of the low N treatment had a nitrogen nutrition index of ~0.8, which indicated that N was just growth limiting. High N supply led to a NNI of 1.3, and increased plant growth rate via stimulated tiller production (data not shown), but leaf-level net assimilation rates of young leaves were not affected. These results indicate that our low N and high N treatment represented rather 'nearlyadequate' and 'copious supply' of N for leaf-level photosynthesis. High N supply increased leakiness, in correspondence with a lower  $A_{sat}$ /CE ratio, again indicating the involvement of the relative activities of Rubisco/PEPc. This interpretation was further supported by the positive correlation between N<sub>mass</sub> and CE, and simultaneous absence of such a relationship between  $N_{mass}$  and  $A_{sat}$ . These results suggest that under copious N supply to C. squarrosa, the activity of PEPc was more strongly enhanced relative to that of Rubisco, in line with experimental results on Amaranthus retroflexus (Sage et al., 1987). Thus, our results are also in agreement with the hypothesis that C<sub>4</sub> plants have less plasticity in increasing the



**Fig. 5.** Correlations between  $N_{mass}$  and  $A_{sat}$  (A), CE (B) and  $A_{sat}$  (CE (C) of leaves grown at low or high N fertilizer supply (N1, circles; or N2, triangles) combined with low or high VPD (V1, open symbols; or V2, filled symbols). Each symbol represents a data point of an individual plant.

amount of Rubisco compared with  $C_3$  plants, due to spatial restrictions in bundle-sheath cells (Sage and McKown, 2006).

Another potential origin of the variance in leakiness is related to the bundle-sheath conductance  $(g_{bs}=\phi V_P/(C_{bs}-C_m))$ . However, current understanding on  $g_{bs}$  is limited due to the lack of a methodology for quantification of  $g_{bs}$  that is independent of gas exchange measurements. Furthermore, it is unknown how VPD may affect the properties of bundle-sheath cell walls, limiting to some extent discussions of potential mechanisms. A recent study demonstrated that *Miscanthus×giganteus* plants grown at high N supply had higher bundle-sheath area per unit leaf area, which might increase  $g_{bs}$ ; however, estimates of leakiness did not differ between N levels (Ma et al., 2016). Such results highlight the potential complexity of influences on leakiness by biochemical/physiological and anatomical features. In our study, treatments had little effect on individual leaf area and no effect on thickness (Liu et al., 2016), indicating that effects of leaf anatomy may have been minor. Across all treatments, leakiness measured under growth conditions was negatively related to  $A_{sat}$ /CE, again suggesting that variations of leakiness across treatments was related to the balance between C<sub>3</sub> and C<sub>4</sub> cycles.

# Response of bundle-sheath leakiness to short-term variation of $CO_2$

The response of  $\phi$  to short-term variation of [CO<sub>2</sub>] has been predicted by model analyses (von Caemmerer and Furbank, 1999; Kiirats et al., 2002; Ghannoum, 2009; Yin et al., 2011). Although these models were designed for estimating gas exchange rate rather than leakiness, the present results supported the theoretical predictions. So far, only a small number of studies investigated the response of  $\phi$  to dynamic changes of [CO<sub>2</sub>]. A study using the on-line  $\Delta$  method showed no response of  $\phi$  of F. bidentis to dynamic changes of CO<sub>2</sub> (Pengelly et al., 2012), in agreement with the finding of an earlier study on several species (Henderson et al., 1992). However, in the latter study, each leaf was measured at only two or three levels of  $CO_2$ . Those results suggested a rapid regulation of C<sub>3</sub> and C<sub>4</sub> cycle rates in response to changing CO<sub>2</sub> concentrations. The present results indicate that this kind of regulation did not occur to the same extent during short-term CO<sub>2</sub> responses in C. squarrosa. Clearly, the response of  $\phi$  to short-term variation of CO<sub>2</sub> should be studied in detail with a greater number of species.

### Implications on the ecophysiology of C. squarrosa

This work questions the interpretation of leakiness as a simple indicator for efficiency of C<sub>4</sub> photosynthesis or physiological fitness of C<sub>4</sub> plants, as increased leakiness was related to the enhancement of WUE<sub>i</sub> in C. squarrosa. The latter observation was consistent across N and VPD treatments (long-term response) and short-term variation of CO<sub>2</sub> levels. This result was in line with the theoretical prediction of the simplified model of C<sub>4</sub> discrimination: WUE<sub>i</sub> is positively correlated to leakiness when  $\Delta > 4.4\%$ , i.e. when  $\phi > 0.37$ . This relationship indicates a potential trade-off between quantum yield and water use efficiency: high leakiness under VPD stress might decrease the efficiency of energy use, but may be a condition for maintaining the effectiveness of CCM and minimizing photorespiration at a very low  $g_s$  in this species. This mechanism is meaningful for C4 species in arid or semi-arid grasslands, where shading is uncommon but high VPD or drought

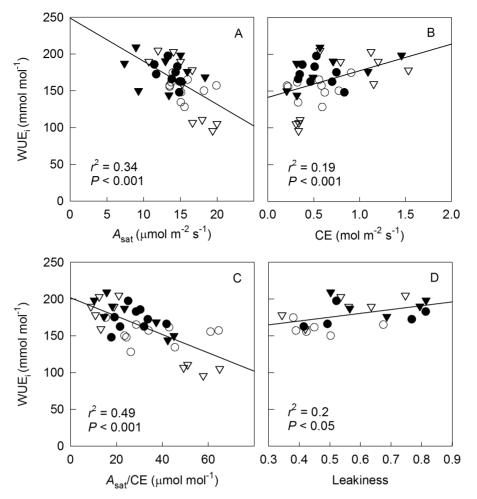


Fig. 6. Correlations between WUE<sub>i</sub> and A<sub>sat</sub> (A), CE (B), A<sub>sat</sub>/CE (C), and leakiness (D) of leaves grown at low or high N fertilizer supply (N1, circles; or N2, triangles) combined with low or high VPD (V1, open symbols; or V2, filled symbols). Each symbol represents a data point of an individual plant.

stress is a common attribute of the habitat. Considering the reduction of Rubisco specificity for  $CO_2$  relative to  $O_2$  by the increase of leaf temperature, this trade-off may be important for  $C_4$  plants in the context of global warming. Observational evidence has indeed shown that this  $C_4$  grass has increased its relative abundance in vast areas of Inner Mongolian grass-lands during regional warming in the last decades (Wittmer *et al.*, 2010).

# Supplementary data

Supplementary data are available at JXB online.

Fig. S1. Relative humidity and leaf-to-air vapour pressure deficit in the leaf cuvette during the measurement of  $CO_2$  response curves.

Fig. S2. Minimum mesophyll conductance to  $CO_2$   $(g_{mmin}=A/C_i)$  and estimated  $g_m$  in response to  $C_i$ .

Fig. S3. Photosynthetic light response curves of leaves of *C. squarrosa*.

Fig. S4.  $V_{\rm p}$  and  $V_{\rm c}$  in response to  $C_{\rm i}$ .

Fig. S5. Bundle-sheath leakiness in response to  $C_i$ , calculated using different models of <sup>13</sup>C discrimination.

Fig. S6. Correlations between  $WUE_i$  and bundle-sheath leakiness of *C. squarrosa*.

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# References

**Barbour MM, Evans JR, Simonin KA, von Caemmerer S.** 2016. Online  $CO_2$  and  $H_2O$  oxygen isotope fractionation allows estimation of mesophyll conductance in  $C_4$  plants, and reveals that mesophyll conductance decreases as leaves age in both  $C_4$  and  $C_3$  plants. New Phytologist **210,** 875–889.

**Bellasio C, Griffiths H.** 2014. Acclimation to low light by  $C_4$  maize: implications for bundle sheath leakiness. Plant, Cell and Environment **37**, 1046–1058.

Berenson ML, Levine DM, Goldstein M. 1983. Intermediate statistical methods and applications: A computer package approach. Englewood Cliffs, NJ: Prentice-Hall.

Cousins AB, Badger MR, von Caemmerer S. 2008. C<sub>4</sub> photosynthetic isotope exchange in NAD-ME- and NADP-ME-type grasses. Journal of Experimental Botany **59**, 1695–1703.

Ehleringer JR, Cerling TE, Helliker BR. 1997.  $C_4$  photosynthesis, atmospheric  $CO_2$ , and climate. Oecologia **112**, 285–299.

Elsig J, Leuenberger MC. 2010. <sup>13</sup>C and <sup>18</sup>O fractionation effects on open splits and on the ion source in continuous flow isotope ratio mass spectrometry. Rapid Communications in Mass Spectrometry **24**, 1419–1430.

**Evans JR, Sharkey TD, Berry JA, Farquhar GD.** 1986. Carbon isotope discrimination measured concurrently with gas-exchange to investigate CO<sub>2</sub> diffusion in leaves of higher plants. Australian Journal of Plant Physiology **13**, 281–292.

**Farquhar GD.** 1983. On the nature of carbon isotope discrimination in  $C_4$  species. Australian Journal of Plant Physiology **10**, 205–226.

Farquhar GD, Cernusak LA. 2012. Ternary effects on the gas exchange of isotopologues of carbon dioxide. Plant, Cell and Environment **35**, 1221–1231.

Farquhar GD, Richards RA. 1984. Isotopic composition of plant carbon correlates with water-use efficiency of wheat genotypes. Australian Journal of Plant Physiology **11**, 539–552.

**Flexas J, Ribas-Carbo M, Diaz-Espejo A, Galmes J, Medrano H.** 2008. Mesophyll conductance to CO<sub>2</sub>: current knowledge and future prospects. Plant, Cell and Environment **31**, 602–621.

**Foyer CH, Valadier MH, Migge A, Becker TW.** 1998. Drought-induced effects on nitrate reductase activity and mRNA and on the coordination of nitrogen and carbon metabolism in maize leaves. Plant Physiology **117**, 283–292.

**Fravolini A, Williams DG, Thompson TL.** 2002. Carbon isotope discrimination and bundle sheath leakiness in three  $C_4$  subtypes grown under variable nitrogen, water and atmospheric  $CO_2$  supply. Journal of Experimental Botany **53**, 2261–2269.

**Furbank RT, Jenkins CLD, Hatch MD.** 1990.  $C_4$  photosynthesis: quantum requirement,  $C_4$  and overcycling and Q-cycle involvement. Australian Journal of Plant Physiology **17**, 1–7.

**Ghannoum O.** 2009. C<sub>4</sub> Photosynthesis and water stress. Annals of Botany **103**, 635–644.

**Ghashghaie J, Badeck F-W, Lanigan G, Nogués S, Tcherkez G, Deléens E, Cornic G, Griffiths H.** 2003. Carbon isotope fractionation during dark respiration and photorespiration in C<sub>3</sub> plants. Phytochemistry Reviews **2**, 145–161.

**Gong XY, Chen Q, Lin S, Brueck H, Dittert K, Taube F, Schnyder H.** 2011. Tradeoffs between nitrogen- and water-use efficiency in dominant species of the semiarid steppe of Inner Mongolia. Plant and Soil **340**, 227–238.

**Gong XY, Schäufele R, Feneis W, Schnyder H.** 2015. <sup>13</sup>CO<sub>2</sub>/<sup>12</sup>CO<sub>2</sub> exchange fluxes in a clamp-on leaf cuvette: disentangling artefacts and flux components. Plant, Cell and Environment **38**, 2417–2432.

**Gresset S, Westermeier P, Rademacher S, Ouzunova M, Presterl T, Westhoff P, Schön CC.** 2014. Stable carbon isotope discrimination is under genetic control in the  $C_4$  species maize with several genomic regions influencing trait expression. Plant Physiology **164,** 131–143.

**Hatch MD.** 1987.  $C_4$  photosynthesis: a unique blend of modified biochemistry, anatomy and ultrastructure. Biochimica et Biophysica Acta **895,** 81–106.

Henderson SA, von Caemmerer S, Farquhar GD. 1992. Short-term measurements of carbon isotope discrimination in several  $C_4$  species. Australian Journal of Plant Physiology **19**, 263–285.

Henderson SA, von Caemmerer S, Farquhar GD, Wade L, Hammer H. 1998. Correlation between carbon isotope discrimination and transpiration efficiency in lines of the  $C_4$  species *Sorghum bicolor* in the glasshouse and the field. Australian Journal of Plant Physiology **25**, 111–123.

**Kiirats O, Lea PJ, Franceschi VR, Edwards GE.** 2002. Bundle sheath diffusive resistance to  $CO_2$  and effectiveness of  $C_4$  photosynthesis and refixation of photorespired  $CO_2$  in a  $C_4$  cycle mutant and wild-type *Amaranthus edulis*. Plant Physiology **130**, 964–976.

**Kromdijk J, Griffiths H, Schepers HE.** 2010. Can the progressive increase of  $C_4$  bundle sheath leakiness at low PFD be explained by incomplete suppression of photorespiration? Plant, Cell and Environment **33**, 1935–1948.

Lanigan GJ, Betson N, Griffiths H, Seibt U. 2008. Carbon isotope fractionation during photorespiration and carboxylation in *Senecio*. Plant Physiology **148**, 2013–2020.

**Lehmeier CA, Lattanzi FA, Schäufele R, Wild M, Schnyder H.** 2008. Root and shoot respiration of perennial ryegrass are supplied by the same substrate pools: assessment by dynamic <sup>13</sup>C labeling and compartmental analysis of tracer kinetics. Plant Physiology **148**, 1148–1158.

Lemaire G, Jeuffroy MH, Gastal F. 2008. Diagnosis tool for plant and crop N status in vegetative stage theory and practices for crop N management. European Journal of Agronomy **28**, 614–624. Liu HT, Gong XY, Schäufele R, Yang F, Hirl RT, Schmidt A, Schnyder

**H.** 2016. Nitrogen fertilization and  $\delta^{18}$ O of CO<sub>2</sub> have no effect on <sup>18</sup>O-enrichment of leaf water and cellulose in *Cleistogenes squarrosa* (C<sub>4</sub>) – is VPD the sole control? Plant, Cell and Environment, DOI: 10.1111/ pce.12824.

**Long SP.** 1999. Environmental responses. In: Sage RF, Monson RK, eds.  $C_4$  plant biology. San Diego: Academic Press, 215–249.

Ma J, Sun W, Koteyeva NK, Voznesenskaya E, Stutz SS, Gandin A, Smith-Moritz AM, Heazlewood JL, Cousins AB. 2016. Influence of light and nitrogen on the photosynthetic efficiency in the C4 plant *Miscanthus* × *giganteus*. Photosynthesis Research, DOI: 10.1007/s11120-016-0281-7.

**Meinzer F, Zhu J.** 1998. Nitrogen stress reduces the efficiency of the  $C_4$  CO<sub>2</sub> concentrating system, and therefore quantum yield, in *Saccharum* (sugarcane) species. Journal of Experimental Botany **49**, 1227–1234.

Osborn HL, Alonso-Cantabrana H, Sharwood RE, Covshoff S, Evans JR, Furbank RT, von Caemmerer S. 2016. Effects of reduced carbonic anhydrase activity on  $CO_2$  assimilation rates in *Setaria viridis*: a transgenic analysis. Journal of Experimental Botany, DOI: 10.1093/jxb/erw357.

**Pengelly JJL, Sirault XRR, Tazoe Y, Evans JR, Furbank RT, von Caemmerer S.** 2010. Growth of the C<sub>4</sub> dicot *Flaveria bidentis*: photosynthetic acclimation to low light through shifts in leaf anatomy and biochemistry. Journal of Experimental Botany **61,** 4109–4122.

**Pengelly JJL, Tan J, Furbank RT, von Caemmerer S.** 2012. Antisense reduction of NADP-malic enzyme in *Flaveria bidentis* reduces flow of  $CO_2$  through the  $C_4$  cycle. Plant Physiology **160,** 1070–1080.

**Pyankov VI, Gunin PD, Tsoog S, Black CC.** 2000. C<sub>4</sub> plants in the vegetation of Mongolia: their natural occurrence and geographical distribution in relation to climate. Oecologia **123**, 15–31.

**Rogers A.** 2014. The use and misuse of  $V_{c,max}$  in Earth System Models. Photosynthesis Research **119**, 15–29.

Sage RF, McKown AD. 2006. Is  $C_4$  photosynthesis less phenotypically plastic than  $C_3$  photosynthesis? Journal of Experimental Botany **57**, 303–317.

**Sage RF, Pearcy RW, Seemann JR.** 1987. The nitrogen use efficiency of  $C_3$  and  $C_4$  Plants III. Leaf nitrogen effects on the activity of carboxylating enzymes in *Chenopodium album* (L) and *Amaranthus retroflexus* (L). Plant Physiology **85**, 355–359.

**Saliendra NZ, Meinzer FC, Perry M, Thom M.** 1996. Associations between partitioning of carboxylase activity and bundle sheath leakiness to CO<sub>2</sub>, carbon isotope discrimination, photosynthesis, and growth in sugarcane. Journal of Experimental Botany **47**, 907–914.

Schnyder H, Schäufele R, Lötscher M, Gebbing T. 2003. Disentangling CO<sub>2</sub> fluxes: direct measurements of mesocosm-scale natural abundance  ${}^{13}CO_2/{}^{12}CO_2$  gas exchange,  ${}^{13}C$  discrimination, and labelling of CO<sub>2</sub> exchange flux components in controlled environments. Plant, Cell and Environment **26**, 1863–1874.

**Ubierna N, Sun W, Cousins AB.** 2011. The efficiency of  $C_4$  photosynthesis under low light conditions: assumptions and calculations with  $CO_2$  isotope discrimination. Journal of Experimental Botany **62**, 3119–3134.

**Ubierna N, Sun W, Kramer DM, Cousins AB.** 2013. The efficiency of  $C_4$  photosynthesis under low light conditions in *Zea mays, Miscanthus* x *giganteus* and *Flaveria bidentis*. Plant, Cell and Environment **36**, 365–381.

von Caemmerer S, Furbank RT. 1999. Modeling  $C_4$  photosynthesis. In: Sage RF, Monson RK, eds.  $C_4$  plant biology. San Diego: Academic Press, 173–211.

von Caemmerer S, Ghannoum O, Pengelly JJL, Cousins AB. 2014. Carbon isotope discrimination as a tool to explore  $C_4$  photosynthesis. Journal of Experimental Botany **65**, 3459–3470.

**Wang ZC, Kang SZ, Jensen CR, Liu FL.** 2012. Alternate partial rootzone irrigation reduces bundle-sheath cell leakage to  $CO_2$  and enhances photosynthetic capacity in maize leaves. Journal of Experimental Botany **63,** 1145–1153.

**Watling JR, Press MC, Quick WP.** 2000. Elevated  $CO_2$  induces biochemical and ultrastructural changes in leaves of the  $C_4$  cereal sorghum. Plant Physiology **123**, 1143–1152.

Williams DG, Gempko V, Fravolini A, Leavitt SW, Wall GW, Kimball BA, Pinter PJ Jr, LaMorte R, Ottman M. 2001. Carbon isotope discrimination by *Sorghum bicolor* under CO<sub>2</sub> enrichment and drought. New Phytologist **150**, 285–293.

Wittmer MHOM, Auerswald K, Bai Y, Schäufele R, Schnyder H. 2010. Changes in the abundance of  $C_3/C_4$  species of Inner Mongolia

grassland: evidence from isotopic composition of soil and vegetation. Global Change Biology **16**, 605–616.

Yang H, Auerswald K, Bai Y, Wittmer MHOM, Schnyder H. 2011. Variation in carbon isotope discrimination in *Cleistogenes squarrosa* (Trin.) Keng: patterns and drivers at tiller, local, catchment, and regional scales. Journal of Experimental Botany **62**, 4143–4152.

Yang F, Gong XY, Liu HT, Schäufele R, Schnyder H. 2016a. Effects of nitrogen and vapour pressure deficit on phytomer growth and development in a  $C_4$  grass. AoB Plants, doi: 10.1093/aobpla/plw075.

Yang Z, Liu J, Tischer SV, Christmann A, Windisch W, Schnyder H, Grill E. 2016b. Leveraging abscisic acid receptors for efficient water use in *Arabidopsis*. Proceedings of the National Academy of Sciences of the United States of America **113**, 6791–6796.

**Yin XY, Sun ZP, Struik PC, Van der Putten PEL, Van leperen W, Harbinson J.** 2011. Using a biochemical C<sub>4</sub> photosynthesis model and combined gas exchange and chlorophyll fluorescence measurements to estimate bundle-sheath conductance of maize leaves differing in age and nitrogen content. Plant, Cell and Environment **34,** 2183–2199.